

1 **Healthy microbiome – a mere idea or a sound concept?**

2

3 Lucie Najmanová¹, Petra Vídeňská², Monika Cahová³

4

5 ORCID IDs

6 Monika Cahova 0000-0003-2640-5084

7 Petra Videnska 0000-0002-7377-7377

8 Lucie Najmanová 0000-0003-0691-7250

9

10 ¹Institute of Microbiology of the CAS, Prague, Czech Republic;

11 ²RECETOX, Faculty of Science Masaryk University, Brno, Czech Republic;

12 ³Institute for Clinical and Experimental Medicine, Prague, Czech Republic;

13

14

15 Corresponding author

16 Monika Cahová, monika.cahova@ikem.cz

17

18 Short title

19 What is a healthy microbiome?

20

21

22

23

24

25

26

27

28

29

30

31

32

33	<i>Content</i>	
34	Content.....	2
35	Holobiont concept.....	4
36	Human-associated microbiome from the ecological perspective	5
37	How to define “being healthy”?.....	7
38	Healthy microbiome in the „one health concept“	7
39	Microbiome-related diseases.....	8
40	Microbiota-focused therapy	9
41	There is nothing like “human healthy microbiome”	10
42	Dynamic character of the microbiome.....	11
43	The variability of human body niches	12
44	Interaction of individual microbiomes/body niches	13
45	The dysbiosis precedes the clinical signs of the disease	14
46	One size does not fit all	15
47	How to describe the microbiome	17
48	Examples of healthy microbiomes	19
49	Vaginal microbiome.....	20
50	Oral microbiome.....	20
51	Summary	21
52		
53		
54		
55		
56		
57		
58		
59		
60		
61		
62		
63		
64		

65 **Abstract**

66 Hundreds of studies in last decades have aimed to compare the microbiome of patients
67 suffering from diverse diseases with that of healthy controls. The microbiome-related
68 component was additionally identified in pathophysiology of many diseases formerly
69 considered to depend only on the host physiology. This, however, opens important
70 questions like: “What is the healthy microbiome?” or “Is it possible to define it
71 unequivocally?”. In this review, we describe the main hindrances complicating the definition
72 of “healthy microbiome” in terms of microbiota composition. We discuss the human
73 microbiome from the perspective of classical ecology and we advocate for the shift from the
74 stress on microbiota composition to the functions that microbiome ensures for the host.
75 Finally, we propose to leave the concept of ideal healthy microbiome and replace it by focus
76 on microbiome advantageous for the host, which always depends on the specific context like
77 the age, genetics, dietary habits, body site or physiological state.

78

79 **Key words**

80 holobiont, core microbiome function, resilience, microbiome ecology, one health
81 hypothesis, dysbiosis

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97 ***Holobiont concept***

98 Progress in sequencing techniques opened new areas of research and revealed that all
99 multicellular organisms, including humans, live in a tight co-existence with rich and highly
100 variable resident microbiota (the bacteria, archaea, viruses, and fungi) that have a significant
101 influence on the host development and health. The term microbiota is, in the literature,
102 often replaced by the term microbiome, which, however, has two other, equally used,
103 distinct definitions. The genetic definition uses this term to describe only the sum of genetic
104 information of the resident microbiota, while the ecological point of view uses the term to
105 describe the microbial community including its habitat with typical physical and chemical
106 conditions, i.e. a dynamic and interactive microecosystem. Additionally, when speaking
107 about microbiota or microbiome, the researchers often have in mind only bacteria, as they
108 represent the most abundant and also most studied portion of microbial population
109 associated with human body. This is also the case of this review.

110 The human associated microbiota (microbiome) is now being recognized as a “new
111 organ” that complements the host’s missing functions. Research focused on the role of
112 microbiota in health and disease or on microbiome-based therapy, opens questions like:
113 “What are the most important characteristics of healthy microbiome?”, “What core
114 functions should it ensure for the host?” and “How could it be described?”. Growing
115 understanding of the complexity of microbial ecosystems and their relationships with their
116 environment unravels that there is probably nothing like one ideal healthy microbiome
117 community.

118 The achievements in the study of the human microbiome shifted the perception of
119 multicellular organisms: they are not only a single entity by themselves, but should be
120 considered as a whole together with a highly variable resident microbiota (the bacteria,
121 archaea, viruses, and fungi), hence the term "holobionts" (1). Both the eukaryotic and
122 prokaryotic components are tightly interconnected and live in a state of dynamic balance.
123 Furthermore, the microbiome component is being continually challenged and replenished by
124 contact with the surrounding environment (**Figure 1**).

125 The holobiont concept brought yet another new term, hologenome, describing
126 collective genomes of the host and its microbiota, where the host (human, animal, plant
127 etc.) genes are only a minority. In the human holobiont, microbial genomes probably
128 outnumber the human genes approx. 100times (2). From this perspective, even a birth event

129 is not only a new human, but also a new community “infant plus its microbiota”(3). Since the
130 start of the Human Microbiome Project, scientists have aimed to characterize the human
131 healthy/beneficial microbiota, however, even after more than a decade there is still no
132 sufficient insight on its nature or how it should behave. Here we summarize the main
133 challenges we face in the field and highlight the most promising approaches.

134

135 ***Human-associated microbiome from the ecological perspective***

136 Microbial communities inhabiting various niches of the human body are communities
137 that meet the criteria of macroecosystems and therefore, it is useful and justifiable to
138 borrow the concepts and methods from classical ecology. The ecosystem consists of all
139 organisms living in a defined area and their interaction with the physical environment. This
140 definition encompasses the complex, adaptive system that is characterized by historical
141 dependency, nonlinear dynamics, threshold effects (i.e. factors promoting the return to the
142 stable state after the disturbance), multiple basins of attraction (i.e. stable states), and
143 limited predictability (4).

144 The behavior of the system could be described using the model of “stability landscape”
145 (5). In this model, the basins of attraction (depressions) represent the stable states. Within
146 the basins, the systems tend to return to equilibrium with the lowest energy. The
147 disturbances, i.e. substantial changes in the environment or community structure and
148 composition, allow the system to pass the threshold and to set in a new stable state (**Figure**
149 **2**). They are invaluable sources of stimuli leading to ecosystem adaptation and evolution if
150 they occur in a predictive manner and manageable scale. On the other hand, if it is
151 unpredictable and erratic, the community would suffer losses and even eventually become
152 extinct (6).

153 Stability, resistance, and resilience are essential characteristics of any ecosystem
154 including the human microbiome (7). According to Pimm, a system is stable if key variables
155 describing the system return to equilibrium values after displacement, the functions of the
156 system are maintained and there is limited variability of key system parameters over time
157 (8). Resistance is defined as the capacity of an ecosystem to remain unchanged on
158 perturbation (9). Ecological resilience was conceptualized by Holing in 1996 and could be
159 defined as a capacity of a system to absorb disturbance and reorganize while undergoing
160 change, so as to still retain essentially the same function, structure, identity, and feedback

161 (10). The combined and often synergistic effects of anthropogenic pressures can make
162 ecosystems less resilient and thus more vulnerable to changes that could have been
163 previously absorbed.

164 In the human microbiome context, the initial state (**Figure 2A**) may represent the stable
165 microbiome of a healthy individual (i.e. advantageous for the host). An intermediate level of
166 disturbance modifies the community composition and its metabolic function, but the
167 microbiome can revert to the original state (**Figure 2B**). If the intensity of the disturbance
168 exceeds the adaptive capacity of the ecosystem, it passes the threshold and reaches a new
169 stable state (**Figure 2C**). The mild disturbance might be a diversified diet or an exposure to a
170 microbial-rich environment. An intensive disturbance could be provoked by the massive use
171 of antibiotics, extensive sanitation, etc. and will push the system to a new stable state,
172 potentially disadvantageous for the host (further referred as unhealthy or dysbiotic).

173 The stable microbial system is intuitively considered healthy and indeed, it probably is –
174 but from its own point of view, i.e. from point of view of the microbiome – not necessarily
175 also the host. For example, in inflammatory bowel disease (IBD) or recurrent *Clostridioides*
176 *difficile* infection, the gut microbiome could also be stable and resilient and as such, it
177 becomes a significant obstacle to therapeutic intervention and contributes to the chronicity
178 of the disease (11, 12). A stable state per se is not a sufficient indicator of a beneficial
179 function, but understanding how stability is established and maintained is essential for
180 diagnosis and successful therapy of many diseases.

181 A key factor for microbiome stability and resilience is the microbial diversity and the
182 consequent functional redundancy. This observation, originally described in grassland
183 savanna ecosystems (13), was repeated in a laboratory “micro-setting”. Naeem and Li
184 performed an experiment on a wide set of artificial microbial communities with a different
185 representation of key functional microbial groups representing terrestrial and aquatic
186 ecosystems and variable amount of available nutrients. They found that the capacity of the
187 system to maintain productivity was dependent on the balanced representation of the
188 number of species per functional group and concluded that the “redundancy is a valuable
189 commodity” (14).

190 These observations resulted in the formulation of the “biological insurance hypothesis”
191 (15) according to which compensation by one species for loss or decline in another preserves

192 long-term average ecosystem performance and reduces variability in performance,
193 promotes the long-term probability of persistence, and enhances resilience to perturbations.
194

195 ***How to define “being healthy”?***

196 Even though the question seems to be simple, the answer is extremely complicated. The
197 first problem represents the term “healthy”. Oxford Dictionary defines health as “the state
198 of being free from illness and injury”. On the opposite end of the scale is WHO definition
199 that describes health as “a state of complete physical, mental and social wellbeing and not
200 merely the absence of disease or infirmity”. Both definitions received substantial criticism.
201 While the former is negative and only excludes the state of illness, the latter is too complex
202 and impossible to measure. Furthermore, the increase in the prevalence of chronic diseases
203 would mean that many people with even minor health complications would be persistently
204 considered as being ill (16, 17). Despite the profound differences between these two
205 definitions, they both share one common feature – they are static.

206 In 1982, Stokes et al. proposed following definition: “Health is a state characterized by
207 anatomical, physiological, and psychological integrity; an ability to perform personally valued
208 family, work, and community roles; an ability to deal with physical, biological, psychological,
209 and social stress.” (18). Interestingly, this definition introduces an important aspect, the
210 ability to cope with stress, which moves the perception of health towards a dynamics
211 process – seeking a balance. From this perspective, health is “a dynamic condition,
212 encompassing resilience to stress and recovery from damage” (16, 17).

213 Human microbiome(s) are very dynamic structures and there is no way to define and
214 describe if they are a priori beneficial or harmful. The concept of dynamic health allows
215 characterizing healthy or pathological microbiomes according to specific conditions.
216 According to the concept of the human holobiont, illness could be related to a non-resilient
217 microbiome unable to meet the physiological demands of the host (19).

218

219 ***Healthy microbiome in the „one health concept“***

220 According to one health concept in its simplified version, it is impossible or at least
221 highly improbable to stay healthy in an unhealthy environment. To understand the holobiont
222 physiology in its complexity we should, therefore, consider not only the two-component

223 system, i.e. the host and its microbiome, and their mutual interactions, but also the
224 holobiont interactions with its environment.

225 The maintenance of a healthy microbiome is critically dependent on the continuous
226 acquisition of microorganisms and appropriate supporting substrates through feeding,
227 drinking, breathing, and other interactions with the environment (20-22). For example, the
228 gut microbiome of hunters and gatherers still surviving in small communities living in
229 relatively pristine areas and in close contact with their natural surroundings is characterized
230 by a higher stability as well as higher diversity when compared to the western population
231 living in urban areas. The gut microbiome diversity of western populations is reduced at all
232 taxonomic levels, meaning that not only species, but also whole large groups, encompassing
233 hundreds of species, are absent. This results in a loss of redundancy and thus of essential
234 functions. The diet common in western societies, characterized by an oversupply of animal
235 protein and fat and a low amount of plant polysaccharides, is associated with a poor capacity
236 to digest carbohydrates (23, 24).

237 Paradoxically, recent advances in medicine and better housing act against the natural
238 self-renewing capacity of our microbiome resulting from the close exposure to the external
239 reservoirs. The massive exposure to antibiotics often results in the depletion of keystone
240 bacterial taxa or whole functional groups called guilds (22, 25, 26). Resulting changes in
241 microbiome composition of citizens of developed countries have been correlated with a low
242 level of resilience, chronic sub-inflammation, and compromised setting of the immune
243 system (27, 28). The “hygiene hypothesis” postulates that reduction in the frequency of
244 infections contributes directly to the increase in the frequency of autoimmune and allergic
245 diseases while the contact with environments rich in microbial diversity protects against
246 these disorders (29-31).

247

248 ***Microbiome-related diseases***

249 The enormously growing microbiome research has important implications in the
250 perception of the mechanisms underlying the onset and development of many NCDs. The
251 way of life in modern, westernized society is profoundly different from the conditions
252 determining the co-evolution of human hosts and their microbiomes. Relatively mild, but
253 long-term influence of conditions like western-type lifestyle with unhealthy diets (32-34),
254 high hygiene standards and extensive usage of cosmetics (35-37), overuse of medicine

255 including antibiotics and proton-pump inhibitors (38), disturbances of the circadian rhythm
256 (39, 40) etc. can cause the deterioration of the human body-associated microbiome
257 ecosystem. In detail, it could be manifested as the loss of key bacterial taxa/guilds, loss or
258 reduction of essential microbiome-mediated functions and metabolites, aberrant
259 stimulation of immune system and compromised control against pathogen attack (19, 22,
260 41). Such changes may belong to the principal drivers of the rise of non-communicable
261 diseases (NCDs) prevalence throughout the last decades (27, 42, 43). The traditional
262 definition of NCDs like asthma, heart disease, obesity, type 2 diabetes, cancer,
263 neurodegenerative conditions or autoimmune diseases rules out microbes as causative
264 agents. Recently the links between NCDs and altered, mainly, but not exclusively, gut
265 microbiome were reported and the therapeutic implications have attracted keen interest
266 among scientists. Several studies suggest that at least in some NCDs there is substantial
267 microbiota-related component and thus they may be to some degree communicable among
268 humans (43, 44). That might as well be the case, but some caution when interpreting the
269 data and translating them into human context is desirable. Many of the disease –
270 microbiota associations are based on correlation studies, i.e. comparison of microbiota in
271 apparently healthy and diseased population. This type of study suffer from two limitations,
272 i.e. (i) correlation does not prove causation (45) and (ii) symptomatically invisible dysbiosis
273 often precedes the disease onset as will be discussed further. The widely used proof-of-
274 concept approach is the transplantation of fecal microbiota from individuals with and
275 without a disease into germ-free animals. The subsequent recapitulation of the diseased
276 phenotype is considered as the proof of causality and was demonstrated for many
277 pathophysiological states, e.g. cardiovascular disease(46), IBD (47), type 2 diabetes (48),
278 obesity (49) and others. Even though the outcomes of these studies are generally accepted,
279 this experimental design has inherent limitations complicating the interpretation of the
280 results (50). The authors definitely do not intend taking the role of microbiome in health and
281 disease into question but it is necessary to keep in mind that oversimplified associations may
282 lead to misinterpretation of experimental results and false identification of specific
283 microbiota composition as “healthy” or “dysbiotic”.

284

285 ***Microbiome-focused therapy***

286 Having in mind the holobiont concept, it seems shortsighted to focus the medical and
287 scientific attention only on the host and his/her physiological processes and to neglect the
288 therapeutic potential of our co-inhabitants. The identification of microbiota-related
289 component in various diseases opens new field of microbiome-focused therapy that may be
290 either untargeted (probiotics, prebiotics, fecal microbiota transfer) or targeted (engineered
291 bacteria, postbiotics, phages) (51). Among these options, the fecal microbiota transfer (FMT)
292 has the greatest potential to induce significant shift in whole gut microbiota community (12)
293 and therefore, to replenish the missing function(s) of the microbiome in complexity.
294 Currently, only recurrent *Clostridioides difficile* infection is approved for FMT therapy in both
295 USA and EU (52) but at this moment, there are 150 clinical trials registered in
296 clinicaltrials.gov investigating FMT therapeutic potential in many other pathological
297 conditions, i.e. IBD, obesity, liver diseases, neurological diseases etc. One of the main
298 challenges that hinder wider application of this otherwise safe and inexpensive therapy in
299 clinical practice is the lack of reliable criteria for the donor. According to the current
300 standards, donors are meticulously tested for potential pathogen presence but the risk of
301 the transmission of more complex “microbiota setting” is still not addressed. Indeed, one
302 case study documented the transmission of an obese phenotype from an overweight donor
303 to a lean patient following FMT for *Clostridioides difficile* infection (52, 53). Particularly from
304 this point of view, the definition of healthy microbiome is of utmost importance. In the
305 following paragraphs, we will discuss the uniqueness of host-microbiome interaction what
306 opens the question whether it is feasible to establish the requirements of one super-donor
307 or whether is better to adjust the requirements for the donor to the needs of specific
308 recipient.

309

310 ***There is nothing like “human healthy microbiome”***

311 It seems a healthy microbiome ensures a better health. However, the fundamental
312 question, how the healthy microbiome should look like, has not been answered. To describe
313 the healthy microbiome, we face several challenges, related to its variability in both time
314 and space: 1) the individual microbiome exhibits both long-term and short-term dynamics. 2)
315 Each body niche harbor a different microbial community adapted to highly variable local
316 conditions. 3) The microbiota communities of different body niches are not separated but
317 interact and influence each other. Therefore, the “unhealthy” state originating in one

318 location may spread to other niches. 4) Since the dysbiosis often precedes the clinical signs
319 of the disease, the microbiome of an apparently healthy individual can already be dysbiotic.
320 5) Usefulness of specific microbiota for the host is context-dependent. Specific microbiota
321 can, depending on other circumstances, represent both the life-saving condition as well as
322 the serious threat.

323

324 Dynamic character of the microbiome

325 The short-term fluctuations are caused for example by a change in the type of the
326 physiological status, circadian rhythms, mechanical stimuli etc. while the long-term
327 variability can result from hormonal shifts or changes connected with aging. The oral
328 microbiome undergoes daily short-term dynamics. The tooth surface and supragingival
329 community is challenged several times a day by teeth-brushing or intake of some foodstuff
330 (e.g. simple sugars) and is naturally restored from other niches in the mouth as well as from
331 the external sources (54). The vaginal microbiome in some women exhibited remarkable
332 variations in time during the menstrual cycle, however for other women it remained
333 relatively stable (55). The skin microbiome is generally considered to be highly stable in time,
334 however, some parts of the foot also exhibited remarkable variability (56).

335 The long-term dynamics of the human microbiome are driven by physiological changes
336 related to ontogenesis and aging (**Figure 3**). In this context, the gut microbiome is probably
337 the most studied one. During the very first days/weeks, the newborn gut microbiome is
338 dominated by aerobic and facultative anaerobic bacteria. As the oxygen content in the gut
339 gradually decreases, obligate anaerobes subsequently prevail. By the age of three years, the
340 distal gut microbiota composition is represented almost entirely by obligate anaerobes (57).
341 After the third year, the gut microbiome becomes less dynamic, however, the stable adult
342 microbiome is established approximately at the end of the second decade of life and
343 persists, again only approximately, up to the age of seventy. Aged microbiome is
344 characterized by a continuous decline in the physiological functions affecting a wide
345 spectrum of metabolic and immunological processes (58) resulting in a chronic pro-
346 inflammatory status called “inflammaging”. Despite significant individual and geographical
347 variability, there are some common features of age-related changes in gut microbiota
348 composition: (i) decreased alpha diversity (59); (ii) increase of potentially pathogenic
349 bacteria, e.g. Streptococcaceae, Staphylococcaceae, and Enterobacteriaceae (60); (iii)

350 reduction of the abundance of potentially beneficial bacteria like *Faecalibacterium*
351 *prausnitzii*, *Roseburia* or *Bifidobacterium* (27, 61). Finally, the changes in microbiota
352 composition are reflected by an altered functional performance, e.g. decreased production
353 of beneficial short-chain fatty acids and increased production of branched-chain fatty acids.
354 In general, aging is associated with a shift from predominantly saccharolytic metabolism
355 towards predominantly putrefactive metabolism in the elderly, with more fermentation of
356 proteins, which concomitantly produces different harmful fermentation metabolites (62, 63)
357

358 The variability of human body niches

359 The multicellular organism is composed of many, often highly variable, niches providing
360 its microbial inhabitants with a wide range of living conditions. The oxygen pressure varies
361 from fully aerobic; e.g. on the skin, to strictly anaerobic conditions; e.g. in the deep
362 periodontal pockets or in the distal gut (cecum and colon). The temperature may be quite
363 stable (~37 °C) in the gastrointestinal (GIT) or urogenital tract or highly variable on the skin
364 surface depending on the environment, activities, and living habits of the host. The pH value
365 can vary from strongly acidic in the stomach (pH=2), mildly acidic on the skin surface
366 (pH=5.5) to more-less neutral in the oral cavity or small intestine. The energy sources vary a
367 lot even throughout the gastrointestinal tract (GIT) and of course, any part of the GIT will
368 provide a more rich and variable source of energy when compared for example to the vagina
369 or scalp.

370 GIT harbors many extremely different microbial communities. In the healthy oral cavity,
371 there are at least four diverse ecological niches: the tongue, buccal mucosa, teeth surface,
372 and gingival crevice, which differ in oxygen and nutrition availability, and saliva flow. In fact,
373 the saliva could be considered another ecological niche. Continuing further through the GIT,
374 the dominant environmental factors affecting the microbiome composition are acidity,
375 oxygen pressure, bile acid composition and nutrient availability (**Figure 4**).

376 The body surface provides variable environments as well. In general, we distinguish dry,
377 moist and oily (sebaceous) areas on the skin and in addition some areas exhibiting
378 topography-related specific features (foot toes), each harboring distinct microbial
379 communities, for review see (56). The oily sites are typically colonized by *Cutibacterium*
380 species while the moist environment of groin or navel is more suitable for
381 *Corynebacteriaceae* and the bottom of heel is dominated by *Staphylococcaceae* (64). This

382 skin microbiome variability is, however, true rather for the skin surface. Bay et al.
383 demonstrated, that the microbiome of the lower dermal layers exhibits lower topologic
384 diversity, is well conserved, and functionally distinct from the epidermal community (65).

385

386 Interaction of individual microbiomes/body niches

387 The microbiota communities separated in space communicate both directly, e.g. by
388 transfer of microbiota and other material including microbial metabolites through the GIT,
389 and indirectly via influencing immune system, neural network, and/or hormones. The most
390 studied model is the oral-gut axis. On average, humans swallow 1.5 liters of saliva containing
391 1.5×10^{12} bacteria per day (66, 67). The oral and gut microbiome seemed to be separated by
392 physical barriers and chemical hurdles like a strongly acidic milieu in the stomach or primary
393 bile acids in the duodenum. Despite these obstacles, however, the presence of oral bacteria
394 has been demonstrated in many body sites (68-72). Live oral bacteria were described not
395 only in lower GIT, but also in the aortic tissue (73), skin (74, 75), atherosclerotic plaques (76),
396 human breast milk (77), brain of Alzheimer-affected patients (78), and healthy placenta (79).
397 For long, the translocation of oral bacteria into lower GIT and other locations was considered
398 to be rare, and it was supposed to be a consequence of the failure of defense mechanisms
399 and hence a hallmark of the disease. Oral bacteria detected in lower GIT have been linked to
400 several pathological states like IBD, colorectal carcinoma (80), pancreatic ductal
401 adenocarcinoma (81) or rheumatoid arthritis (82). In an experiment, *Klebsiella* strains
402 isolated from the saliva of human patients induced IBD in healthy germ-free mice (83).
403 Recently it has been shown at a large scale that despite oral-gut barriers, a substantial part
404 of the oral microbes freely and frequently traverse the GIT and colonize different niches (84).
405 The transmissible bacteria included both pathogenic and commensal oral species (for
406 example *Prevotella* strains or *Fusobacterium nucleatum* subspecies), however, the
407 transmission scores were significantly increased for known opportunistic oral pathogens,
408 causative agents of dental caries, and plaque-dwelling bacteria (84). Endocarditis-associated
409 species (*Haemophilus*, *Aggregatibacter*, *Streptococcus*) exhibited increased transmission
410 scores as well. Taken together, this is an example how the microbiota originating from one
411 niche may modify the composition of distant microbial communities.

412 The mechanism of migration of bacteria to out-of-GIT destination has not been fully
413 elucidated yet but there is a growing body of evidence that alive bacteria could translocate

414 through a leaky intestinal barrier and migrate via the circulation to the distal destinations
415 (57). Alternatively, the oral bacteria could reach the blood circulation system through minor
416 injuries caused by tooth brushing or during dental treatment as formulated in the theory of
417 focal infection reviewed recently by Olsen et al. (85). Furthermore, the direct translocation
418 of bacteria from one niche to another is not the only way how two or more microbial
419 communities communicate and shape each other. At least two other mechanisms are well
420 described, i.e. via the modulation of the host immune system and via bacterial fermentation
421 products released into the circulation (86).

422

423 The dysbiosis precedes the clinical signs of the disease

424 In microbiota-related diseases, the dysbiosis often precedes the clinical signs of the
425 disease (87, 88), and the shift in the microbiome composition could serve as a marker of the
426 risk of the disease development. However, it remarkably challenges the definition of the
427 “healthy microbiome”, because having no clinical signs of the disease does not automatically
428 mean that the microbiome is not already dysbiotic.

429 An example of this phenomenon is the history of an effort to find a microbial signature
430 of colorectal carcinoma. Numerous studies analyzed the gut microbiome in colorectal
431 carcinoma patients with variable outcomes (89). The most consistent result is the increased
432 abundance of *Fusobacterium nucleatum* (90) both in the feces and in mucosa-associated
433 with the tumor. Several other genera were reported to be either elevated
434 (*Peptostreptococcus*, *Streptococcus*, *Porphyromonas*, *Selenomonas*, *Enterococcus*,
435 *Escherichia/Shigella*, *Klebsiella*) or decreased (*Roseburia*, Lachnospiraceae) in colorectal
436 carcinoma patients but the pattern was not uniform (91). This controversy might be
437 explained, at least partly, by the fact that the colorectal carcinoma-associated microbiome is
438 being studied in a situation when the malignant conversion already occurred. For ethical
439 reasons, it is difficult or even impossible to study the colon carcinogenesis “from the
440 beginning” in humans. Therefore, we cannot be sure whether the observed alterations in the
441 microbiome composition is the cause or the consequence of the cancer (92).

442 The driver-passenger model has been proposed by Tjalsma et al. (93) to explain steps
443 leading to malignant conversion of colon epithelium and the role of bacteria in this process
444 (**Figure 5**). In this model, there are bacterial drivers and passengers, which contain bacteria
445 with similar effects (94). Several specific bacteria, the “drivers”, with pro-carcinogenic

446 features initiate colorectal carcinoma development and start the process of malignant
447 transformation of the healthy epithelium into tumor tissue. These key pathogens disappear
448 as they failed to compete with opportunistic bacteria called “bacterial passengers” that are
449 better adapted to the microenvironment of human colorectal carcinoma tumors (95).
450 Therefore, bacterial drivers can be considered as an indicator of a high risk of colorectal
451 carcinoma, while the disappearance of bacterial drivers and the appearance of bacterial
452 passengers may be indicators of the already established colorectal carcinoma (94). So far,
453 several “drivers” and “passengers” species have been proposed. *Helicobacter pylori*,
454 *Enterococcus faecalis*, *Streptococcus bovis/gallolyticus*, and enterotoxigenic strains of
455 *Bacteroides fragilis* are representatives of the “drivers” (96). Bacterial “passengers” are
456 bacteria well-adapted to the tumor microenvironment that in turn produce metabolites
457 favoring the growth of transformed colonocytes. A characteristic feature of “passenger”
458 bacteria is formation of biofilms what substantially increases their viability and provides
459 them competitive advantage over non-aggregated microorganisms. The typical biofilm-
460 forming “passenger” bacteria is *Fusobacterium nucleatum*, which hampers the growth of
461 butyrate-producing bacteria and thus reduces the release of butyrate, one of the main
462 anticancer bacterial metabolites (92).

463 The described example illustrate the motto of this paragraph “dysbiosis precedes the
464 onset of the disease”. In the case of colorectal carcinoma, the first events promoting
465 tumorigenesis occur in the restricted area of the gut and predominantly low-abundant
466 mucosa-associated bacteria are involved. The dysbiosis is local and in the first stages, it is not
467 projected into easily accessible fecal microbiota and the disease is still not overtly
468 manifested. The microbiota associated with fully developed tumor may not be in causative
469 relationship with the disease onset and merely reflects the altered state in the malignant
470 tissue.

471

472 One size does not fit all

473 The vast majority of the microbes inhabiting various human body niches balance
474 between commensalism (one partner benefits while the other is apparently unaffected) and
475 mutualism (co-dependence among symbionts, in which both partners experience increased
476 fitness) (97), some cause harm only under specific circumstances (opportunistic pathogens)
477 and only few are currently considered to be strictly pathogenic. The actual relationship

478 between the particular microorganism and the host depends on many conditions and what
479 is beneficial in one setting may become detrimental in a different context. A growing body of
480 information describing the multifaceted relationship among hosts and their microbial
481 dwellers suggests that mutualism and pathogenicity are two sides of the same coin (22) and
482 the actual interrelationship depends on the context.

483 Here we bring several examples that “one size does not fit all”. In a landmark study,
484 Riquelme et al. showed that pancreatic adenocarcinoma tumors have a specific microbiome
485 (103). This microbiome is derived from the gut microbiota and more importantly, the tumor
486 microbiome composition differs in patients with long- and short-term survival (97). One of
487 the key components of long-term survival tumor microbiota, *Saccharopolyspora*, was
488 implicated in the inflammatory lung disease and was associated with cytokine
489 overproduction (98). The authors suggested that tumor microbiota associated with long-
490 term survival contributes to the anti-tumor immune response by favoring recruitment and
491 activation of CD8+ T cells, i.e. by inducing a pro-inflammatory immune response within the
492 tumor microenvironment. Thus, in the context of pancreatic adenocarcinoma, the pro-
493 inflammatory microbiota pattern, usually and justly considered unhealthy, brings a literally
494 life-saving advantage to the host.

495 Most of the human gut bacteria possess the genetic equipment allowing for
496 fermentation of substrates inaccessible to the host and thus increase the energy extracted
497 from the food - but some strains are more efficient than others. In an elegant series of
498 experiments on mono-colonized mice, Schwarzer et al. demonstrated that *Lactobacillus*
499 *plantarum* promotes juvenile growth and moreover, it buffered the adverse effects of
500 chronic undernutrition (99). Therefore, having these *Lactobacillus* strains in the gut
501 microbiota may represent an advantage if the host faces the risk of malnutrition; however, it
502 is a substantial disadvantage when the energy is in excess.

503 The gut microbiome is being adapted to the prevailing diet and lifestyle of the host. Few
504 studies addressed the gut microbiome of still surviving communities of hunters and
505 gatherers, among them Hadza people living in Tanzania (100). The diet of the Hadza is very
506 rich in diverse plant polysaccharides but low in amino acids. Compared to the urban
507 communities living in Italy or USA, their microbiome is enriched in several bacterial genera
508 including *Prevotella*. *Prevotella* species possess the enzymatic capacity to degrade
509 carbohydrates and have a high capacity for branched-chain amino acid (BCAA) biosynthesis

510 (101, 102). BCAA are essential amino acids that must be supplemented as food or from
511 bacterial metabolism (103). In the natural Hadza environment, *Prevotella* provides their
512 hosts an advantage by increasing their capacity to process a vast array of refractory and
513 resistant plant polysaccharides and supplementing BCAA missing in the diet.

514 At the same time, *Prevotella* may represent a health risk for the people living in urban
515 areas. There is a long-lasting evidence that elevated circulating BCAA associate with insulin
516 resistance, obesity, and diabetes (104) and may even predict cancer development (105). The
517 association between *Prevotella*-rich gut microbiome and insulin resistance was
518 demonstrated. This particular example illustrates how diverse are interactions between the
519 host – microbiome - environment. In one setting, the metabolic equipment (BCAA
520 biosynthesis) may represent either an evolutionary advantage (in case of low availability of
521 animal proteins) or a risk factor (in the situation of protein overnutrition). High fibrinolytic
522 capability may be of utmost importance (when most of the calories are obtained from plant
523 polysaccharides not easily accessible to humans) or negligible factor (when fiber in the diet is
524 rare).

525 Other examples can be found in oral microbiome studies. When comparing healthy
526 community with periodontitis patients, in almost every sufficiently big cohort there are few
527 outliers of both types – clinically healthy individuals with a clearly dysbiotic microbiome and
528 on the other hand severely affected patients with “healthy microbiomes” (88). The authors
529 hypothesize that some individuals possess an over-reactive immune system that triggers the
530 proinflammatory reaction to otherwise symbiotic bacteria while subject with a less reactive
531 immune system are more tolerant to pathogens. Several other examples of situations when
532 people do not develop the same level of oral disease under the same circumstances are
533 discussed in the review by Rosier et al. (54).

534

535 ***How to describe the microbiome***

536 As mentioned above, the microbiome is a complex and dynamic structure and the
537 choice of appropriate measures is a challenging task. We can ask about its taxonomic
538 composition (“Who is present? How abundant is each component?”), about the functional
539 potential (“What are the consortium members able to do?”), about their actual metabolic
540 performance (“What are they doing just now?”) or how is the community stable or
541 vulnerable.

542 The taxonomic composition could be addressed in principle by two approaches, 16S
543 rRNA gene sequencing or shotgun metagenomic sequencing (WMS) each of them answering
544 a somewhat different question. 16S rRNA gene sequencing provides, rapidly and for
545 relatively low cost, information about taxonomic composition with limited precision and
546 depth of identification. WMS informs us not only about the presence of individual taxa but
547 also about the metabolic potential of the community, i.e. the presence of respective marker
548 genes representing metabolic pathways, however, for the sake of higher costs and
549 requirements for advanced computational skills (106). An alternative approach is RNA
550 sequencing which is similar to WMS in the principle, just instead of the microbial DNA,
551 mRNA serves as a template. RNA sequencing identifies only genes that are actively
552 transcribed at the time of sampling, i.e. it takes into consideration only the alive
553 microorganisms and informs about their functional profile (106).

554 Most bacteria possess a wide metabolic repertoire and individual metabolic pathways
555 could be easily switched on and off to maximize the energy yield from the available
556 substrate(s). Therefore, the same bacteria are capable to produce a very different spectrum
557 of metabolites. The simple list of bacteria present in a sample or the metagenomic analysis
558 including a list of encoded enzymes/metabolic pathways thus provide only partial
559 information about the actual state of the studied community. In contrast, the metabolome
560 has been proposed as a functional read-out of the human microbiome (107), reflective of
561 microbiome–host interactions with an immediate impact on host health. Metabolomics
562 identifies already biosynthesized metabolites/small molecules and therefore provides
563 reliable information about the performance of the microbiota as a whole. On the other
564 hand, we cannot assign particular metabolites to specific members of the consortium or to
565 the host, and there are several other technological biases: There are two main approaches
566 to metabolome analysis – targeted and untargeted. The targeted analysis focused on the
567 preselected group of metabolites ensures the high reproducibility and accuracy of the
568 outcome but the obtained information is limited to a narrow spectrum of compounds. The
569 untargeted analysis aims to identify as many compounds as possible allowing for the
570 elimination of selection bias. At the same time, this approach faces several limitations. First,
571 the identification of hundreds of compounds is laborious, time-consuming, and sometimes
572 impossible. Second, the selection of the sample processing and separation methods always
573 limits the outputs only to part of the present metabolites. Third, the quantification of the

574 obtained signals is complicated and usually, the quantity of a particular compound could be
575 expressed only as a portion of the total, i.e. in percent, but not in absolute concentrations.

576 All the above-mentioned methodological approaches – metagenomics,
577 metatranscriptomics, metabolomics – share one common feature, they produce a huge
578 amount of data. The enormous technological development somewhat outruns our tools and
579 ability to understand, visualize and interpret this reality and seriously complicates the
580 integration of outcomes from different studies. The complexity of the microbiome systems
581 impose enormous demands on the whole research pipeline what results in the
582 reproducibility crisis (108). Searching for the roots of this problem numerous studies were
583 undertaken and unraveled that the chosen method significantly influence the outcome at
584 virtually each step of the experimental procedure – from sample collection and DNA
585 extraction (109) , library preparation (110) to the bioinformatics pipeline (111) and data
586 handling method (112). In response to this challenge, guidelines for “wet lab procedures”
587 (MBQC project, IHMS project) were established (110, 113). Standard guidelines tailored to
588 microbiome study reporting called STORMS (“Strengthening The Organizing and Reporting of
589 Microbiome Studies”) checklist were developed by a consortium of multidisciplinary
590 specialists (114). STORMS provides a tool to organize study planning and manuscript
591 preparation, to improve the clarity of manuscripts, and to facilitate reviewers and readers in
592 assessing these studies. Unfortunately, there is no general consensus on how to handle
593 omics data on bioinformatics level so far and several approaches exist, all of them having
594 their plus and cons (115). The authors would like to stress that the selection of
595 bioinformatics method and biostatistical approach always determines the outcome. At
596 present, the only solution of this bottleneck is the openness in sharing the original
597 sequencing data with sufficient metadata allowing for their re-analysis.

598 So far, we addressed only the cross-sectional description of the microbial community,
599 i.e. “here and now”. Aiming to the description of a healthy microbiome, whatever it is, the
600 more important issue is the assessment of sustainability measures of the microbial
601 ecosystem, its stability, resistance, and resilience. Unfortunately, this field is still at the very
602 beginning and the development of new methodological approaches is highly needed.

603

604 ***Examples of healthy microbiomes***

605 All the above-mentioned facts make the postulation of the healthy microbiome of a
606 specific human body site uncertain and complicated. Nevertheless, in few cases the
607 scientists succeeded at least to describe the taxa as generally beneficial for their hosts,
608 which thus could be considered a healthy microbiome of the niche.

609

610 Vaginal microbiome

611 The best example could be a relatively simple vaginal microbiome (116). During the
612 reproductive age, it is mainly dominated by *Lactobacillus* sp. which metabolites keep low pH
613 protecting thus the genital tract and fetus from pathogenic microorganisms, for review see
614 (117). It can be affected by ethnicity (118), age, and hormonal state - negligibly by menstrual
615 cycle (55) but remarkably during puberty and pregnancy (119). In pregnancy, the species
616 richness generally decreases (116) but the alpha diversity depends on the gestation week
617 and could serve even as a predictive marker of the pre-term delivery risk (120). The human
618 vaginal microbiota is generally assigned to several vaginotypes or community state types
619 (CSTs), first described by Ravel et al. (121), but following scientific papers in the field differ in
620 the number of identified CSTs as well as in their characterization, which is always dependent
621 on the clustering analysis of the entire evaluated sample set.

622 Nevertheless, we can conclude, that the vaginal microbiome of healthy adult women is
623 predominantly composed of one or more *Lactobacillus* sp. and that some small percentage
624 of women harbor a mixed population of non-*Lactobacillus* species based on *Gardnerella*
625 *vaginalis*, *Prevotella*, *Atopobium*, *Klebsiella* and others. The *Lactobacillus* sp.-based CSTs are
626 considered beneficial (keeping low pH and producing metabolites protective against
627 urogenital infections) while the mixed *Gardnerella*-based CST can indicate the risk of
628 bacterial vaginosis. Among *Lactobacillus*-based CSTs, the predominance of *L. crispatus* in
629 pregnancy is considered protective against the risk of preterm delivery, while the *L. inners*
630 seems to indicate an increased risk of prematurity as well as the mixed *Gardnerella* based
631 CST.

632

633 Oral microbiome

634 A much more complicated situation is in the oral cavity. After the gut, the oral cavity has
635 the second largest and diverse microbiota (122). It even gained its own database HOMD
636 (Human oral microbiome database; <https://www.homd.org/>) harboring currently 774 oral

637 bacterial species, 26% of them being known only as uncultivated phylotypes (123). Last, but
638 not least, in the majority of scientific studies employing 16S rDNA-based taxonomy and
639 clustering analysis comparing variable healthy and diseased groups, there are outliers, i.e.
640 clearly diseased patients with the seemingly “healthy” microbiome and vice versa. For all
641 these reasons, the estimation of the healthy oral microbiome is extremely tricky and it is
642 clear, that “one size does not fit all”. Nevertheless, the current state of knowledge enables
643 us to define at least some characteristics of beneficial oral microbiome of Caucasian
644 individuals living in developed countries.

645 The healthy oral microbiome is generally based on variable species of *Streptococcus*,
646 mainly *S. mitis*, *S. oralis*, *S. gordonii*, *S. sanguinis* or *S. parasanguinis* (*S. mutans* is associated
647 with dental caries so it cannot be considered beneficial); further various *Haemophilus*
648 species, *Neisseria*, *Rothia*, *Gemella*, *Lautropia* and probably also *Veillonella* (88), which, as an
649 anaerobic microorganism, could be considered a transient taxon on the way to dysbiosis.
650 Such oral microbiome often comprises also *Fusobacterium nucleatum*, which cannot be
651 considered beneficial but its low percentage in oral cavity probably does not cause any harm
652 (however, its presence in GIT is associated e.g. with increased risk of colon cancer) (81, 92).

653 However, when these more-less aerobic species are gradually replaced by *F. nucleatum*,
654 *Porphyromonas* sp. like *P. pasteri* and *P. catoniae*, and *Capnocytophaga* sp., the
655 microenvironment becomes more suitable for true periopathogenic taxa like red-complex
656 bacteria *Porphyromonas gingivalis*, *Tannerella forsythia*, *Treponema denticola*,
657 *Fretibacterium* sp. and *Filifactor alocis*. Such oral microbiome is considered dysbiotic and the
658 respective individual is at a high risk of developing periodontal disease or is already
659 symptomatic (88). The interplay between the oral microbiome and immune system of the
660 host is highly individual and the clear definition of the level of dysbiosis already critical for
661 the development of the disease is not available. The tools enabling the evaluation of the
662 dysbiosis based on the taxonomic composition of the oral microbiome, thus can be used to
663 place the patient in question on a scale from health to the disease based on comparison with
664 a database of already diagnosed individuals, however, it is only based on the statistic
665 probability and there always would be some individuals misclassified (124).

666

667 **Summary**

668 An overwhelming amount of evidence proves that the human microbiome fully deserves
669 to be considered an additional organ of the human body. Unfortunately, we still lack the
670 appropriate measures allowing for the objective evaluation of whether the individual
671 microbiome is healthy or not. Even the term “healthy” is misleading. It would be more
672 appropriate to assess whether the microbiome composition and performance are
673 (dis)advantageous for the host. The suitability of the particular microbiome composition for
674 the host is always dynamic and depends on the situation of the host and the conditions of
675 the environment; therefore, it is impossible to define one idealized community of specific
676 microbes. The more promising approach may be to concentrate our effort on the definition
677 of the essential (core) set of functions and metabolic modules that a healthy holobiont
678 should possess – no matter if provided by its prokaryotic or eukaryotic part. Their absence
679 could be predictive of the disease onset, especially in cases when the dysbiosis precedes the
680 manifestation of the clinical symptoms. The therapeutic interventions should rather be
681 focused on the replenishment of the attenuated/missing functions of the microbiome than
682 on the simple provision of selected probiotic strains.

683 Furthermore, one of the key characteristics of a healthy microbiome is its resilience, i.e.
684 the ability to maintain the necessary function in the changing environment even when it
685 means the reorganization and changes in the composition of the community. The
686 disturbances imposed on the human microbiome ecosystem are in most cases inevitable.
687 Our efforts to reduce the resulting undesired shifts in the microbiome structure should
688 preferentially address and strengthen the resilience rather than try to achieve some ideal
689 composition.

690 Finally, to our opinion, the human microbiome must be envisioned as a complex system
691 tightly interconnected with other macro- and micro-ecosystems in our environment. Our, i.e.
692 human, microbiome cannot stay healthy in an otherwise unhealthy environment, and
693 therefore, it is essential to pay similar attention to all components of the planetary
694 ecosystem.

695

696

697 **Acknowledgement**

698 This study was supported by the Ministry of Health of the Czech Republic, grant NV-18-
699 01-00040, by the project National Institute of virology and bacteriology (Programme

700 EXCELES, ID Project No. LX22NPO5103) - Funded by the European Union - Next Generation
701 EU and by the project National Institute for Research of Metabolic and Cardiovascular
702 Diseases (Programme EXCELES, Project No. LX22NPO5104) - Funded by the European Union -
703 Next Generation EU.

704

705 **References**

- 706 1. Gordon J, Knowlton, N., Relman, D. A., Rohwer, F., & Youle, M. Superorganisms and
707 holobionts. *Microbe*. 2013;8:152-3.
- 708 2. Human Microbiome Project C. Structure, function and diversity of the healthy human
709 microbiome. *Nature*. 2012;486(7402):207-14.
- 710 3. Gilbert SF. A holobiont birth narrative: the epigenetic transmission of the human
711 microbiome. *Front Genet*. 2014;5:282.
- 712 4. Polasky S, Levin, Simon A. *Fragile Dominion: Complexity and the Commons*. Reading MA:
713 Perseus Books, 1999, 254 pp., \$@@-@@27.00. *American Journal of Agricultural Economics*.
714 2001;83(1):246-7.
- 715 5. Carl F, Steve C, Brian W, Marten S, Thomas E, Lance G, et al. Regime Shifts, Resilience, and
716 Biodiversity in Ecosystem Management. *Annual Review of Ecology, Evolution, and Systematics*.
717 2004;35(1):557-81.
- 718 6. Relman DA. The human microbiome: ecosystem resilience and health. *Nutr Rev*. 2012;70
719 Suppl 1:S2-9.
- 720 7. Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience
721 of the human gut microbiota. *Nature*. 2012;489(7415):220-30.
- 722 8. Pimm SL. *The Balance of Nature?: Ecological Issues in the Conservation of Species and*
723 *Communities*: University of Chicago Press; 1991.
- 724 9. Sommer F, Anderson JM, Bharti R, Raes J, Rosenstiel P. The resilience of the intestinal
725 microbiota influences health and disease. *Nat Rev Microbiol*. 2017;15(10):630-8.
- 726 10. Holling CS. Resilience and Stability of Ecological Systems. *Annual Review of Ecology and*
727 *Systematics*. 1973;4(1):1-23.
- 728 11. Moustafa A, Li W, Anderson EL, Wong EHM, Dulai PS, Sandborn WJ, et al. Genetic risk,
729 dysbiosis, and treatment stratification using host genome and gut microbiome in inflammatory
730 bowel disease. *Clin Transl Gastroenterol*. 2018;9(1):e132.
- 731 12. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, et al. Duodenal
732 infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med*. 2013;368(5):407-15.
- 733 13. McNaughton SJ. Diversity and Stability of Ecological Communities: A Comment on the Role of
734 Empiricism in Ecology. *The American Naturalist*. 1977;111(979):515-25.
- 735 14. Naeem S, Li S. Biodiversity enhances ecosystem reliability. *Nature*. 1997;390(6659):507-9.
- 736 15. Yachi S, Loreau M. Biodiversity and ecosystem productivity in a fluctuating environment: the
737 insurance hypothesis. *Proc Natl Acad Sci U S A*. 1999;96(4):1463-8.
- 738 16. Huber M, Knottnerus JA, Green L, van der Horst H, Jadad AR, Kromhout D, et al. How should
739 we define health? *BMJ*. 2011;343:d4163.
- 740 17. Oleribe OO, Ukwedeh O, Burstow NJ, Gomaa AI, Sonderup MW, Cook N, et al. Health:
741 redefined. *Pan Afr Med J*. 2018;30:292.
- 742 18. Stokes J, 3rd, Noren J, Shindell S. Definition of terms and concepts applicable to clinical
743 preventive medicine. *J Community Health*. 1982;8(1):33-41.
- 744 19. Dietert RR. Microbiome First Medicine in Health and Safety. *Biomedicines*. 2021;9(9).
- 745 20. David LA, Materna AC, Friedman J, Campos-Baptista MI, Blackburn MC, Perrotta A, et al. Host
746 lifestyle affects human microbiota on daily timescales. *Genome Biol*. 2014;15(7):R89.

- 747 21. Dethlefsen L, McFall-Ngai M, Relman DA. An ecological and evolutionary perspective on
748 human-microbe mutualism and disease. *Nature*. 2007;449(7164):811-8.
- 749 22. Larsen OFA, van de Burgwal LHM. On the Verge of a Catastrophic Collapse? The Need for a
750 Multi-Ecosystem Approach to Microbiome Studies. *Front Microbiol*. 2021;12:784797.
- 751 23. De Filippo C, Cavalieri D, Di Paola M, Ramazzotti M, Poullet JB, Massart S, et al. Impact of diet
752 in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa.
753 *Proc Natl Acad Sci U S A*. 2010;107(33):14691-6.
- 754 24. Yatsunenkov T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, et al. Human
755 gut microbiome viewed across age and geography. *Nature*. 2012;486(7402):222-7.
- 756 25. Banerjee S, Schlaeppi K, van der Heijden MGA. Keystone taxa as drivers of microbiome
757 structure and functioning. *Nat Rev Microbiol*. 2018;16(9):567-76.
- 758 26. Bach JF. Revisiting the Hygiene Hypothesis in the Context of Autoimmunity. *Front Immunol*.
759 2020;11:615192.
- 760 27. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota
761 composition correlates with diet and health in the elderly. *Nature*. 2012;488(7410):178-84.
- 762 28. Voorhies AAL, H. A. The Challenge of Maintaining a Healthy Microbiome during Long-
763 Duration Space Missions. *Front Astron Space Sci*. 2016;3(23):1-7.
- 764 29. Bach JF. The hygiene hypothesis in autoimmunity: the role of pathogens and commensals.
765 *Nat Rev Immunol*. 2018;18(2):105-20.
- 766 30. Rook GA. Review series on helminths, immune modulation and the hygiene hypothesis: the
767 broader implications of the hygiene hypothesis. *Immunology*. 2009;126(1):3-11.
- 768 31. von Hertzen L, Hanski I, Haahtela T. Natural immunity. Biodiversity loss and inflammatory
769 diseases are two global megatrends that might be related. *EMBO Rep*. 2011;12(11):1089-93.
- 770 32. Kostovcikova K, Coufal S, Galanova N, Fajstova A, Hudcovic T, Kostovcik M, et al. Diet Rich in
771 Animal Protein Promotes Pro-inflammatory Macrophage Response and Exacerbates Colitis in Mice.
772 *Front Immunol*. 2019;10:919.
- 773 33. Zhang X, Dong Y, Sun G, Hasan AA, Tian M, Zeng S, et al. Paternal Programming of Liver
774 Function and Lipid Profile Induced by a Paternal Pre-Conceptional Unhealthy Diet: Potential
775 Association with Altered Gut Microbiome Composition. *Kidney Blood Press Res*. 2019;44(1):133-48.
- 776 34. Sun CY, Zheng ZL, Chen CW, Lu BW, Liu D. Targeting Gut Microbiota With Natural
777 Polysaccharides: Effective Interventions Against High-Fat Diet-Induced Metabolic Diseases. *Front*
778 *Microbiol*. 2022;13:859206.
- 779 35. Panelli S, Epis S, Cococcioni L, Perini M, Paroni M, Bandi C, et al. Inflammatory bowel
780 diseases, the hygiene hypothesis and the other side of the microbiota: Parasites and fungi.
781 *Pharmacol Res*. 2020;159:104962.
- 782 36. Fyhrquist N. The Human Microbiota and Its Relationship with Allergies. *Gastroenterol Clin*
783 *North Am*. 2019;48(3):377-87.
- 784 37. Olunoiki E, Rehner J, Bischoff M, Koshel E, Vogt T, Reichrath J, et al. Characteristics of the Skin
785 Microbiome in Selected Dermatological Conditions: A Narrative Review. *Life (Basel)*. 2022;12(9).
- 786 38. Tramper-Stranders G, Ambrozej D, Arcolaci A, Atanaskovic-Markovic M, Boccabella C, Bonini
787 M, et al. Dangerous liaisons: Bacteria, antimicrobial therapies, and allergic diseases. *Allergy*.
788 2021;76(11):3276-91.
- 789 39. Matenchuk BA, Mandhane PJ, Kozyrskyj AL. Sleep, circadian rhythm, and gut microbiota.
790 *Sleep Med Rev*. 2020;53:101340.
- 791 40. Frazier K, Chang EB. Intersection of the Gut Microbiome and Circadian Rhythms in
792 Metabolism. *Trends Endocrinol Metab*. 2020;31(1):25-36.
- 793 41. Pickard JM, Zeng MY, Caruso R, Nunez G. Gut microbiota: Role in pathogen colonization,
794 immune responses, and inflammatory disease. *Immunol Rev*. 2017;279(1):70-89.
- 795 42. Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, et al. Gut microbiota from twins
796 discordant for obesity modulate metabolism in mice. *Science*. 2013;341(6150):1241214.
- 797 43. Datar A, Nicosia N. Assessing Social Contagion in Body Mass Index, Overweight, and Obesity
798 Using a Natural Experiment. *JAMA Pediatr*. 2018;172(3):239-46.

799 44. Christakis NA, Fowler JH. The spread of obesity in a large social network over 32 years. *N Engl*
800 *J Med.* 2007;357(4):370-9.

801 45. Cani PD. Human gut microbiome: hopes, threats and promises. *Gut.* 2018;67(9):1716-25.

802 46. Gregory JC, Buffa JA, Org E, Wang Z, Levison BS, Zhu W, et al. Transmission of atherosclerosis
803 susceptibility with gut microbial transplantation. *J Biol Chem.* 2015;290(9):5647-60.

804 47. Elinav E, Strowig T, Kau AL, Henao-Mejia J, Thaiss CA, Booth CJ, et al. NLRP6 inflammasome
805 regulates colonic microbial ecology and risk for colitis. *Cell.* 2011;145(5):745-57.

806 48. Thaiss CA, Itav S, Rothschild D, Meijer MT, Levy M, Moresi C, et al. Persistent microbiome
807 alterations modulate the rate of post-dieting weight regain. *Nature.* 2016;540(7634):544-51.

808 49. Turnbaugh PJ, Ridaura VK, Faith JJ, Rey FE, Knight R, Gordon JI. The effect of diet on the
809 human gut microbiome: a metagenomic analysis in humanized gnotobiotic mice. *Sci Transl Med.*
810 2009;1(6):6ra14.

811 50. Walter J, Armet AM, Finlay BB, Shanahan F. Establishing or Exaggerating Causality for the Gut
812 Microbiome: Lessons from Human Microbiota-Associated Rodents. *Cell.* 2020;180(2):221-32.

813 51. Bajaj JS, Ng SC, Schnabl B. Promises of microbiome-based therapies. *J Hepatol.*
814 2022;76(6):1379-91.

815 52. Marotz CA, Zarrinpar A. Treating Obesity and Metabolic Syndrome with Fecal Microbiota
816 Transplantation. *Yale J Biol Med.* 2016;89(3):383-8.

817 53. Alang N, Kelly CR. Weight gain after fecal microbiota transplantation. *Open Forum Infect Dis.*
818 2015;2(1):ofv004.

819 54. Rosier BT, Marsh PD, Mira A. Resilience of the Oral Microbiota in Health: Mechanisms That
820 Prevent Dysbiosis. *J Dent Res.* 2018;97(4):371-80.

821 55. Chaban B, Links MG, Jayaprakash TP, Wagner EC, Bourque DK, Lohn Z, et al. Characterization
822 of the vaginal microbiota of healthy Canadian women through the menstrual cycle. *Microbiome.*
823 2014;2:23.

824 56. Oh J, Byrd AL, Deming C, Conlan S, Program NCS, Kong HH, et al. Biogeography and
825 individuality shape function in the human skin metagenome. *Nature.* 2014;514(7520):59-64.

826 57. Nagpal R, Yadav H. Bacterial Translocation from the Gut to the Distant Organs: An Overview.
827 *Ann Nutr Metab.* 2017;71 Suppl 1:11-6.

828 58. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell.*
829 2013;153(6):1194-217.

830 59. O'Toole PW, Jeffery IB. Gut microbiota and aging. *Science.* 2015;350(6265):1214-5.

831 60. Odamaki T, Kato K, Sugahara H, Hashikura N, Takahashi S, Xiao JZ, et al. Age-related changes
832 in gut microbiota composition from newborn to centenarian: a cross-sectional study. *BMC Microbiol.*
833 2016;16:90.

834 61. Salazar N, Gonzalez S, Nogacka AM, Rios-Covian D, Arboleya S, Gueimonde M, et al.
835 *Microbiome: Effects of Ageing and Diet.* *Curr Issues Mol Biol.* 2020;36:33-62.

836 62. Nagpal R, Mainali R, Ahmadi S, Wang S, Singh R, Kavanagh K, et al. Gut microbiome and
837 aging: Physiological and mechanistic insights. *Nutr Healthy Aging.* 2018;4(4):267-85.

838 63. Rios-Covian D, Gonzalez S, Nogacka AM, Arboleya S, Salazar N, Gueimonde M, et al. An
839 Overview on Fecal Branched Short-Chain Fatty Acids Along Human Life and as Related With Body
840 Mass Index: Associated Dietary and Anthropometric Factors. *Front Microbiol.* 2020;11:973.

841 64. Trivedi B. Microbiome: The surface brigade. *Nature.* 2012;492(7429):S60-1.

842 65. Bay L, Barnes CJ, Fritz BG, Thorsen J, Restrup MEM, Rasmussen L, et al. Universal Dermal
843 Microbiome in Human Skin. *mBio.* 2020;11(1).

844 66. Humphrey SP, Williamson RT. A review of saliva: normal composition, flow, and function. *J*
845 *Prosthet Dent.* 2001;85(2):162-9.

846 67. Sender R, Fuchs S, Milo R. Revised Estimates for the Number of Human and Bacteria Cells in
847 the Body. *PLoS Biol.* 2016;14(8):e1002533.

848 68. Berthelot JM, Bandiaky ON, Le Goff B, Amador G, Chaux AG, Soueidan A, et al. Another Look
849 at the Contribution of Oral Microbiota to the Pathogenesis of Rheumatoid Arthritis: A Narrative
850 Review. *Microorganisms.* 2021;10(1).

851 69. Khan I, Khan I, Jianye Z, Xiaohua Z, Khan M, Hilal MG, et al. Exploring blood microbial
852 communities and their influence on human cardiovascular disease. *J Clin Lab Anal.*
853 2022;36(4):e24354.

854 70. Martinez M, Postolache TT, Garcia-Bueno B, Leza JC, Figuero E, Lowry CA, et al. The Role of
855 the Oral Microbiota Related to Periodontal Diseases in Anxiety, Mood and Trauma- and Stress-
856 Related Disorders. *Front Psychiatry.* 2021;12:814177.

857 71. Newman KL, Kamada N. Pathogenic associations between oral and gastrointestinal diseases.
858 *Trends Mol Med.* 2022.

859 72. Sumida K, Han Z, Chiu CY, Mims TS, Bajwa A, Demmer RT, et al. Circulating Microbiota in
860 Cardiometabolic Disease. *Front Cell Infect Microbiol.* 2022;12:892232.

861 73. Hajishengallis G. Periodontitis: from microbial immune subversion to systemic inflammation.
862 *Nat Rev Immunol.* 2015;15(1):30-44.

863 74. Salem I, Ramser A, Isham N, Ghannoum MA. The Gut Microbiome as a Major Regulator of the
864 Gut-Skin Axis. *Front Microbiol.* 2018;9:1459.

865 75. Sanchez-Pellicer P, Navarro-Moratalla L, Nunez-Delegido E, Ruzafa-Costas B, Aguera-Santos J,
866 Navarro-Lopez V. Acne, Microbiome, and Probiotics: The Gut-Skin Axis. *Microorganisms.* 2022;10(7).

867 76. Chhibber-Goel J, Singhal V, Bhowmik D, Vivek R, Parakh N, Bhargava B, et al. Linkages
868 between oral commensal bacteria and atherosclerotic plaques in coronary artery disease patients.
869 *NPJ Biofilms Microbiomes.* 2016;2:7.

870 77. Williams JE, Carrothers JM, Lackey KA, Beatty NF, Brooker SL, Peterson HK, et al. Strong
871 Multivariate Relations Exist Among Milk, Oral, and Fecal Microbiomes in Mother-Infant Dyads During
872 the First Six Months Postpartum. *J Nutr.* 2019;149(6):902-14.

873 78. Poole S, Singhrai SK, Kesavalu L, Curtis MA, Crean S. Determining the presence of
874 periodontopathic virulence factors in short-term postmortem Alzheimer's disease brain tissue. *J*
875 *Alzheimers Dis.* 2013;36(4):665-77.

876 79. Xu B, Han YW. Oral bacteria, oral health, and adverse pregnancy outcomes. *Periodontol*
877 *2000.* 2022;89(1):181-9.

878 80. Nakatsu G, Li X, Zhou H, Sheng J, Wong SH, Wu WK, et al. Gut mucosal microbiome across
879 stages of colorectal carcinogenesis. *Nat Commun.* 2015;6:8727.

880 81. Mitsuhashi K, Noshio K, Sukawa Y, Matsunaga Y, Ito M, Kurihara H, et al. Association of
881 *Fusobacterium* species in pancreatic cancer tissues with molecular features and prognosis.
882 *Oncotarget.* 2015;6(9):7209-20.

883 82. Zhang X, Zhang D, Jia H, Feng Q, Wang D, Liang D, et al. The oral and gut microbiomes are
884 perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat Med.* 2015;21(8):895-
885 905.

886 83. Atarashi K, Suda W, Luo C, Kawaguchi T, Motoo I, Narushima S, et al. Ectopic colonization of
887 oral bacteria in the intestine drives TH1 cell induction and inflammation. *Science.*
888 2017;358(6361):359-65.

889 84. Schmidt TS, Hayward MR, Coelho LP, Li SS, Costea PI, Voigt AY, et al. Extensive transmission
890 of microbes along the gastrointestinal tract. *Elife.* 2019;8.

891 85. Olsen I, van Winkelhoff AJ. Acute focal infections of dental origin. *Periodontol* 2000.
892 2014;65(1):178-89.

893 86. Park SY, Hwang BO, Lim M, Ok SH, Lee SK, Chun KS, et al. Oral-Gut Microbiome Axis in
894 Gastrointestinal Disease and Cancer. *Cancers (Basel).* 2021;13(9).

895 87. Liu B, Faller LL, Klitgord N, Mazumdar V, Ghodsi M, Sommer DD, et al. Deep sequencing of
896 the oral microbiome reveals signatures of periodontal disease. *PLoS One.* 2012;7(6):e37919.

897 88. Lenartova M, Tesinska B, Janatova T, Hrebicek O, Mysak J, Janata J, et al. The Oral
898 Microbiome in Periodontal Health. *Front Cell Infect Microbiol.* 2021;11:629723.

899 89. Jahani-Sherafat S, Alebouyeh M, Moghim S, Ahmadi Amoli H, Ghasemian-Safaei H. Role of
900 gut microbiota in the pathogenesis of colorectal cancer; a review article. *Gastroenterol Hepatol Bed*
901 *Bench.* 2018;11(2):101-9.

902 90. Lawrence GW, Begley M, Cotter PD, Guinane CM. Potential Use of Biotherapeutic Bacteria to
903 Target Colorectal Cancer-Associated Taxa. *Int J Mol Sci.* 2020;21(3).

904 91. Wang T, Cai G, Qiu Y, Fei N, Zhang M, Pang X, et al. Structural segregation of gut microbiota
905 between colorectal cancer patients and healthy volunteers. *ISME J.* 2012;6(2):320-9.

906 92. Ranjbar M, Salehi R, Haghjooy Javanmard S, Rafiee L, Faraji H, Jafarpor S, et al. The dysbiosis
907 signature of *Fusobacterium nucleatum* in colorectal cancer-cause or consequences? A systematic
908 review. *Cancer Cell Int.* 2021;21(1):194.

909 93. Tjalsma H, Boleij A, Marchesi JR, Dutilh BE. A bacterial driver-passenger model for colorectal
910 cancer: beyond the usual suspects. *Nat Rev Microbiol.* 2012;10(8):575-82.

911 94. Xing J, Fang Y, Zhang W, Zhang H, Tang D, Wang D. Bacterial driver-passenger model in
912 biofilms: a new mechanism in the development of colorectal cancer. *Clin Transl Oncol.*
913 2022;24(5):784-95.

914 95. Castellarin M, Warren RL, Freeman JD, Dreolini L, Krzywinski M, Strauss J, et al.
915 *Fusobacterium nucleatum* infection is prevalent in human colorectal carcinoma. *Genome Res.*
916 2012;22(2):299-306.

917 96. Avril M, DePaolo RW. "Driver-passenger" bacteria and their metabolites in the pathogenesis
918 of colorectal cancer. *Gut Microbes.* 2021;13(1):1941710.

919 97. Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, et al. Tumor Microbiome
920 Diversity and Composition Influence Pancreatic Cancer Outcomes. *Cell.* 2019;178(4):795-806 e12.

921 98. Kim YI, Park JE, Brand DD, Fitzpatrick EA, Yi AK. Protein kinase D1 is essential for the
922 proinflammatory response induced by hypersensitivity pneumonitis-causing thermophilic
923 actinomycetes *Saccharopolyspora rectivirgula*. *J Immunol.* 2010;184(6):3145-56.

924 99. Schwarzer M, Makki K, Storelli G, Machuca-Gayet I, Srutkova D, Hermanova P, et al.
925 *Lactobacillus plantarum* strain maintains growth of infant mice during chronic undernutrition.
926 *Science.* 2016;351(6275):854-7.

927 100. Schnorr SL, Candela M, Rampelli S, Centanni M, Consolandi C, Basaglia G, et al. Gut
928 microbiome of the Hadza hunter-gatherers. *Nat Commun.* 2014;5:3654.

929 101. Dahl WJ, Rivero Mendoza D, Lambert JM. Diet, nutrients and the microbiome. *Prog Mol Biol*
930 *Transl Sci.* 2020;171:237-63.

931 102. Yue SJ, Liu J, Wang AT, Meng XT, Yang ZR, Peng C, et al. Berberine alleviates insulin resistance
932 by reducing peripheral branched-chain amino acids. *Am J Physiol Endocrinol Metab.*
933 2019;316(1):E73-E85.

934 103. Gojda J, Cahova M. Gut Microbiota as the Link between Elevated BCAA Serum Levels and
935 Insulin Resistance. *Biomolecules.* 2021;11(10).

936 104. Newgard CB, An J, Bain JR, Muehlbauer MJ, Stevens RD, Lien LF, et al. A branched-chain
937 amino acid-related metabolic signature that differentiates obese and lean humans and contributes to
938 insulin resistance. *Cell Metab.* 2009;9(4):311-26.

939 105. Katagiri R, Goto A, Nakagawa T, Nishiumi S, Kobayashi T, Hidaka A, et al. Increased Levels of
940 Branched-Chain Amino Acid Associated With Increased Risk of Pancreatic Cancer in a Prospective
941 Case-Control Study of a Large Cohort. *Gastroenterology.* 2018;155(5):1474-82 e1.

942 106. Wensel CR, Pluznick JL, Salzberg SL, Sears CL. Next-generation sequencing: insights to
943 advance clinical investigations of the microbiome. *J Clin Invest.* 2022;132(7).

944 107. Zierer J, Jackson MA, Kastenmuller G, Mangino M, Long T, Telenti A, et al. The fecal
945 metabolome as a functional readout of the gut microbiome. *Nat Genet.* 2018;50(6):790-5.

946 108. Schloss PD. Identifying and Overcoming Threats to Reproducibility, Replicability, Robustness,
947 and Generalizability in Microbiome Research. *mBio.* 2018;9(3).

948 109. Greathouse KL, Sinha R, Vogtmann E. DNA extraction for human microbiome studies: the
949 issue of standardization. *Genome Biol.* 2019;20(1):212.

950 110. Costea PI, Zeller G, Sunagawa S, Pelletier E, Alberti A, Levenez F, et al. Towards standards for
951 human fecal sample processing in metagenomic studies. *Nat Biotechnol.* 2017;35(11):1069-76.

952 111. O'Sullivan DM, Doyle RM, Temisak S, Redshaw N, Whale AS, Logan G, et al. An inter-
953 laboratory study to investigate the impact of the bioinformatics component on microbiome analysis
954 using mock communities. *Sci Rep.* 2021;11(1):10590.

955 112. Clausen DS, Willis AD. Evaluating replicability in microbiome data. *Biostatistics.*
956 2022;23(4):1099-114.

957 113. Sinha R, Abu-Ali G, Vogtmann E, Fodor AA, Ren B, Amir A, et al. Assessment of variation in
958 microbial community amplicon sequencing by the Microbiome Quality Control (MBQC) project
959 consortium. *Nat Biotechnol.* 2017;35(11):1077-86.

960 114. Mirzayi C, Renson A, Genomic Standards C, Massive A, Quality Control S, Zohra F, et al.
961 Reporting guidelines for human microbiome research: the STORMS checklist. *Nat Med.*
962 2021;27(11):1885-92.

963 115. Nearing JT, Douglas GM, Hayes MG, MacDonald J, Desai DK, Allward N, et al. Microbiome
964 differential abundance methods produce different results across 38 datasets. *Nat Commun.*
965 2022;13(1):342.

966 116. Mancabelli L, Tarracchini C, Milani C, Lugli GA, Fontana F, Turrone F, et al. Vaginitypes of the
967 human vaginal microbiome. *Environ Microbiol.* 2021;23(3):1780-92.

968 117. Verstraelen H, Vieira-Baptista P, De Seta F, Ventolini G, Lonnee-Hoffmann R, Lev-Sagie A. The
969 Vaginal Microbiome: I. Research Development, Lexicon, Defining "Normal" and the Dynamics
970 Throughout Women's Lives. *J Low Genit Tract Dis.* 2022;26(1):73-8.

971 118. Dominguez-Bello MG. Gestational shaping of the maternal vaginal microbiome. *Nat Med.*
972 2019;25(6):882-3.

973 119. Auriemma RS, Sciarati R, Del Vecchio G, Liccardi A, Verde N, Pirchio R, et al. The Vaginal
974 Microbiome: A Long Urogenital Colonization Throughout Woman Life. *Front Cell Infect Microbiol.*
975 2021;11:686167.

976 120. Haque MM, Merchant M, Kumar PN, Dutta A, Mande SS. First-trimester vaginal microbiome
977 diversity: A potential indicator of preterm delivery risk. *Sci Rep.* 2017;7(1):16145.

978 121. Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SS, McCulle SL, et al. Vaginal microbiome of
979 reproductive-age women. *Proc Natl Acad Sci U S A.* 2011;108 Suppl 1:4680-7.

980 122. Deo PN, Deshmukh R. Oral microbiome: Unveiling the fundamentals. *J Oral Maxillofac Pathol.*
981 2019;23(1):122-8.

982 123. Escapa IF, Chen T, Huang Y, Gajare P, Dewhirst FE, Lemon KP. New Insights into Human
983 Nostril Microbiome from the Expanded Human Oral Microbiome Database (eHOMD): a Resource for
984 the Microbiome of the Human Aerodigestive Tract. *mSystems.* 2018;3(6).

985 124. Najmanova L, Sabova L, Lenartova M, Janatova T, Mysak J, Vetrovsky T, et al. R/G Value-A
986 Numeric Index of Individual Periodontal Health and Oral Microbiome Dynamics. *Front Cell Infect*
987 *Microbiol.* 2021;11:602643.

988
989
990
991
992
993
994
995
996
997
998
999
1000
1001

1002 **Figure legends**

1003

1004 **Figure 1** *Human holobiont and its interaction with the environment.* © Linda Čihařová.

1005

1006 **Figure 2** *Stability landscape model.* (A) original state; (B) transition state; (C) new stable
1007 state. Dashed arrows indicates the disturbance, solid arrows the adaptation of the system to
1008 the disturbance. Adapted according to Folke et al. (5).

1009

1010 **Figure 3** *Long-life dynamics of gut microbiome.*

1011

1012 **Figure 4** *Variability of the environmental conditions along the gastrointestinal tract.*

1013

1014 **Figure 5** *Drivers – passengers model.* Adapted according to Tjalsma et al. (93).

1015

1016

1017

1018

1019

1020

1021

1022

1023

1024

1025

1026

1027

1028

1029

1030

1031

1032

1033

1034

1035

1036

1037

1038

1039

1040

1041



1042

1043

1044 **Figure 1** *Human holobiont and its interaction with the environment.* © Linda Čihařová.

1045

1046

1047

1048

1049

1050

1051

1052

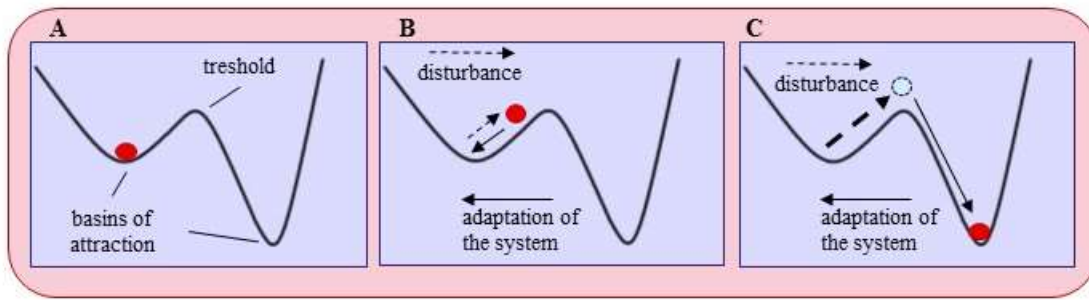
1053

1054

1055

1056

1057



1058

1059 **Figure 2** *Stability landscape model*. (A) original state; (B) transition state; (C) new stable

1060 state. Dashed arrows indicates the disturbance, solid arrows the adaptation of the system to

1061 the disturbance. Adapted according to Folke et al. (5).

1062

1063

1064

1065

1066

1067

1068

1069

1070

1071

1072

1073

1074

1075

1076

1077

1078

1079

1080

1081

1082

1083

1084

1085

1086

1087

1088

1089

1090

1091

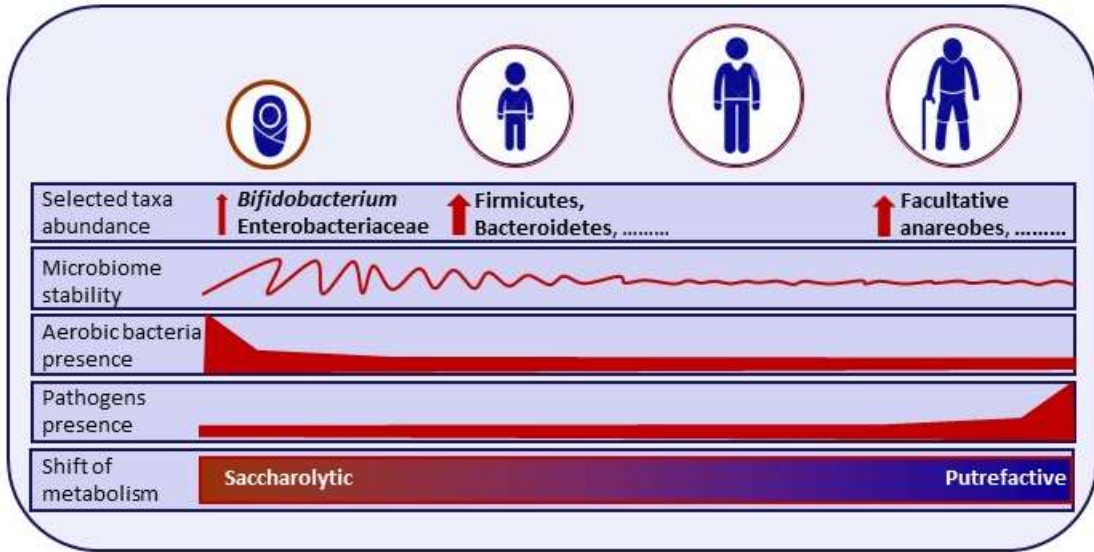
1092

1093

1094

1095

1096



1097

1098

1099 **Figure 3** Long-life dynamics of gut microbiome.

1100

1101

1102

1103

1104

1105

1106

1107

1108

1109

1110

1111

1112

1113

1114

1115

1116

1117

1118

1119

1120

1121

1122

1123

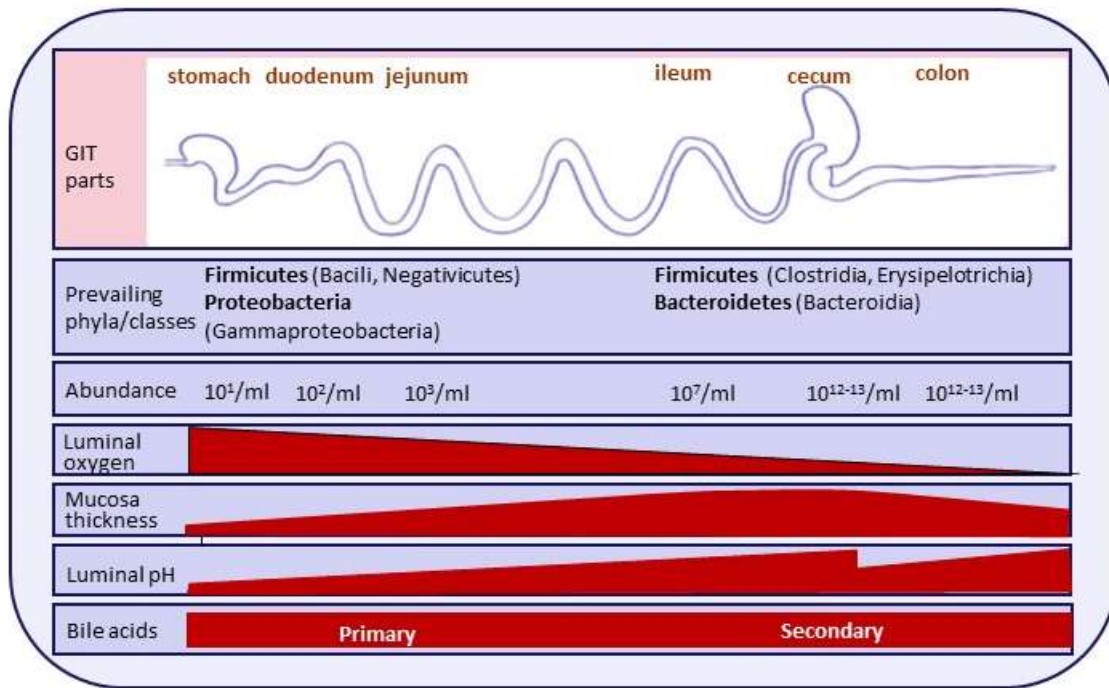
1124

1125

1126

1127

1128



1129

1130

1131 **Figure 4** Variability of the environmental conditions along the gastrointestinal tract.

1132

1133

1134

1135

1136

1137

1138

1139

1140

1141

1142

1143

1144

1145

1146

1147

1148

1149

1150

1151

1152

1153

1154

1155

1156

1157

Malignant transformation	Healthy epithelium	Tumour tissue
	Drivers	Passengers
Bacteria	inducing DNA damage in colonocytes via production of genotoxins, superoxide etc.	adapted to CRC microenvironment
Indicators of	risk of CRC	established CRC
Characteristic feature	DNA damage	favor the growth of transformed colonocytes
Representatives	<i>Helicobacter pylori</i> , <i>Enterococcus faecalis</i> , <i>Streptococcus bovis/galloyticus</i> , <i>Bacteroides fragilis</i>	<i>Fusobacterium nucleatum</i> , <i>Clostridium septicum</i> , Commensals: <i>Slackia</i> , <i>Collinsella</i> spp., <i>Roseburia</i>

1158

1159

1160 **Figure 5 Drivers – passengers model.** Adapted according to Tjalsma et al. (93).

1161

1162

1163

1164

1165

1166

1167

1168

1169