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Gender specific influence of fish oil or atorvastatin on functional properties of renal Na,K-ATPase in healthy Wistar and hypertriglyceridemic rats

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Abstract

For better understanding of pathophysiological processes leading to increased retention of sodium as a consequence of hyperlipidemia, the properties of renal Na,K-ATPase, a key enzyme involved in maintaining sodium homeostasis in the organism, were studied. Enzyme kinetics of renal Na,K-ATPase were used for characterization of ATP- and Na⁺-binding sites after administration of fish oil (FO) (30 mg day⁻¹) or atorvastatin (0.5 mg $100 \text{ g}^{-1} \text{ day}^{-1}$) to healthy Wistar rats and rats with hereditary hypertriglyceridemia of both genders. Untreated healthy Wistar and also hypertriglyceridemic female rats revealed higher Na,K-ATPase activity as compared to respective untreated male groups. Hypertriglyceridemia itself was accompanied with higher Na,K-ATPase activity in both genders. Fish oil improved the enzyme affinity to ATP and Na⁺, as indicated by lowered values of K_m and K_{Na} in Wistar female rats. In Wistar male rats FO deteriorated the enzyme in the vicinity of the Na⁺binding site as revealed from the increased K_{Na} value. In hypertriglyceridemic rats FO induced a significant effect only in females in the vicinity of the sodium binding sites resulting in improved affinity as documented by the lower value of K_{Na}. Atorvastatin aggravated the properties of Na,K-ATPase in both genders of Wistar rats. In hypertriglyceridemic rats protection of Na,K-ATPase was observed, but this effect was bound to females only. Both treatments protected renal Na,K-ATPase in a gender specific mode, resulting probably in improved extrusion of excessive intracellular sodium out of the cell affecting thus the retention of sodium in hHTG females only.

Key words: sodium pump, kidney, hereditary hypertriglyceridemia, sex difference

Introduction

Hypertriglyceridemia, independently of cholesterol level, is associated with an increased risk of cardiovascular diseases and it may be followed by development of atherosclerosis and hypertension (Hokanson *et al.* 1996). In management of hypertriglyceridemia various therapies have been selected. A systematic search of randomized controlled trials, comparing any lipid-lowering intervention with placebo or usual diet with respect to mortality, resulted in conclusion that statins and n-3 polyunsaturated fatty acids (PUFA) are the most favorable lipid-lowering interventions with reduced risks of overall and cardiac mortality (Studer *et al.* 2005). Numerous other studies have shown that in HTG treatment statins and a diet rich in omega-3 polyunsaturated fatty acids (particularly docosahexaenoic acid and eicosapentaenoic acid) lower lipid levels in plasma and have an important impact in primary and secondary prevention of cardiovascular disease and atherosclerosis as well.

For better understanding of pathophysiological processes leading to cardiovascular events as a consequence of hyperlipidemia, several experimental models have been intensively studied. One of these alternatives, known as Prague hereditary hypertriglyceridemic (hHTG) rats, was developed as a model of human hypertriglyceridemia (Vrána and Kazdová 1990). A large number of studies have revealed, that Prague hHTG rats represent a suitable model for the study of metabolic disturbances in relation to blood pressure. Numerous abnormalities of blood pressure regulation as well as alterations in the structure and function of the cardiovascular system were found in this model (for review see Zicha et al. 2006). The Prague hHTG rats showed a significant increase of diastolic blood pressure accompanied with hypertrophy of the left ventricle (Klimeš et al. 1997). In other studies increase of systolic blood pressure (Šimko et al. 2002) and hypertrophy of the right ventricle were documented (Šimko et al. 2005). The aorta is also altered in hHTG rats, as documented by quantitative and qualitative changes of endothelium and connexin 43 (Dlugosova et al. 2009). Besides the increased pressure overload also the altered retention of sodium and subsequent enlargement of circulating volume may be co-responsible for the development of cardiac hypertrophy (Klimeš et al. 1997). The kidneys of hypertriglyceridemic rats were more susceptible to cyclosporin-induced nephrotoxicity which was reduced in hHTG rats by PUFA (Bohdanecká et al. 1999). Administration of PUFA and also atorvastatin resulted in

antiarrhythmic effect via protection of cardiomyocytes and cell-to-cell junction integrity (Bacova *et al.* 2010).

The mammalian kidney plays a crucial role in maintaining the extracellular homeostasis of sodium ions. One of the key systems involved in this process is renal Na,K-ATPase or so called sodium pump which transports 3 Na⁺ ions out of the cell and 2 K⁺ ions into the cell using the energy derived from hydrolysis of one molecule of ATP. Previous studies have documented that Na,K-ATPase is sensitive to the administration of PUFA (Gerbi *et al.* 1997, Djemli-Shipkolye *et al.* 2002). Administration of atorvastatin or simvastatin to hyperlipidemic patients also increased the activity of Na,K-ATPase in erythrocytes (Broncel *et al.* 1997).

Trying to characterize the utilization of ATP and Na⁺-binding properties of renal Na,K-ATPase during hypertriglyceridemia and its treatment with PUFA and atorvastatin, the present study was designed to investigate the kinetic properties of the enzyme in male and female hHTG rats.

Methods

Animal model

Experiments were performed on adult Wistar rats and rats with hereditary hypertriglyceridemia (HTG) of both genders.

At the beginning of experiments, 3-month-old Wistar and HTG animals of both genders were divided into 3 groups (n=8 in each experimental group): Wistar and HTG rats fed with n-3 polyunsaturated fatty acid diet isolated from fish oil (eicosapentaenoic acid and docosahexaenoic acid, Vesterålen, Norway, 30 mg·day⁻¹) for two months (WfFO – Wistar females, HfFO – HTG females, WmFO – Wistar males, HmFO – HTG males), Wistar and HTG rats treated with atorvastatin (Zentiva, Slovakia, 0.5-mg·100 g⁻¹ body weight per day) for two months (WfA, HfA, WmA, HmA), and untreated Wistar and HTG control rats (WfC, HfC, WmC, HmC). All rats were allowed free access to food and drinking water. The animal room was air-conditioned and the environment was continually monitored for the temperature of 23 \pm 1°C with relative humidity of 55 \pm 10%. At the end of experiment, the rats were anesthetized by thiopental anesthesia, the excised kidneys were immediately frozen in liquid nitrogen and stored for further investigations of Na,K-ATPase properties. All experiments were approved by the Veterinary Council of the Slovak Republic

(Decree No. 289, part 139, July 9th 2003) and were conform to Principles of Laboratory Animal Care (NIH publication 83-25, revised 1985).

Sample isolation

The plasmalemmal membrane fraction from rat kidney was isolated according to Jorgensen (1974) with slight modifications. Briefly, the renal tissue was homogenized in cold isolation medium containing in mmol.l⁻¹: 250 sucrose, 25 imidazol, 1 EDTA (pH 7.4) using a tissue disruptor (3 x 10 sec at a setting of 4, Polytron PT-20). The homogenate was centrifuged at 6000g for 15 min. The sediment was re-homogenized and centrifuged again at 6000g for 15 min. The collected supernatants from both centrifugations were re-centrifuged at 48000g for 30 min and the final sediment was re-suspended in the isolation medium. An aliquot was removed for determination of proteins by the method of Lowry *et al.* (1951) using bovine serum albumin as standard.

Enzyme kinetics

ATP-kinetics of Na,K-ATPase was estimated at the temperature of 37°C measuring the hydrolysis of ATP by 10 µg plasmalemmal proteins in the presence of increasing concentrations of the substrate ATP (0.16–8.0 mmol·l⁻¹). The total volume of the medium was 0.5 ml containing (in mmol·l⁻¹): MgCl₂ 4, KCl 10, NaCl 100 and imidazole 50 (pH 7.4). After 20 min of pre-incubation in substrate-free medium, the reaction was started by addition of ATP and after 20 min the reaction was stopped by addition of 0.3 ml 12% ice-cold solution of trichloroacetic acid. The liberated inorganic phosphorus was determined according to Taussky and Shorr (1953). In order to establish the Na,K-ATPase activity, the ATP hydrolysis that occurred in the presence of Mg²⁺ only was subtracted. The Na,K-ATPase kinetics for cofactor Na⁺ was determined by the same method, in the presence of increasing concentrations of NaCl (2.0–100.0 mmol·l⁻¹). The amount of ATP was constant (8 mmol·l⁻¹). The kinetic parameters V_{max}, K_m, K_{Na} were evaluated from the obtained data by direct nonlinear regression. The parameter V_{max} represents the maximal velocity, K_m and K_{Na} values represent the concentrations of ATP or Na⁺ necessary for half maximal activation of the enzyme.

Statistical analysis

All results were expressed as mean \pm S.E.M. The significance of differences between the individual groups was determined using the one-way analysis of variance (ANOVA) by Student-Newman-Keuls test. A value of p < 0.05 was regarded as significant.

Results

When activating renal Na,K-ATPase with increasing concentrations of substrate, its activity was higher in hHTG groups throughout the applied range of ATP concentrations as compared to Wistar groups (Fig. 1). This effect was observed in female as well as in male groups. Evaluation of the data by nonlinear regression resulted in higher V_{max} value in the WfC group by 23% as compared to the WmC group. Similar gender specificity was observed also in hHTG animals where the V_{max} value in the HfC group was by 18% higher as compared to the HmC group. Hypertriglyceridemia itself induced a significant increase of V_{max} value in both genders, representing 25% in males and 21% in females. The K_m value remained similar in all four compared groups (Table 1).

Activation of the enzyme with increasing concentrations of cofactor Na⁺ resulted again in higher activity in hHTG groups throughout the applied range of NaCl concentrations as compared to Wistar groups in both genders (Fig. 2). Evaluation of kinetic parameters revealed higher V_{max} value in the WfC group by 16% as compared to the WmC group. Similar gender specificity was observed also in hHTG animals where the V_{max} value in the HfC group was by 16% higher as compared to the HmC group. Hypertriglyceridemia itself induced a significant increase of V_{max} value in both genders representing 22%. The K_{Na} value showed gender specificity in Wistar rats, where this parameter was higher by 24% in WfC as compared to the WmC group (Table 1).

Administration of fish oil to Wistar females induced a slight increase of Na,K-ATPase activities in the presence of low concentrations of ATP, but in the presence of higher concentrations exceeding 2 mmol.l⁻¹ ATP the activities were slightly lower as compared to untreated controls (Fig. 3). This biphasic effect of fish oil administration was reflected in a decrease of the K_m value by 15% in WfFO as compared to the WfC group. In Wistar males the enzyme activity was similar

throughout the concentration range of ATP applied (Fig. 5), without any alterations in kinetics parameters (Table 2).

Administration of atorvastatin to Wistar rats was followed by decreased enzyme activities throughout the whole concentration range of ATP in both genders (Fig. 3). Evaluation of kinetics parameters resulted in significant decrease of V_{max} value in females only, as shown by 23% decrease in WfA compared to the WfC group (Table 2).

During activation of renal Na,K-ATPase with increasing concentrations of NaCl, a slight stimulation of the enzyme activity was observed in Wistar females treated with fish oil. This effect we observed in the presence of lower concentrations below 10 mmol.l⁻¹ of NaCl, above this concentration the effect was lost (Fig. 4). Evaluation of this effect revealed unchanged V_{max} with decreased K_{Na} value by 17% in the WfFO as compared to the WfC group (Table 2). Administration of fish oil to Wistar males induced again a slight stimulation of Na,K-ATPase activity but the stimulatory effect increased stepwise with increasing concentration of NaCl (Fig. 4) resulting in increased values of V_{max} by 17% and K_{Na} by 25% in WmFO as compared to the WmC group (Table 2).

Administration of atorvastatin to Wistar rats was followed by decreased enzyme activities throughout the whole concentration range of NaCl in both genders (Fig. 4). Evaluation of kinetics parameters resulted in significant decrease of V_{max} value by 21% in WfA as compared to the WfC group. The K_{Na} value was significantly increased by 22% in males, however in females the value was decreased by 21%, as compared to respective controls (Table 2).

Administration of fish oil to hHTG rats induced a slight increase of Na,K-ATPase activities throughout the investigated concentration range of ATP in both genders as compared to untreated animals (Fig. 5), however this slight effect was not reflected either in changes of V_{max} or of K_m values (Table 3).

Administration of atorvastatin to hHTG rats induced a slight stimulation in female rats only (Fig. 5), resulting in an 18% increase of V_{max} and 15% of K_m value (Table 3).

When activating renal Na,K-ATPase with increasing concentrations of NaCl, we observed a slight stimulation of the enzyme activity only in females of hHTG animals treated with fish oil (Fig. 6). Evaluation of this effect revealed significant decrease of K_{Na} value by 11% in HfFO as compared to the HfC group (Table 3).

Administration of atorvastatin to hHTG female rats was followed by increased enzyme activities throughout the whole concentration range of NaCl (Fig. 6), resulting in 14% increase of V_{max} value in HfA as compared to the HfC group (Table 3). On the other hand, in males we observed a slight decrease of enzyme activity below 10 mmol·l⁻¹ concentration of NaCl (Fig. 6) resulting in a small but statistically significant increase of K_{Na} value by 12% in HmA as compared to the HmC group (Table 3).

Discussion

There are cumulative data indicating that sodium transport across the cell membrane is gender dependent (Grikiniene *et al.* 2004). Healthy females showed a significantly lower intracellular concentration of Na⁺ (Smith *et al.* 1988, Taylor *et al.* 1991) as compared to males. Gender-related differences in intracellular Na⁺ concentration result most likely from differences in the function of sodium transport systems including Na,K-ATPase. This hypothesis is confirmed also by our present study, documenting increased activity of renal Na,K-ATPase in control female Wistar rats. This is in agreement with previous data of Quintas *et al.* (1997). However, some other experimental studies documented similarity in activities and expression of renal Na,K-ATPase during normal physiological conditions in both genders (Fekete *et al.* 2004, 2006).

Our data provided evidence that in the situation of pathophysiological overload the activity of renal Na,K-ATPase was again increased in females as comaperd to male hHTG rats. The increased activity of renal Na,K-ATPase in hHTG females may be explained by a hypothetically higher presence of the active enzyme molecules in the renal tissue in females as compared to males. This hypothesis is supported by our data concerning the increased V_{max} value in both types of enzyme kinetics in all female groups as compared to respective male groups. This finding, suggesting better adaptation of the enzyme to pathophysiological overload in females, is supported by the observation of higher mRNA expression of Na,K-ATPase catalytic α_1 subunit, as well as higher activity in female rats as compared to males after ischemia reperfusion injury in the kidney (Fekete *et al.* 2004). The sexual disparity of Na,K-ATPase properties was demonstrated in various pathophysiological conditions also in other tissues, like the aorta (Palacios *et al.* 2006), heart (Vlkovicova *et al.* 2005) and erythrocytes (Smith *et al.* 1988).

The mechanism of the protection of Na,K-ATPase in females may be ascribed to estradiol, because it was previously shown that this hormone, besides other effects, stimulates directly and also indirectly the Na,K-ATPase molecule in the cardiac tissue (Dzurba *et al.* 1997). This explanation is supported also by the finding that the significantly higher activity of the sodium pump is a consequence of estradiol-induced increase in Na,K-ATPase α 2 subunit expression in the female aorta (Palacios *et al.* 2004) and α 1 subunit expression in the female kidney (Fekete *et al.* 2004). Estradiol also stimulated Na,K-ATPase activity and the expression of α 1 subunit via multiple signaling cascades, including phosphatidyl inositol-3 kinase and p42/44 mitogen-activated protein kinase in vascular smooth muscle (Sudar *et al.* 2008).

It is necessary to mention that renal Na,K-ATPase activity was higher in both genders of hHTG rats as compared to respective control Wistar groups. So the improved extrusion of intracellular sodium out of the cell as a consequence of increased Na,K-ATPase activity documented in the present study, may be the reason for increased retention of sodium in hHTG rats, as shown by Klimeš *et al.* (1997).

Previous studies reported that the Na,K-ATPase was sensitive to the administration of n-3 polyunsaturated fatty acids (PUFA). The effect of fatty acids was dependent on physiological or pathophysiological conditions and it showed also tissue dependence. The positive effect of PUFA on Na,K-ATPase was documented during diabetes in red blood cells (Djemli-Shipkolye *et al.* 2002), in hearts (Gerbi *et al.* 1997) and in sciatic nerves (Gerbi *et al.* 1999). On the other hand, Djemli-Shipkolye *et al.* (2002) observed a negative effect of PUFA on Na,K-ATPase in sciatic nerves in diabetic as well as in healthy rats. It should be mentioned that all previous studies were carried out exclusively on males.

It is known that Na,K-ATPase reveals different properties in males and females, as discussed previously. Therefore in our study we tried to broaden the knowledge about the effect of PUFA from the view of possible gender specificity. Our new data suggest a gender specific influence of PUFA on renal Na,K-ATPase. In Wistar female rats, fish oil improved the enzyme affinity to ATP and Na⁺, as indicated by lowered values of K_m and K_{Na}. In Wistar males fish oil caused deterioration of the enzyme in the vicinity of the Na⁺-binding site, as revealed from the increased K_{Na} value. In hHTG rats fish oil induced a significant effect only in females in the vicinity of the sodium binding sites, resulting in improved affinity, as documented by a lower value of K_{Na}. Our data indicate that renal Na,K-ATPase in healthy and also in

hypertriglyceridemic female rats seems to be more sensitive to the protective effect of PUFA administration.

Atorvastatin did not influence the vicinity of the ATP binding site in Wistar males nor in Wistar females, as suggested by unchanged K_m values. In the vicinity of the Na⁺ binding site of Na,K-ATPase in male Wistar rats, atorvastatin induced probably conformational changes, resulting in deteriorated affinity to sodium, as suggested by increased value of K_{Na} . Concerning Wistar female rats, atorvastatin reduced the number of active Na,K-ATPase molecules as indicated by the lowered V_{max} value for both types of enzyme activation. This finding may suggest a decrease of transmembraneous transport of sodium out of the cell as a consequence of decreased Na,K-ATPase activity, presumably followed by lowered retention of sodium in normotensive Wistar rats.

In male hypertriglyceridemic rats atorvastatin did not affect the functional properties of renal Na,K-ATPase, while in hHTG female rats it stimulated Na,K-ATPase in the whole concentration range of ATP or Na⁺. This effect is probably caused by an increased number of active enzyme molecules, as suggested by increased V_{max} value. So the enzyme in female rats treated with atorvastatin was capable to increase its activity also in the presence of high concentrations of ATP/Na⁺, while the enzyme in untreated hHTG rats was already saturated. Administration of atorvastatin to healthy Wistar rats was followed by deteriorated properties of the renal Na,K-ATPase in both genders. While a positive effect was observed in the protection of renal Na,K-ATPase in hypertriglyceridemic rats, but this effect was strictly bound to females only.

In conclusion, treatment with fish oil or atorvastatin improved the functional properties of renal Na,K-ATPase in a gender specific manner, inducing probably improved extrusion of intracellular sodium out of the cell affecting thus the retention of sodium in hHTG females only.

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Table.1. Kinetic parameters of renal Na,K-ATPase during activation with ATP and NaCl in Wistar rats and hypertriglyceridemic (hHTG) rats divided in to the following groups: control female Wistar rats (WfC), control female hHTG rats (HfC), control male Wistar rats (WmC), control male hHTG rats (HmC). Data represent means \pm S.E.M, n=8 in each group. a: p < 0.05 as compared to the WmC group, b: p < 0.05 as compared to the HmC group.

Groups of	V _{max}	K _m	V _{max}	K _{Na}
rats	ATP-kinetics		Na-kinetics	
WmC	31.9 ± 2.5	0.83 ± 0.04	33.5 ± 2.1	11.25 ± 0.59
WfC	39.1 ± 2.6 ^a	0.92 ± 0.04	38.9 ± 2.1 ^a	13.96 ± 0.75 ^a
HmC	39.9 ± 2.0 ^a	0.84 ± 0.05	40.9 ± 1.6 ^a	10.14 ± 0.34
HfC	$47.2 \pm 2.0^{b,c}$	0.93 ± 0.03	$47.4 \pm 2.3^{b,c}$	11.16 ± 0.43 ^b

Table.2. Effect of *in vivo* administration of fish oil and atorvastatin on kinetic parameters of renal Na,K-ATPase during activation with ATP and NaCl in Wistar rats divided in to the following groups: control male rats (WmC), male rats treated with fish oil (WmFO), male rats treated with atorvastatin (WmA), control female rats (WfC), female rats treated with fish oil (WfFO), male rats treated with atorvastatin (WmA). Data represent means \pm S.E.M, n=8 in each group. a: p < 0.05 as compared to the WfFO group, b: p < 0.05 as compared to the WfC group, c: p < 0.05 as compared to the WfFO group.

Groups of	V _{max}	K _m	V _{max}	K _{Na}
rats	ATP-kinetics		Na-kinetics	
WmC	31.9 ± 2.5	0.83 ± 0.04	33.5 ± 2.1	11.25 ± 0.59
WmFO	34.6 ± 2.4	0.89 ± 0.05	39.3 ± 2.0^{a}	14.10 ± 0.93 ^a
WmA	29.5 ± 0.6	0.94 ± 0.04	31.5 ± 0.1	13.78 ± 0.62 ^a
WfC	39.1 ± 2.6 ^a	0.92 ± 0.04	38.9 ± 2.1 ^a	13.96 ± 0.75 ^a
WfFO	36.1 ± 1.9	0.78 ± 0.04^{b}	39.6 ± 2.3	11.61 ± 0.39 ^b
WfA	30.2 ± 0.6^{b}	$0.95 \pm 0.05^{\circ}$	30.8 ± 1.1 ^b	11.02 ± 0.75 ^b

Table. 3. Effect of *in vivo* administration of fish oil and atorvastatin on kinetic parameters of renal Na,K-ATPase during activation with ATP and NaCl in hypertriglyceridemic (hHTG) rats divided in to the following groups: control male rats (HmC), male rats treated with fish oil (HmFO), male rats treated with atorvastatin (HmA), control female rats (HfC), female rats treated with fish oil (HfFO), male rats treated with atorvastatin (HmA). Data represent means \pm S.E.M, n=8 in each group. a: p < 0.05 as compared to the HmC group, b: p < 0.05 as compared to the HmC group, d: p < 0.05 as compared to the HmC group.

Groups of	V _{max}	K _m	V _{max}	K _{Na}
rats	ATP-kinetics		Na-kinetics	
HmC	39.9 ± 2.0	0,84 ± 0.05	40.9 ± 1.6	10.14 ± 0.34
HmFO	41.4 ± 1.2	0.80 ± 0.03	40.7 ± 1.4	10.65 ± 0 33
HmA	39.9 ± 1.7	0.85 ± 0.03	41.5 ±1.4	11.36 ±0.29 ^a
HfC	47.2 ± 2.0 ^a	0.93 ± 0.03	47.4 ± 2.3 ^a	11.16 ± 0.43
HfFO	50.3 ± 2.2^{b}	0.91 ± 0.09	50.3 ± 2.3^{b}	9.94 ± 0.42^{d}
HfA	55.5 ± 1.2 ^{c,d}	1.07 ± 0.04 ^{c,d}	54.1 ±0.9 ^{c,d}	11.30 ± 0.63



Fig. 1. Activation of renal Na,K-ATPase by low concentrations of substrate ATP in Wistar rats and hypertriglyceridemic (hHTG) rats divided in to the following groups: **left** panel~ control female Wistar rats (WfC), control female hHTG rats (HfC), **right panel**~ control male Wistar rats (WmC), control male hHTG rats (HmC). Insert: activation of the enzyme in the whole range of ATP concentrations.



Fig. 2. Activation of renal Na,K-ATPase by low concentrations of NasCl in Wistar rats (W) and hypertriglyceridemic (hHTG) rats divided in to the following groups: left panel~ control female Wistar rats (WfC), control female hHTG rats (HfC), right panel~ control male Wistar rats (WmC), control male hHTG rats (HmC). Insert: activation of the enzyme in the whole range of ATP concentrations.



Fig. 3. Effect of *in vivo* administration of fish oil and atorvastatin on the activation of renal Na,K-ATPase by low concentrations of substrate ATP in Wistar rats divided in to the following groups: left panel~ control female rats (WfC), female rats treated with fish oil (WfFO), female rats treated with atorvastatin (WfA), right panel~ control male rats (WmC), male rats treated with fish oil (WmFO), male rats treated with fish oil (WmFO), male rats treated with atorvastatin (WmA). Insert: activation of the enzyme in the whole range of ATP concentrations.



Fig. 4. Effect of *in vivo* administration of fish oil and atorvastatin on the activation of renal Na,K-ATPase by low concentrations of NaCl in Wistar rats divided in the following groups: left panel~ control female rats (WfC), female rats treated with fish oil (WfFO), female rats treated with atorvastatin (WfA), right panel~ control male rats (WmC), male rats treated with fish oil (WmFO), male rats treated with fish oil (WmFO), male rats treated with atorvastatin (WfA). Insert: activation of the enzyme in the whole range of ATP concentrations.



Fig. 5. Effect of *in vivo* administration of fish oil and atorvastatin on the activation of renal Na,K-ATPase by low concentrations of substrate ATP in hypertriglyceridemic (hHTG) rats divided in to the following groups: **left panel~** control female rats (HfC), female rats treated with fish oil (HfFO), female rats treated with atorvastatin (HfA), **right panel~** control male rats (HmC), male rats treated with fish oil (HmFO), male rats treated with atorvastatin (HmA). Insert: activation of the enzyme in the whole range of ATP concentrations.



Fig. 6. Effect of *in vivo* administration of fish oil and atorvastatin on the activation of renal Na,K-ATPase by low concentrations of NaCl in hypertriglyceridemic (hHTG) rats divided in to the following groups: left panel~ control female rats (HfC), female rats treated with fish oil (HfFO), female rats treated with atorvastatin (HfA), right panel~ control male rats (HmC), male rats treated with fish oil (HmFO), male rats treated with fish oil (HmFO), male rats treated with fish oil (HmA). Insert: activation of the enzyme in the whole range of ATP concentrations.

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