A SIMPLE AND ROBUST APPROACH FOR GENOTYPING IN MUSACEAE

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Genotyping using SSR markers

- SSR markers have been successfully applied in molecular genotyping
- The use of SSR markers opens a possibility for automation and multiplexing which significantly increases the throughput of genotyping
- Standardized SSR genotyping platform enables to analyze large sets of accessions as well as a few individuals
- Alternative methods DArTs and GBS (Genotyping By Sequencing) are suitable for analysis of large set of accessions

We have used 19 SSR markers selected out of the initial 22 marker set (http://www.musagenomics.org/cetest_firstpage1/genomic_dna.html), for their clear reproducible amplification pattern (Crouch *et al.* 1998; Lagoda *et al.* 1998; Hippolyte *et al.* 2010)

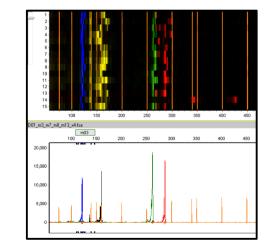
Experimental design

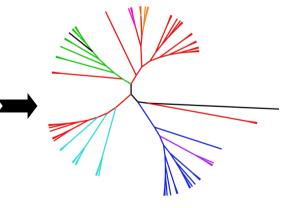
- Ploidy level estimation using flow cytometry
- Genomic DNA isolation
- PCR with fluorescently labeled primers and capillary electrophoresis analysis of resulting fragments (ABI 3730xl)
- Call for alleles and cluster analysis

Analysis of ITS sequence region, if needed



3730xl DNA analyzer



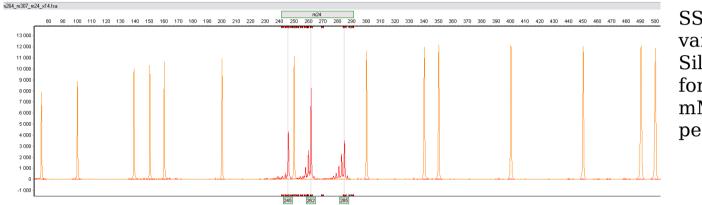


Cluster analysis

Call for alleles using GeneMarker (Softgenetics) followed by manual check

Advantages of the fragment analysis

Precise estimation of allele size, high resolution



SSR profile of a variety Sport of Silk (AAB) for marker mMaCIR24 (red peaks)

High-throughput, possible automation, possible multiplexing (5-dye set)



Multiplexed SSR profile of variety Mbwazirume (AAA) for SSR markers mMaCIR152 (green) mMaCIR195 (blue) Ma1_32 (black) mMaCIR260 (red)

Results



A platform for efficient genotyping in *Musa* using microsatellite markers

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Example of ITS analysis

• incongruence in the blind test results of the ITC 0250 accessions

• ITS analysis confirmed that the blind sample no. 4 *M. acuminata* ssp. *malaccensis* ITC 0250 is not the same genotype as the *M. acuminata* ssp. *malaccensis* ITC 0250, that we obtained from the Genbank and stored in our greenhouse

0610con 1511con2 0070con1 0609con 0093 0249 0660 0627 0672 GH D250CON2 PS BT 0250 0728 orziconi oo12con1 0543conz 06371754548 0563con1 1411con2 0637 0269con1 1411con. 0896con1 0563con2 0253con1 1027con3 0653con3 0543con1 0253con2 0370 0287 0528 0917con2 1330con1 1027con2 0011 0094con2 0638con1 1207con1 1330con2 0588con1 0543con3 0₈₇₃ 1411con3 05391TS454b 0246con2 05391754548 1120con1 0539conz 0247 0614conz 0545 Ologaconi PKWCort 1010HSASAR PUMBATS 107017545410 00^{7,3} 0246con1 1070 Mumurenot 1120con2 1021cont 0248 0927con2 Nagensium . 1072con3 1389 0927con1 1389ITS454a 1387 Musella_lassiocarpa 0.02

brown - Australimusa/Callimusa clade, green – Rhodochlamys species dark blue – BB genotypes light blue – AA genotypes pink – M. schizocarpa entries

Analysis of SSRs - conclusions

- SSR genotyping is a powerful tool for molecular characterization of *Musa* germplasm
- SSR genotyping is suitable for classification of the unknown samples
- A prior knowledge of ploidy level of an unknown sample is important prior to analysis - can be determined by flow cytometry
- In case of uncertain results, ITS sequence analysis can be employed to unravel the identity of an unknown sample
- A centralized database of molecular profiles keeps growing with every new sample, resulting in stepwise improvement in the grouping
- To facilitate building of the database of electrophoretic profiles, guarantee standard genotyping conditions and reproducibility of results, the genotyping should be centralized