# Physiological Research Pre-Press Article

## Antioxidant Defense is Increased in Aged Hearts Following Omega-3 Supplementation in the Absence of Changes in Inflammation

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Short title: Omega-3, oxidative stress, and inflammation in aged heart

#### Summary

The purpose of this study was to determine the effect of a 15-week omega-3 rich diet on age-related differences in myocardial antioxidant defense and inflammation. 20 mature (M) (6 mo.) and 20 old (O) (15 mo.) male Fisher 344 rats were assigned to two diet groups: Control (CON) or Fish Oil (FO). Following the diet, animals were sacrificed and left ventricular (LV) heart tissue was harvested for biochemical assays and western blot analysis. No differences were observed in expression of LV interleukin-6 (IL-6) and tumor necrosis factor- $\alpha$  as well as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) production between MCON and OCON. However, LV catalase protein expression and activity were increased in OCON vs. MCON and accompanied by increased expression of superoxide dismutase (SOD)-1. In contrast, LV IL-6 was lower in MFO vs. old rats, and LV H<sub>2</sub>O<sub>2</sub> was decreased in MFO and OFO relative to respective control groups. Protein expression and activity of LV catalase and SOD-1 expression were increased in OFO similarly to OCON, but LV SOD activity was also increased in OFO vs. mature rats. In summary, FO supplementation increased myocardial antioxidant defense in all animals and augmented age-associated increases in antioxidant capacity in the absence of changes in inflammation.

Key words: aging, oxidative stress, omega 3 fatty acids, inflammation

The risk of heart disease (HD) increases with age and diet is an important modifiable risk factor that may interact with reactive oxygen species (ROS) and inflammatory markers (de Winther, Kanters, Kraal, Hofker 2005, Devaraj, Tang, Adams-Huet, Harris, Seenivasan, de Lemos, Jialal 2007). Diets high in total fat, cholesterol, and particularly saturated fat have been shown through several long-term epidemiological studies to increase the risk of atherosclerosis and subsequent development of HD (Shekelle, Shryock, Paul, Lepper, Stamler, Liu, Raynor 1981, Kromhout, Menotti, Bloemberg, Aravanis, Blackburn, Buzina, Dontas, Fidanza, Giampaoli, Jansen, et al. 1995, Hu, Stampfer, Rimm, Ascherio, Rosner, Spiegelman, Willett 1999, Millen, Quatromoni 2001, Xu, Eilat-Adar, Loria, Goldbourt, Howard, Fabsitz, Zephier, Mattil,Lee 2006). While high intake of atherogenic lipids such as saturated fat are detrimental to the heart (Esrey, Joseph, Grover 1996), polyunsaturated fats, particularly omega-3 fatty acids have been shown to be beneficial (Sinclair 1956, Kromhout, Bosschieter, de Lezenne Coulander 1985, Dolecek, Granditis 1991). Evidence from several observational studies, randomized controlled trials, and animal studies suggest that habitual intake of omega-3 fatty acids reduce the risk of HD (Kris-Etherton, Harris, Appel, American Heart Association. Nutrition 2002, Wang, Harris, Chung, Lichtenstein, Balk, Kupelnick, Jordan, Lau 2006, von Schacky, Harris 2007, Serhan, Chiang 2008).

Omega-3 fatty acids are long chain polyunsaturated fatty acids, including eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) recommended by several societies including the American Heart Association (Kris-Etherton *et al.* 2002) and the Guidelines on Cardiovascular Prevention of nine European societies (Graham, Atar, Borch-Johnsen, Boysen, Burell, Cifkova, Dallongeville, De Backer, Ebrahim, Gjelsvik, Herrmann-Lingen, Hoes, Humphries, Knapton, Perk, Priori, Pyorala, Reiner, Ruilope, Sans-Menendez, Op Reimer, Weissberg, Wood, Yarnell, Zamorano,Guidelines 2007). The mechanism behind omega-3 related cardioprotection has been attributed to both anti-inflammatory and antioxidant effects. Inflammatory plasma markers including interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF-α) decrease following EPA and DHA administration (Bhatnagar,Durrington 2003, Ferrucci, Cherubini, Bandinelli, Bartali, Corsi, Lauretani, Martin, Andres-Lacueva, Senin,Guralnik 2006, Farzaneh-Far, Harris, Garg, Na,Whooley 2009) and circulating levels of the antioxidant glutathione peroxidase increase with omega-3 supplementation (Castillo, Arias,Farias 2014). In addition, chronic omega-3 supplementation reduces myocardial ischemic injury in rodents as related to reinforcement of endogenous antioxidant defense (Castillo *et al.* 2014).

Despite the established cardioprotective effects of omega-3 fatty acids, it remains unknown whether supplementation can alter antioxidant defense and inflammation in the hearts of aging rodents. Therefore, the purpose of this study was to determine the effect of a 15-week omega-3 rich diet using (FO) supplementation on age-related differences in myocardial antioxidant defense and inflammation. We hypothesized that FO supplementation would augment antioxidant defense and lower levels of inflammation in the heart of aged animals.

Our experiments were approved by the University of Delaware Animal Care and Use Committee and conducted following the guidelines established by the National Institutes of Health Office of Laboratory Animal Welfare for the use of animals in research. Twenty mature and 20 old male Fisher 344 rats (Harlan Laboratories) were housed in a temperature controlled facility and maintained on a 12 hour light-dark cycle. Both mature and old animals were randomly assigned to one of two diet groups: (1) control (CON); or (2) fish oil (FO). LabDiet provided the commercial diets (Quakertown, PA). The FO diet (LabDiet 5001 with 10% Menhaden Fish oil added) contained 2.88% omega-3 fatty acids while the CON diet (Modified 5001 with 8% total fat) contained 0.43%. The Menhaden fish oil contained 30% omega-3 fatty acids with a 1.16:1 ratio of EPA to DHA. Menhaden fish oil supplementation is a common approach to increase omega 3 content of rodent chow and has been successfully used elsewhere (Hock, Beck, Bodine, Reibel 1990). The four groups were mature control (MCON), mature fish oil (MFO), old control (OCON), and old fish oil (OFO). Animals were maintained on the diet for 15-weeks and thus were approximately 6 and 15 months when sacrificed. Animals were anesthetized with ketamine-xylazine (100mg/kg) and sacrificed on the last day of the diet. The heart was removed and the left ventricle (LV) was separated, minced, and homogenized using glass on glass homogenization in cold 100 mM potassium phosphate buffer (1:20 wt/vol; pH 7.4). The homogenate was centrifuged (4°C) for 10 min at 400 g. The supernatant was decanted and assayed to determine total protein content using the Bradford Protein assay (Bradford 1976) along with the activities of superoxide dismutase (SOD), catalase (CAT), and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). SOD activity was measured using a colorimetric assay from Cayman Chemical (Ann Arbor, Michigan), CAT activity was measured using an assay by Aebi et al. (Aebi 1984) and  $H_2O_2$  was measured using the Amplex Red Hydrogen Peroxide/peroxidase kit from Invitrogen (Molecular Probes, Grand Island, NY). All activities were normalized

to protein content. Tumor necrosis factor-alpha (TNF-α) and IL-6 were measured using ELISA kits from R&D Systems, Inc. (Minneapolis, MN).

To determine the effect of fish oils on induction of myocardial antioxidants, polyacrylamide gel electrophoresis and immunoblotting were performed. Briefly, LV samples from mature and old animals were homogenized and one-dimensional sodium dodecyl sulfate polyacrylamide gel electrophoresis was performed to separate proteins by molecular weight. After separation, proteins were transferred to nitrocellulose membranes (0.45 mm thick, Bio-Rad; Hercules, CA) using the Bio-Rad transblot electrophoretic transfer cell at a constant voltage of 100V for one hour. After protein transfer, membranes were blocked overnight at 4°C using 5% nonfat dry milk. Blots were incubated for 1 hour at room temperature using the following antibodies from Santa Cruz Biotechnology (Santa Cruz, CA): mouse monoclonal antibody for SOD-1 (sc-271014), mouse monoclonal antibody for SOD-2 (sc-58425), mouse monoclonal antibody for catalase (sc-271803), mouse monoclonal antibody for GPx-1/2 (sc-133160), and beta-actin (sc-47778). Antibodies were then reacted with a rabbit polyclonal secondary antibody to mouse IgG (ab6728) (Abcam; Cambridge, MA) for one hour at room temperature. Membranes were exposed to chemiluminescent reagents (GE Healthcare) for 1 minute and developed. Bands were normalized to beta-actin and quantification of the bands was performed using Image J software (NIH).

A two-way ANOVA was run to test for an interaction between treatment and age. When a significant main effect was seen, a Bonferroni post hoc test was run to test for group differences. The level of significance was set at P<0.05 and values are reported as means ± SE. At the end of 15 weeks of diet, both aged groups were significantly heavier (OCON: 485±9; OFO: 512±14 g) than MCON (448±4 g; P<0.05) while MFO (473±3 g) was only different from MCON (P<0.05). Heart mass was lower in the MCON group (1.25±0.01 g) relative to the other three groups (MFO, 1.30±0.03; OCON, 1.32±0.03; OFO, 1.42±0.03 g; p<0.05) which coincided with the differences in total body weight. However, once heart weight was normalized to body weight, no differences were seen among groups.

To determine the effect of the FO diet on markers of inflammation, LV TNF- $\alpha$  and IL-6 were measured. Both are thought to play an important role in cardiovascular disease risk and development (Cesari, Penninx, Newman, Kritchevsky, Nicklas, Sutton-Tyrrell, Tracy, Rubin, Harris,Pahor 2003). No differences were seen in TNF- $\alpha$  concentrations among the groups; however, IL-6 was greater in both aged groups (p<0.05) with no effect of diet suggesting that the FO supplementation was not sufficient to overcome the aging effects on myocardial inflammation. IL-6 has been shown to be a very reliable inflammatory marker of aging (Johnson 2006) and is increased in healthy, older subjects (Wei, Xu, Davies,Hemmings 1992). Further, two months of omega 3 supplementation lowered circulating IL-6 levels in atherosclerotic patients (Eftekhari, Aliasghari, Beigi,Hasanzadeh 2014). IL-6 was measured in the aged rat heart in this study in the absence of apparent cardiovascular disease, which may explain the discrepancies with previous findings.

Endogenous antioxidant expression and activity was increased in the LV of OCON animals. LV CAT activity was increased in OCON (P<0.05; Table 1) compared to MCON in conjunction with increased protein expression of SOD-1 and CAT (Figure 1).

OFO animals had similar increases in LV CAT expression and activity as well as SOD-1 expression but LV SOD activity was also increased in the OFO vs. mature rats (P<0.05; Table 1). In addition,  $H_2O_2$  levels were decreased in both MFO and OFO relative to their respective control groups. No differences were observed in protein expression of GPx-1/2 between all groups and SOD-2 expression trended similarly as SOD-1 in old animals but did not reach statistical significance (P=0.09 for an effect of age). Overall, endogenous myocardial antioxidant defense was increased with FO supplementation in all animals as indicated by decreased levels of ROS production. FO also augmented age-associated increases in antioxidant capacity with increased SOD activity and decreased  $H_2O_2$  levels (Table 1). Similarly, isolated cardiac myocytes from omega-3 supplemented rats have demonstrated less asynchronous contractile activity in the presence of  $H_2O_2$  and had increased SOD expression relative to a control and saturated fat supplemented diet (Dolecek,Granditis 1991).

Our findings are consistent with previous investigation demonstrating increased antioxidant capacity with omega-3 supplementation (Erdogan, Fadillioglu, Ozgocmen, Sogut, Ozyurt, Akyol,Ardicoglu 2004, Castillo *et al.* 2014). Eight weeks of DHA and EPA supplementation via gavage providing 1.2 g/kg resulted in an increase in antioxidant activity (CAT, SOD1, GPx) in 4-month old rodents (Castillo *et al.* 2014) while others have shown an increase in SOD and a reduction in oxidative stress in rodents supplemented orally with a fish oil rich capsule containing 0.4 g/kg (Erdogan *et al.* 2004). In this study, only SOD activity became significantly elevated with the FO diet in old animals as compared to controls. Our dose of fish oil was provided in the standard rat chow at 28.8 g/kg as opposed to gavage supplementation. This is similar to other

studies that have provided diets rich in omega 3 fatty acids in the rodent chow (Hock *et al.* 1990, Yang, Saldeen, Nichols,Mehta 1993). While our omega 3 content was higher, access to chow was not controlled. Based on typical rodent consumption, our animals consumed approximately 0.7g of omega 3 fatty acids each day. Further, the differences in antioxidant expression profiles between studies could be due to the age and/or strain of animals utilized. However this study does not suggest that omega-3 supplementation increases levels of oxidative stress as reported previously (Sinclair 1956).

In summary, the purpose of this study was to determine the effects of 15 weeks of an omega-3 rich diet on markers of inflammation and endogenous antioxidant defenses in the hearts of mature and old rodents. The primary finding was that a diet rich in omega-3 fatty acids increased myocardial antioxidant defenses in all animals and augmented age-associated increases in antioxidant capacity in the absence of a change in inflammation. Further study is warranted to better characterize whether these changes result in functional changes in the aged heart.

#### Acknowledgement

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## Figure Legend

**Figure 1.** Representative western blots (A) and quantification (B) of left ventricular antioxidant enzymes for MCON, MFO, OCON and OFO groups. This includes superoxide dismutase-1 (SOD1), superoxide dismutase-2 (SOD2), catalase (CAT), and glutathione peroxidase 1/2 (GPx1/2). CAT and SOD1 were increased in the older groups alone. Values are mean  $\pm$  SEM and samples were normalized to MCON. \* P<0.05 vs. respective MCON; + P<0.05 vs. respective MFO.

## Table 1: Markers of Oxidative stress and inflammation

	MCON	MFO	OCON	OFO
Antioxidants				
Catalase (U/mg protein)	3.01 ± 0.33	3.95 ± 0.24	4.49 ± 0.52 *	4.25 ± 0.30*
SOD (U/mg protein)	24.1 ± 1.3	25.4 ± 1.8	26.9 ± 1.6	33.8 ± 3.1*§
Oxidative Stress				
H <sub>2</sub> O <sub>2</sub> (nM/mg protein)	123.7 ± 9.2	105 ± 7.9*	137 ± 18.7	97.1 ± 7.7¶
Inflammatory Markers				
IL-6 (pg/mg)	30.3 ±1.85	26.6 ± 0.67	37.8 ± 3.8*	37.7 ± 2.23§
TNF-α (pg/mg)	10.3 ± 0.27	13.1 ± 1.03	11.9 ± 1.2	11.4 ± 0.84

Values are means ± SE. N=10 in each group. H<sub>2</sub>O<sub>2</sub>, hydrogen peroxide; MCON, mature control; MFO, mature fish oil diet; OCON, old control diet; OFO, old fish oil diet. \* P < 0.05 from MCON; § P < 0.05 from MFO; ¶ P<0.05 from OCON.

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## **FIGURE 1**

