

# Laboratory of Epigenetic Regulations

RNA degradation, dsRNA, RNAi, miRNA, chromatin

**Petr Svoboda**

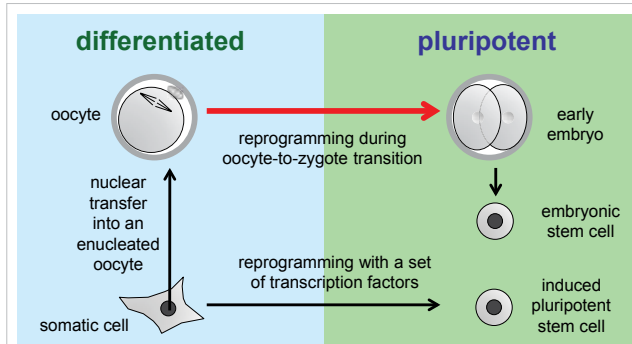
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The zygotic genome activation is the first step in the execution of the genome-encoded program that forms a new organism from a single fertilized cell and it is an essential event in the life of every sexually reproducing organism. The zygotic genome activation is closely associated with formation of pluripotency, i.e. the ability of cells to differentiate into any body cell type. Pluripotency is most studied in two artificial cell types, which maintain pluripotency during *in vitro* culture: embryonic stem cells [ESCs], which are derived from the inner cell mass of the blastocyst, and induced pluripotent stem cells [iPSCs], which form upon reprogramming gene expression in somatic cells with specific pluripotency factors that include transcription factors from the core transcription factor network controlling ESC renewal and pluripotency. A similar network forms in a stepwise manner during the mouse zygotic genome activation, which initiates at the early two-cell stage.

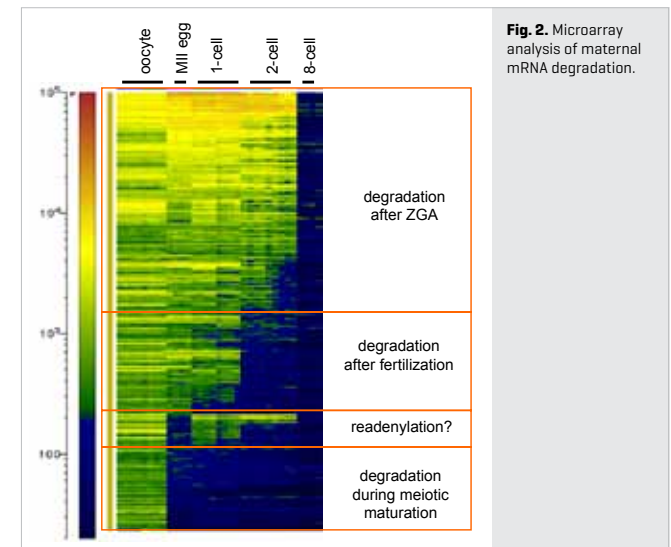
We study reprogramming of oocytes into pluripotent blastomeres of an early embryo [oocyte-to-embryo transition]. This model is the natural parallel to the artificial reprogramming of somatic cells into iPSCs. The oocyte-to-embryo transition, however, is distinct. It is a unidirectional transient process executed by cytoplasmic factors, as demonstrated by animal cloning by nuclear transfer. Our primary research interest is in post-transcriptional mechanisms underlying oocyte-to-embryo transition. These mechanisms include control of maternal mRNA stability, small regulatory RNAs in miRNA and RNAi pathways, and production of maternal transcription factors, which will control gene expression in the embryo. Our goal is to understand how

control of gene expression creates developmental competence *in vivo*.

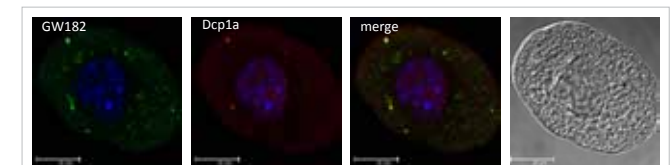
Research of pluripotency is eminent for medicine and biotechnology where pluripotency plays a role in an ever-growing number of applications. Understanding control of the oocyte-to-embryo transition will provide original insights into stem cell biology and will likely contribute to efficient and safe production of pluripotent stem cells, efficient cloning technologies, informative prenatal diagnostics, and understanding of pathology of sterility and developmental defects.



**Fig. 1.** Oocyte-to-zygote transition is a unique model for studying pluripotency. The mammalian oocyte is a highly specialized cell, whose cytoplasm is capable of reprogramming a genome to initiate development of a new organism. The blastomeres of the 2-cell embryo are totipotent as they can give a rise to embryonic and extraembryonic tissues. The pluripotent embryonic stem cells, which have potential to give a rise to any body cell type, are derived from the blastocyst, the final preimplantation embryo stage carrying the first defined cell lineages.



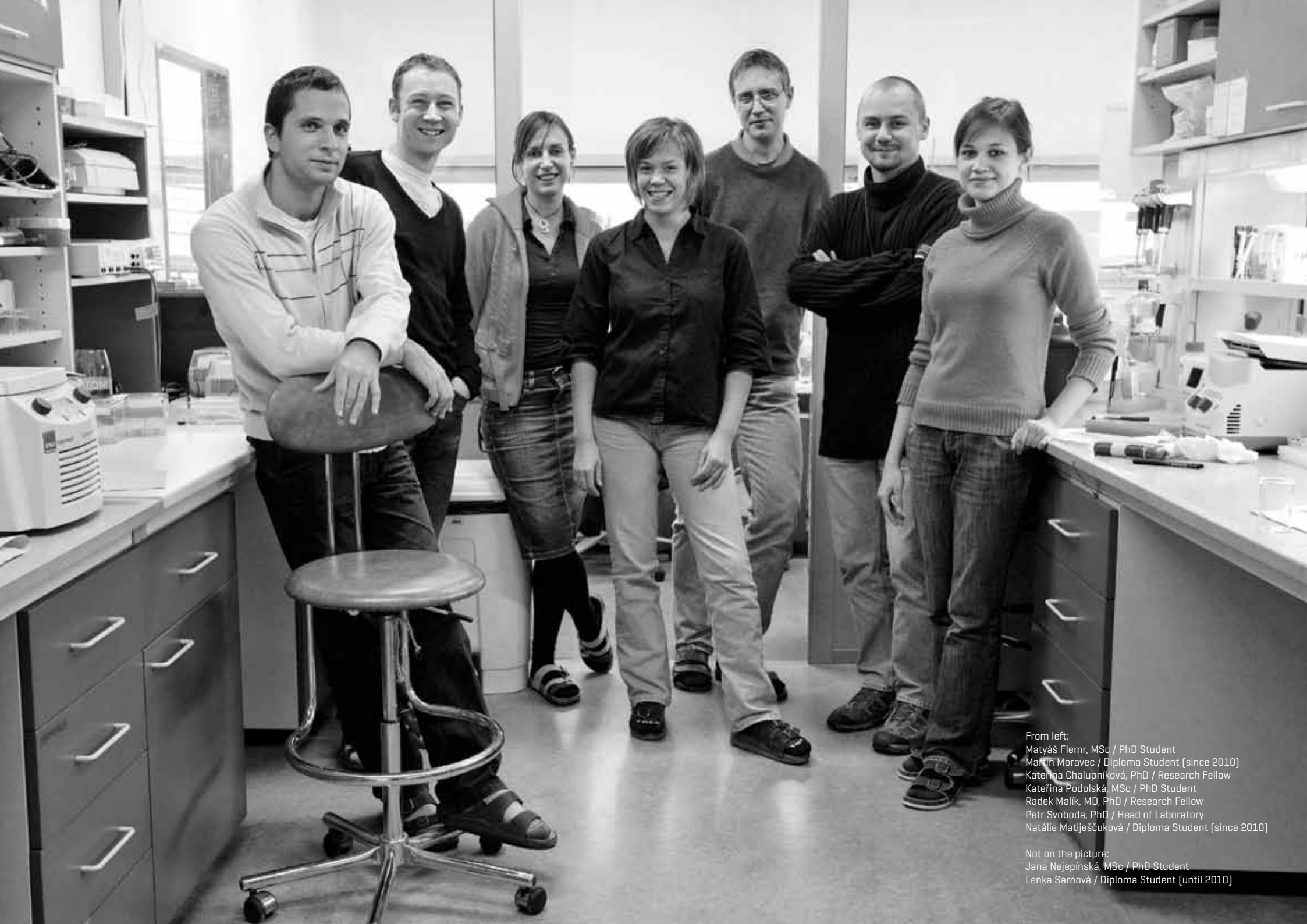
**Fig. 2.** Microarray analysis of maternal mRNA degradation.



**Fig. 3.** Colocalization of p-body components in meiotically incompetent oocytes. P-bodies are centres of mRNA metabolism, including degradation and storage.

- EMBO, 1483 – EMBO Installation Grant, 2007–2011, P. Svoboda
- Ministry of Education, Youth and Sports of the Czech Republic, ME09039 – Role of posttranslational mechanisms in reprogramming mouse oocytes to pluripotent cells, 2009–2012, P. Svoboda
- GA CR, GA204/09/0085 – RNA silencing and long dsRNA in mammalian cells, 2009–2013, P. Svoboda
- GA CR, GAP305/10/2215 – Control of chromatin and pluripotency by microRNAs, 2010–2013, P. Svoboda
- GA Charles University, 18110 – Development and characterization of microRNA pathway inhibitor, 2010–2012, K. Podolská

1. Svoboda P, Flemr M. The role of miRNAs and endogenous siRNAs in maternal-to-zygotic reprogramming and the establishment of pluripotency. **EMBO Rep** 2010 11(8): 590–597.
2. Flemr M, Svoboda P. Ribonucleoprotein localization in mouse oocytes. **Methods** 2011 53(2): 136–141.
3. Ma J, Flemr M, Stein P, Berninger P, Malik R, Zavolan M, Svoboda P, Schultz RM. MicroRNA activity is suppressed in mouse oocytes. **Curr Biol** 2010 20(3): 265–270.
4. Flemr M, Ma J, Schultz RM, Svoboda P. P-Body Loss Is concomitant with formation of a messenger RNA storage domain in mouse oocytes. **Biol Reprod** 2010 82(5): 1008–1017.
5. Sinkkonen L, Hugenschmidt T, Berninger P, Gaidatzis D, Mohn F, Artus-Revel CG, Zavolan M, Svoboda P, Filipowicz W. MicroRNAs control de novo DNA methylation through regulation of transcriptional repressors in mouse embryonic stem cells. **Nat Struct Mol Biol** 2008 15(3): 259–267.



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