# **Physiological Research Pre-Press Article**

1	INVITED REVIEW
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3	Vitamin D <sub>3</sub> Action within the Ovary – an Updated Review
4	Malgorzata GRZESIAK
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6	Department of Endocrinology, Institute of Zoology and Biomedical Research, Jagiellonian
7	University in Krakow, Gronostajowa 9, 30-387 Krakow, Poland
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9	Corresponding author
10	M. Grzesiak, Department of Endocrinology, Institute of Zoology and Biomedical Research,
11	Jagiellonian University in Krakow, Gronostajowa 9, 30-387 Krakow, Poland. E-mail:
12	m.e.grzesiak@uj.edu.pl
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25 Short title: Vitamin  $D_3$  in the ovary.

## 26 Summary

27	Vitamin $D_3$ is well-known as a major regulator of calcium and phosphorus homeostasis. A
28	growing body of evidence highlights its crucial role in the regulation of reproductive
29	processes in females. The role of vitamin $D_3$ in the female reproductive tract has been
30	extensively investigated because its receptor is abundant in reproductive organs, including
31	ovary. Importantly, besides expression of vitamin $D_3$ receptor, the ovary is an extrarenal site
32	of vitamin $D_3$ metabolism. The influence of vitamin $D_3$ on follicular development and ovarian
33	steroidogenesis has been investigated. Furthermore, vitamin D3 deficiency has also been
34	associated with polycystic ovary syndrome, premature ovarian failure and ovarian cancer. The
35	objective of this review is to summarize our knowledge about the contribution of vitamin $D_3$
36	to physiological and pathological processes within the ovary.
37	Key words
38	Vitamin $D_3$ • Vitamin $D_3$ receptor • Ovary • Polycystic Ovary Syndrome • Premature Ovarian
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51	Vitamin $D_3$ (VD <sub>3</sub> ) deficiency is recognized as a global problem which increases the
52	risk of many chronic diseases. The status of $VD_3$ in the organism depends on sun exposure,
53	diet, intake of VD <sub>3</sub> supplements, lifestyle and genetic factors (DeLuca 2004). It is well-known
54	that $VD_3$ is predominantly involved in the regulation of calcium and phosphorus homeostasis
55	and crucial for bone mineralization. However, a growing body of literature indicates its
56	pleiotropic actions within the organism including an influence on various physiological and
57	pathological processes. The classical VD <sub>3</sub> target tissues are the intestine, kidneys and bones.
58	Importantly, among the non-classical sites of VD <sub>3</sub> action are tissues of the female
59	reproductive tract. VD <sub>3</sub> receptor (VDR) and VD <sub>3</sub> metabolic enzymes have been found in the
60	ovary, uterus, fallopian tube, vagina and placenta of both human and animals, confirming the
61	direct role of VD <sub>3</sub> in these organs (Lerchbaum and Obermayer-Pietsch 2012). In recent years
62	there have been an increasing number of scientific papers suggesting a correlation between
63	low VD <sub>3</sub> level and reduced fertility, metabolic and endocrine disorders, polycystic ovary
64	syndrome (PCOS), premature ovarian failure (POF) and ovarian cancer (Muscogiuri et al.
65	2017). This review focuses on the influence of $VD_3$ on physiological processes within the
66	ovary as well as its contribution to ovarian pathologies that is summarized in Figure 1.
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68	VD <sub>3</sub> metabolism and mechanism of action
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70	The main source of circulating $VD_3$ is endogenous synthesis in the skin following
71	ultraviolet-B irradiation (UVB). Only small amount of VD3 is derived from the diet (fatty fish,
72	cod-liver oil, milk, eggs) or supplements (Bikle 2014). In keratinocytes, 7-dehydrocholesterol
73	is converted to previtamin D3 under UVB. Next, previtamin D3 undergoes isomerization
74	under the influence of the body's thermal energy and forms biologically inactive VD <sub>3</sub>
75	(cholecalciferol). $VD_3$ is released from the keratinocyte membranes into the extracellular

space and hence into the blood, where it is transported with vitamin D binding proteins 76 77 (VDBP). In the liver, hydroxylation to 25(OH)D<sub>3</sub> (25-hydroxycalciferol, calcidiol) takes place in the presence of 25-hydroxylases (e.g. CYP2R1, CY24A1). A second hydroxylation in the 78 79 kidneys involves 1α-hydroxylase (CYP27B1) action, resulting in biologically active VD<sub>3</sub>, i.e. 1α,25(OH)<sub>2</sub>D<sub>3</sub> (1α, 25-dihydroxycholecalciferol, calcitriol). Both calcidiol and calcitriol may 80 be degraded as a result of further hydroxylation by CYP24A1(Christakos et al. 2016) (Figure 81 82 2). The concentration of calcitriol circulating in the blood is not a reliable indicator of the  $VD_3$ level in the body because its content and metabolism are controlled by parathormone and 83 depend on the concentration of calcium and phosphorus. Thus, calcidiol is considered the best 84 85 indicator due to its long half-life and lack of mechanisms regulating its level (DeLuca 2004). The biological effect of  $1\alpha$ , 25(OH)<sub>2</sub>D<sub>3</sub> on target cells is mediated by VDR that belongs 86 to the superfamily of ligand-activated steroids receptor and acts as a transcriptional factor. 87 88 VDR is composed of a short N-terminal domain, a highly conserved DNA binding domain, a hinge region and an α-helical C-terminal ligand binding domain (Christakos et al. 2016). 89 Calcitriol binding to the ligand binding domain induces heterodimerization of VDR with the 90 9-cis-retinoic acid receptor (RXR). The VDR-RXR complex is translocated to the nucleus and 91 binds to the VD<sub>3</sub> response element (VDRE) regulating the expression of target genes. The 92 93 activation/inhibition of transcription requires also the recruitment of wide range of coregulators (Keane et al. 2017). Studies on the structure of VDR have shown the presence of 94 two overlapping ligand binding sites in the C-terminal domain. They were defined as the 95 genomic pocket (VDR-GP) and the alternative pocket (VDR-AP). The first of these initiates 96 the genomic response, while the second one can cause both genomic and non-genomic effects 97 (Mizwicki et al. 2004). The final signaling pathway triggered after ligand (calcitriol or its 98 synthetic analogues) binding to VDR also depends on VDR localization in the cell. The 99 receptor has been found in the cytoplasm/nucleus and mitochondria as well as in cell 100

101	membrane cavities, i.e. caveolae. VDR located in caveolae triggers a rapid cell response by
102	activating receptors associated with G proteins, phosphatases, kinases and ion channels
103	(Keane <i>et al.</i> 2017). Recently it has been demonstrated that $1\alpha$ , $25(OH)_2D_3$ can act by
104	interaction with the MARRS (Membrane-Associated Rapid Response Steroid) protein that
105	occurs in caveolae together with VDR. This type of $VD_3$ receptor is also known as GRP58
106	(Glucose Responsive Protein, 58 kDa), ERp57 or ERp60 (Endoplasmic Reticulum Protein
107	57/60 kDa) and Pdia3 (Protein Disulfide Isomerase Family A, Member 3) (Hii and Ferrante
108	2016) (Figure 2).

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#### 110 The role of VD<sub>3</sub> in the regulation of folliculogenesis

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A growing body of literature suggests that VD<sub>3</sub> plays an important role in the 112 regulation of ovarian processes that determine female fertility. Female reproductive potential 113 is expressed as the number of primary follicles in the ovary at birth, known as the ovarian 114 115 reserve. It decreases during postnatal life as a result of the recruitment of primary follicles to the growth (Monniaux et al. 2014). This process is controlled by growth factors and 116 hormones, among which the most important is anti-Müllerian hormone (AMH). It is produced 117 by granulosa cells of preantral and early antral follicles and inhibits initial recruitment of 118 follicles, maintaining the ovarian reserve (Visser et al. 2006). Recently the influence of VD<sub>3</sub> 119 on AMH concentration, and thereby on the ovarian reserve, has been extensively discussed. 120 121 Studies conducted on a group of premenopausal women with regular menstrual cycles showed a positive correlation between the plasma concentration of 25(OH)D<sub>3</sub> and AMH. In addition, 122 a decrease in the level of both hormones was observed in the winter and this effect was 123 reversed after VD<sub>3</sub> administration (Merhi *et al.* 2012). The effect of VD<sub>3</sub> on AMH level is 124 probably due to the presence of the VDRE sequence in the AMH gene promoter as found in 125

prostate cells (Malloy et al. 2009). Furthermore, Merhi et al. (2014) has observed that 126 127 25(OH)D<sub>3</sub> deficiency in follicular fluid correlated with an increased expression of the transcript for AMH type II receptor (AMHR-II) in human granulosa cells. VD<sub>3</sub> has also been 128 shown to reduce the phosphorylation of the Smad 1/5/8 protein that contributes to signal 129 transduction from AMHR-II. Thus, VD<sub>3</sub> may increase the synthesis of AMH but also 130 modulate its effect on follicular cells by regulating intracellular signaling pathways (Irani and 131 132 Merhi 2014). Despite the lack of literature data indicating the direct role of VD<sub>3</sub> in maintaining ovarian reserve, the effect exerted on AMH suggests a synergistic action between 133 both hormones. 134

135 The effect of VD<sub>3</sub> on folliculogenesis has been demonstrated for the first time in studies conducted on Vdr- and Cyp27b1-knockout mice. They displayed increased ovarian 136 interstitial tissue, weakened follicular development and lack of corpus luteum suggesting 137 ovulatory disorders (Kinuta et al. 2000, Panda et al. 2001). The influence of VD3 on follicular 138 development *in vitro* has been studied on primates by Xu *et al.* (2016). They isolated preantral 139 140 follicles and cultured them to the antral stage with addition of a low (25 pg/ml) or high (100 pg/ml) concentration of  $1\alpha$ , 25(OH)<sub>2</sub>D<sub>3</sub>. The low dose had a positive effect on oocyte growth, 141 survival and development of preantral follicles, suggested by the authors as being due to an 142 143 increased sensitivity to follicle-stimulating hormone (FSH). However, after reaching the antral stage, the higher dose of  $1\alpha_2 25(OH)_2 D_3$  was more effective and promoted follicular 144 growth (Xu et al. 2016). These results show that VD<sub>3</sub> affects both the early and late stages of 145 146 folliculogenesis, and that its effect is dose-dependent.

147 The growth and development of ovarian follicles are associated with the proliferation 148 and differentiation of granulosa cells. Yao *et al.* (2017) demonstrated an effect of  $VD_3$  on the 149 proliferation of goat granulosa cells by regulation of oxidative stress and changes in the expression of genes regulated cell cycle. The influence of VD<sub>3</sub> on granulosa cell proliferation
has also been observed in hens (Wojtusik and Johnson 2012).

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#### 153 The role of VD<sub>3</sub> in the regulation of steroidogenesis

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Besides production of germ cells, the ovary synthesizes steroid hormones including 155 156 progesterone, androgens and estrogens. Studies so far indicate that VD<sub>3</sub> regulates the 157 expression and activity of steroidogenic enzymes and that the effect is tissue specific (Lundquist 2014). In human granulosa cells, there is an augmented expression and activity of 158 159  $3\beta$ -hydroxysteroid dehydrogenase ( $3\beta$ -HSD) as well as an increase in progesterone production (Merhi et al. 2014), consistent with the studies of Parikh et al. (2010). Studies on porcine 160 granulosa cells *in vitro* revealed no effect of  $1\alpha$ , 25(OH)<sub>2</sub>D<sub>3</sub> on basal progesterone production, 161 162 but noted its increase after insulin and FSH stimulation (Smolikova et al. 2013). In contrast, other studies on porcine granulosa cells showed a reduced progesterone synthesis following 163 1a,25(OH)<sub>2</sub>D<sub>3</sub> treatment that was associated with decreased cholesterol side-chain cleavage 164 enzyme (CYP11A1) mRNA and protein expression and increased 3β-HSD mRNA and 165 protein expression (Hong et al. 2016). Results from experiments on the effect of VD<sub>3</sub> on 166 ovarian estrogen synthesis are clearer. Studies on human (Parikh et al. 2010), porcine (Hong 167 et al. 2017) and goat (Yao et al. 2017) granulosa cells revealed a stimulatory effect of 168  $1\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub> on estradiol and estrone production and on the expression of aromatase 169 (CYP19A1), which converts androgens to estrogens. These results may be explained by the 170 fact that the VDRE element is present in the promoter of the gene encoding CYP19A1 in 171 human placental cells (Sun et al. 1998). The role of VD3 in ovarian androgen synthesis has not 172 yet been intensively studied. Parikh et al. (2010) found no effect of VD<sub>3</sub> on androgen 173 production in humans. However, it has been observed that testosterone affects the 174

transcriptional activity of VDR in porcine granulosa cells by inhibiting the formation of VDRRXR complexes (Herian *et al.* 2018). This information suggests that VD<sub>3</sub> is an important
modulator of steroidogenesis in the human and mammalian ovary.

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179 VD<sub>3</sub> and PCOS

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PCOS is one of the most common endocrinopathies of women of reproductive age. It 181 is characterized by ovulation disorders, irregular cycles, the presence of ovarian cysts, 182 hyperandrogenism, abnormal level of gonadotropins and metabolic disturbances 183 184 (hyperinsulinemia, insulin resistance, dyslipidemia) resulting in infertility. Recent studies show a reduced calcidiol level in women with PCOS suggesting a relationship between VD<sub>3</sub> 185 deficiency and the occurrence of many PCOS symptoms (Dravecka et al. 2016, Shahrokhi et 186 187 al. 2016). VD<sub>3</sub> deficiency is often associated with a disturbed calcium metabolism, which in women with PCOS may inhibit follicle maturation and ovulation. The diminished level of 188 circulating VD<sub>3</sub> also reduces the activity and expression of CYP19A1, which disturbs 189 conversion of androgens to estrogens. An increase in androgen concentration blocks follicular 190 maturation before ovulation and leads to ovarian cyst appearance (Lorenzen et al. 2017). 191 One of the metabolic symptoms occurring in 60-80% of women with PCOS is insulin 192 resistance. VD<sub>3</sub> has been shown to increase insulin synthesis and secretion, and expression of 193 its receptor. In addition, it increases cell sensitivity to insulin by inhibiting the production of 194 pro-inflammatory cytokines (Sung et al. 2012). The direct effect of VD<sub>3</sub> on insulin secretion 195 and consequently on glucose metabolism is mediated via VDR present in  $\beta$  cells of the 196 pancreas. Importantly, VDRE sequence was found in the promoter of the gene coding insulin 197 (Sung et al. 2012). It is also believed that the indirect effect of VD<sub>3</sub> on insulin sensitivity 198 depends on the regulation of intracellular calcium level, which is necessary for proper cell 199

signaling in the insulin-dependent tissues (muscle and fat) (Pittas et al. 2007). Insulin 200 201 resistance results in elevated glucose concentrations, which may in turn modify proteins, lipids and nucleic acids in a non-enzymatic way leading to the formation of Advanced 202 203 Glycation End-products (AGEs). An increased plasma concentration of AGEs as well as their accumulation in granulosa and theca cells of ovarian follicles has been observed in PCOS. 204 205 These compounds bind to their soluble receptor (sRAGE) and induce the formation of reactive oxygen species and cytokines with pro-inflammatory properties. The involvement of 206 207 AGEs and their receptors in the pathogenesis of PCOS is mainly associated with the disturbance of follicular growth. This effect can be attenuated by  $1\alpha$ ,  $25(OH)_2D_3$ , which has 208 209 anti-inflammatory properties (Merhi 2019).

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#### 211 VD<sub>3</sub> and POF

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POF is defined as the loss of ovarian function before the age of 40 years. It is 213 214 characterized by premature depletion of the ovarian reserve due to autoimmune damage or genetic predisposition. Typical POF symptoms include amenorrhea, high gonadotropins and 215 low estradiol levels, as well as decreased AMH concentration in the plasma (Knauff et al. 216 2009). Keeping in mind the effect of VD<sub>3</sub> on the synthesis of AMH as an ovarian reserve 217 marker, VD<sub>3</sub> contribution to the POF etiology seems to be possible. Research conducted on a 218 population of women with POF showed the negative correlation between VD<sub>3</sub> deficiency and 219 FSH level (Kebapcilar et al. 2013). On the other hand, study that confirmed the characteristic 220 221 hormonal profile (high FSH and LH level, low estradiol level) in patients with POF, has also shown no changes in 25(OH)D<sub>3</sub> concentration (Ersoy et al. 2015). The above mentioned 222 results do not allow to unequivocally confirm the role of VD<sub>3</sub> in the pathogenesis of POF. 223 However, it is suggested that VD<sub>3</sub> deficiency may decrease the AMH level, which in turn 224

leads to increased FSH concentration and the

226 occurrence *al*. 2015).

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#### 228 VD<sub>3</sub> and ovarian cancer

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Epidemiological studies have shown that the occurrence of ovarian cancer is inversely 230 correlated with exposure to UVB radiation, which is necessary for the synthesis of VD<sub>3</sub> in the 231 skin, thus suggesting its involvement in the pathogenesis of ovarian cancer (Guo et al. 2018). 232 Analysis of plasma calcidiol concentrations among ovarian cancer patients showed that this 233 234 was significantly lower (under 20 ng/ml) than in the control group. In addition, a group of patients with calcidiol level below 10 ng/ml had a statistically lower survival rate. From the 235 results, it was noticed that VD<sub>3</sub> deficiency may have greater impact on patients with more 236 237 aggressive cancers (Colonese et al. 2015).

It is believed that VDR gene polymorphisms increase the risk of ovarian cancer. The most common is the single FokI nucleotide polymorphism located at the 5 'end, which leads to the synthesis of VDR protein with a longer amino acid sequence. Further identified polymorphisms - BsmI, ApaI and TaqI - are located at the 3 'end and do not affect the synthesis of functional VDR protein but regulate the stability of VDR mRNA (Guo *et al.* 2018).

The anti-tumor mechanism of VD<sub>3</sub> action involves the inhibition of cell proliferation by affecting the cell cycle regulatory proteins (p21, p27, cyclins). In addition, cell cycle inhibition in the G2/M phases and induction of ovarian cancer cell death by increasing mRNA and GADD45 $\alpha$  protein expression were reported. Further research indicates that VD<sub>3</sub> inhibits cancer angiogenesis and metastasis. It has also been reported that it affects glucose and fatty acids metabolism in cancer cells (Guo *et al.* 2018). Previous studies confirm that

250	administration of $VD_3$ or its analogues is not an effective method of ovarian cancer treatment.
251	On the other hand, proper VD <sub>3</sub> supplementation may reduce the risk of illness (Guo <i>et al</i> .
252	2018).
253	
254	Conclusions
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256	Numerous epidemiological data and results of animal studies confirm that VD <sub>3</sub> plays a
257	key role in supporting ovarian function. It has a positive effect on folliculogenesis and
258	maintenance of the ovarian reserve, and also stimulates steroidogenesis. There is growing
259	concern that the global problem with $VD_3$ deficiency among women contributes to
260	reproductive complications. Therefore, $VD_3$ supplementation seems to be a great opportunity
261	for the treatment and insertion of ovarian pathologies. In conclusion, monitoring of plasma
262	calcidiol level should become a preventive diagnostic for ensuring female health.
263	
264	Acknowledgments
265	The websites https://biorender.com (BioRender) and https://smart.servier.com (SMART
266	Sevier Medical Art) and were applied to prepare Figure 1 and Figure 2, respectively.
267	
268	Conflict of Interest
269	There is no conflict of interest to declare.
270	
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### 379 Legend to Figures

**Fig. 1.** Vitamin D<sub>3</sub> contribution to physiological and pathological processes within the ovary.

381 AGEs: advanced glycation end-products; AMH: anti-Müllerian hormone; AMHRII: AMH

receptor type II; 3β-HSD: 3β-hydroxysteroid dehydrogenase; CYP11A1: cholesterol side-

chain cleavage enzyme; CYP19A1: cytochrome P450 aromatase; FSH: follicle-stimulating

hormone; GCs: granulosa cells; LH: luteinizing hormone; PCOS: polycystic ovary syndrome;

POF: premature ovarian failure; TCs: theca cells; VD<sub>3</sub>: vitamin D<sub>3</sub>; VDR: vitamin D<sub>3</sub> receptor

**Fig. 2.** Overview of vitamin D<sub>3</sub> metabolism and intracellular action. CYP2R1: 25-hydroxylase

present in the endoplasmic reticulum; CYP24A1: 24-hydroxylase; CYP27A1: 25-hydroxylase

- present in the mitochondria; CYP27B1: 1α-hydroxylase; MAARS: membrane-associated
- 389 rapid response steroid protein; RXR: 9-cis-retinoic acid receptor; UVB: ultraviolet-B
- irradiation; VDBP: vitamin D<sub>3</sub> binding protein; VDR: vitamin D<sub>3</sub> receptor; VDRE: vitamin D
- 391 response element

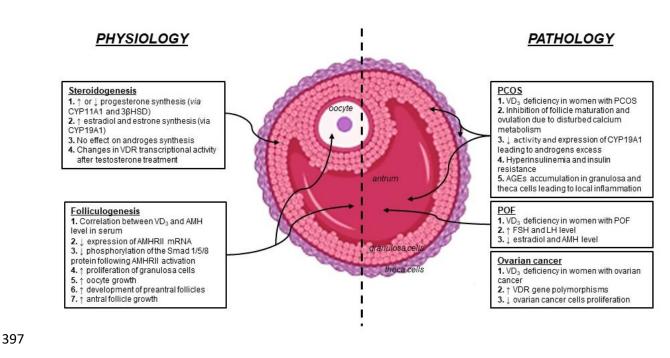
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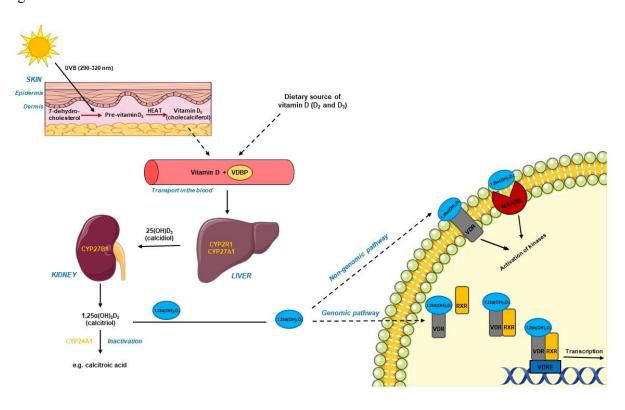
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#### 396 Fig. 1



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399 Fig. 2



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