

UNESCO/IUPAC Postgraduate Course in Polymer Science

Lecture:

Molecular weight and dimensions of polymers and their assemblies

Jiří Horský

Institute of Macromolecular Chemistry ASCR, Heyrovsky sq. 2, Prague -162 06 http://www.imc.cas.cz/unesco/index.html <u>unesco.course@imc.cas.cz</u>

Polymers

M gives independent information

M affects

properties such as density, degree of crystallization, Young modulus, melt viscosity, glass transition temperature etc

Consequently, *M* affects application use and processing methods

Classical methods for determination of molecular weight of polymers (see Polymer solutions in a nutshell)

Osmotic pressure (absolute, 5 10³-10⁶)

$$\Pi_{c \to 0} = RT \ \frac{c}{M}$$

Static Light scattering (absolute, 10⁴-10⁷)

$$\left(\frac{Kc}{R_{\theta}}\right)_{c\to 0} = \frac{1}{M} \left[1 + \frac{16\pi^2 n^2}{3\lambda} \left\langle R_g^2 \right\rangle \sin^2\left(\frac{\theta}{2}\right) + \dots \right] \qquad \int_{0}^{2\theta} \int_{0}^{10\theta} \int_{0}^{1$$

2.2

Zimm plot (2 D plot for 2 independent variables) => M, A_2 , R_g Intrinsic viscosity (relative, >10³)

 $[\eta] = KM^{a}$ (Mark-Houwink Eq.)

And many more - analytical ultracentrifugation, ebullioscopy, endgroup analysis, vapor pressure osmometry, Polymer sample (usually) contains molecules of various polymerization degree Experimental methods give us only some average of the molecular weight

$$\left(\frac{Y}{c}\right)_{c\to 0} = KM^{a} \qquad Y = \sum_{i} Y_{i} \qquad cK\overline{M}^{a} = \sum_{i} c_{i}KM_{i}^{a} \qquad \overline{M} = \left(\sum_{i} w_{i}M_{i}^{a}\right)^{\frac{1}{a}}$$

 $a_{\Pi} = -1$ $a_{LS} = 1$ $a_{\eta} = 0.5 - 0.8$ flexible > 0.8 rigid

Averages with integer a can be defined as the ratio of moments around the origin

$$M_{n} = \left(\sum_{i} w_{i} M_{i}^{-1}\right)^{-1} = \frac{\sum_{i} w_{i} M_{i}^{0}}{\sum_{i} w_{i} M_{i}^{-1}} \qquad M_{w} = \sum_{i} w_{i} M_{i} = \frac{\sum_{i} w_{i} M_{i}^{1}}{\sum_{i} w_{i} M_{i}^{0}} \qquad M_{z} = \frac{\sum_{i} w_{i} M_{i}^{2}}{\sum_{i} w_{i} M_{i}^{1}}$$
number-average weight-average z-average mol. weight
$$M_{n} = \left(\sum_{i} x_{i} M_{i}\right)^{-1} = \frac{\sum_{i} x_{i} M_{i}^{1}}{\sum_{i} x_{i} M_{i}^{0}} \qquad M_{w} = \frac{\sum_{i} x_{i} M_{i}^{2}}{\sum_{i} x_{i} M_{i}^{1}} \qquad M_{z} = \frac{\sum_{i} x_{i} M_{i}^{3}}{\sum_{i} x_{i} M_{i}^{3}}$$

w weight fraction, x molar fraction

Combined averages give only partial information on the sample molecular weight non-uniformity : polydispersity index $M_{\rm w}/M_{\rm n}$ **Full information:** relative amount of each polymerization degree – a distribution function number DF - x_i , weight (mass) DF - w_i cumulative $\sum_{j=1}^{i} x_j = \sum_{j=1}^{i} w_i$ discrete vs continous DF difDF = $\frac{d \text{cumDF}}{d \text{cumDF}}$ cumulative vs differential DF dMMathematically defined DF Poisson $w_P = \frac{Pe^{-a}a^{P-1}}{(a+1)(P-1)!}$ most-probable $w_p = a^2 P(1-a)^{P-1}$ **Discrete DF** 0.1 0.1 Poisson PI=1.01 Poisson PI=1.01 Most probable PI=1.95 0.08 Most probable PI=1.95 0.08 0.06 0.06 0.04 0.04

0.02 0.02

100

75

0.00

50

DP

75

100

0.00 L

Continuous DF

Schulz-Zimm

$$dw = \frac{a^{b+1}}{\Gamma(b+1)} M^b \exp(-aM) dM$$



Principle of Size Exclusion Chromatography



solvent flow

Partition between the flowing solvent and the solvent in the pores Elution time (volume) is a function of the particle (hydrodynamic) size Above certain size all particles are equal, flow only through the interstitial volume



Detectors: (i) mass sensitive (refractometer, evaporative light scattering detection) (ii) molecularly sensitive (end group UV absorption)

(iii) molecular-mass sensitive (light scattering, viscometric)

Standard configuration (i); up-to-date = multidetector (i)&(iii)



Point to point correspondence of cumulative V and M or log M distributions



M(V) – valid for the column, polymer, solvent etc

SEC with a mass detector only

- calibration by polymer standards
- universal calibration
- hydrodynamic volume $[\eta]M(V)$ Mark-Houwink Eq. log $[\eta]M = \log K + (1+a)\log M$
- polystyrene calibration

SEC with mass & molecular-mass detectors

viscosity detector – gives *M* through the universal calibration without Mark-Houwink parameters

static light-scattering detector

Averages of *M*: directly according discreet definitions

(i) LS data are used for the calibration equation
(ii) RI data are treated in standard way



Sedimentation Analysis – Analytical Ultracentrifuge (macromolecular archelogy? - Swedberg 1923, Nobel prize 1926)

Declared clinically dead in 1980's but then resurrected (natural macromolecules)



centrifugal force=friction resistance (f u) => sedimentation velocity u

sedimentation coefficient s

$$s = \frac{u}{\omega^2 r} = \frac{M(1 - \overline{v}\rho)}{N_A f} = \frac{MD(1 - \overline{v}\rho)}{RT}$$



sedimentation creates temporal concentration gradient which is opposed by diffusion, equilibrium gradient is

$$\frac{1}{r}\frac{\mathrm{d}c}{\mathrm{d}r} \sim M$$



Elution time (volume) is a function of the particle (hydrodynamic) size bigger particles (up to 0.1 mm vs. SEC <100 nm) smaller adsorption change of mechanism (bigger particles first), absolute detection LS

Mass Spectrometry

Why MS if we have SEC? MS: $M/w_{1/2}$ =10000

Resolution SEC: $N_{to} = 5.54 (V/w_{1/2})^2 = 10000$

V/w_{1/2}=42



The Nobel Prize in Chemistry 2002

"for the development of soft desorption ionisation methods for mass spectrometric analyses of biological macromolecules"

MALDI-TOF mass spectrometry

Mass spectrometry

Principle: a charge particle is accelerated in electromagnetic field

In electric field

$$\frac{1}{2}mv^2 = zeU$$

For measurement of molecular weight, M,

(i) the sample must be vaporized and charged (ion source)

(ii) particles must be separated according to M/z (mass analyzer)

(iii) particles with the same M/z must be counted (detector)

Low-molecular-weight compounds:

M is used for identification

vaporization puts an upper limit on measured M

the most widely used electron impact ionizations fragments the sample molecules

Fragmentation gives information on the structure of the compound

For a long time, polymer researches used MS only for analyzing

polymer additives





pyrolysis products

etc...

At the same time, they were fascinated by the MS precision

M atrixA ssitedL aserD esorptionI onization

Matrix: 2,5-dihydroxybenzoic acid 1,8,9-trihydroxyantracene

Laser:

337 nm, pulse <4 ns, 40 kW/4mW

T ime O f F light M ass S pectrometry



Sample preparation



- 1- 2 μL per spot (2-10x) 0.2% sample
- 1-2% matrix
- & ionizitation agent (NaCl, AgTFA)







For samples with M>20 000 multidetector SEC is the first choice



MALDI-TOF MS can distinguish individual oligomers



and even isotopic resolution (in reflector mode)



The strength of MALDI-TOF is with samples M<20000 (For samples with M>20 000, the first choice is the multi-detector SEC)

$$M = M_{eg1} + n M_{m} + M_{eg2} + M_{ia}$$

HO $[CH_2CH_2O]$ $[CH_2CH_2O]$ $[CH_2CH_2O]$ H Na⁺



Mixture of two homopolymers



PEG:PPG

PEG : PPG 2 : 1







MALDI-TOF mass spektrum of 2nd generatiom carbosilane dendrimer (defect detection)



Literature:

Montaudo G et al. Prog. Polym. Sci 31, 277-357 (2006) www.iupac.org/publications/books/pbook/PurpleBook-C3.pdf Kostanski LK et al. J. Biochem. Biophys. Methods 58, 159–186 (2004)



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