

Jiří K a ň k a

**Klonování embryí savců -
reprogramace přeneseného jádra**

Ústav živočišné fyziologie a genetiky
Akademie věd České republiky
Rumburská 89
277 21 Liběchov

tel.: 315 639551
fax.: 315 697186
e-mail :kanka@iapg.cas.cz

Dolly 1997

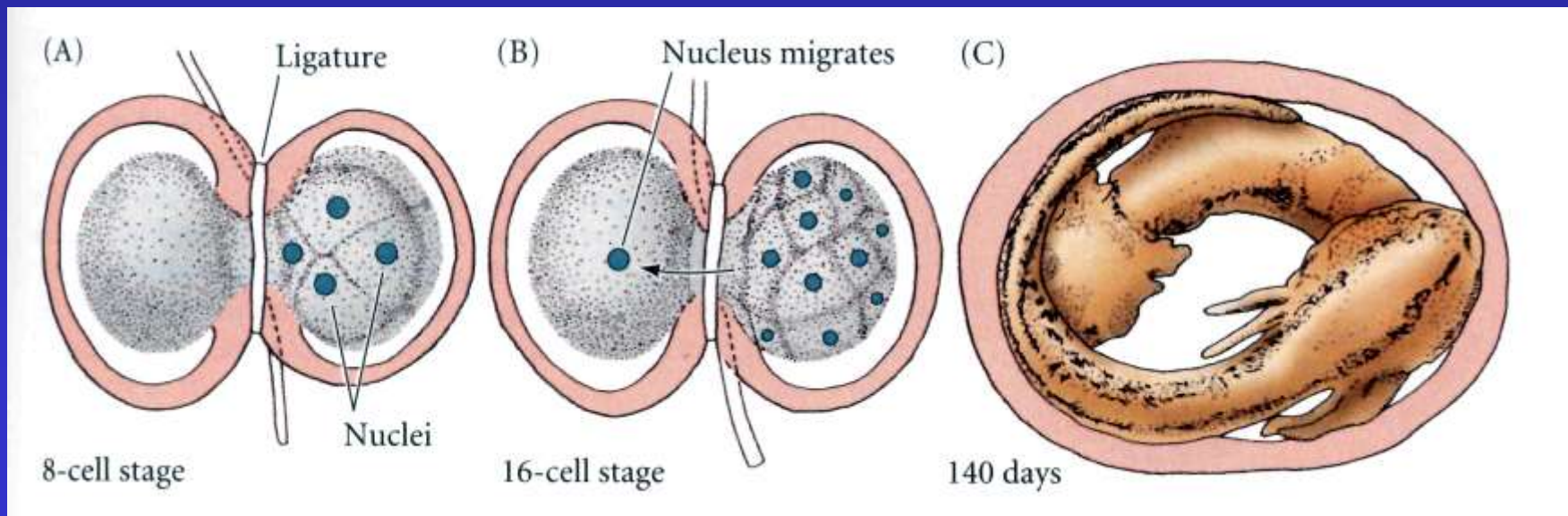


Wilmut, I./Schnieke, A.E./McWhir, J./Kind, A.J./Campbell, K.H.S., Nature, 1997

Historie klonování obratlovců

1885 - August Weissmann předpokládá, že příčinou diferenciací je diferenční dělení jader, která již od prvních dělení ztrácejí část genetické výbavy (*Ascaris megalocephala*).

1914 - Hans Spemann rozdělil vajíčko čolka na dvě části



1952 - R. Briggs a T. J. King klonovali žábu *Rana pipiens*

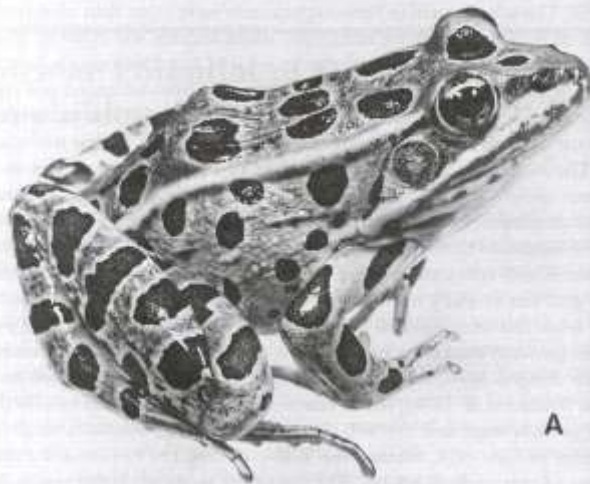
Robert Briggs



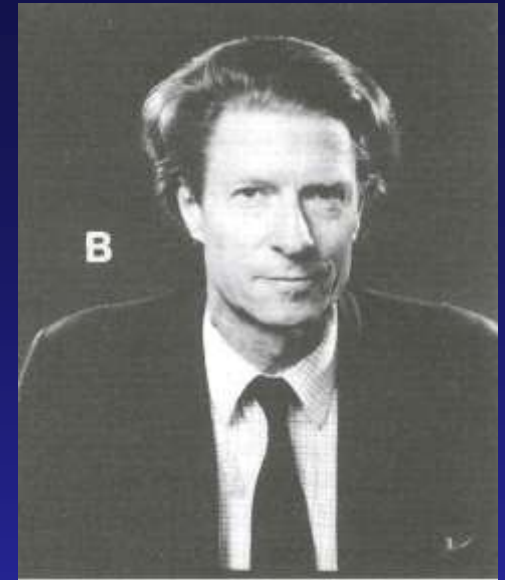
Thomas J. King



Rana pipiens

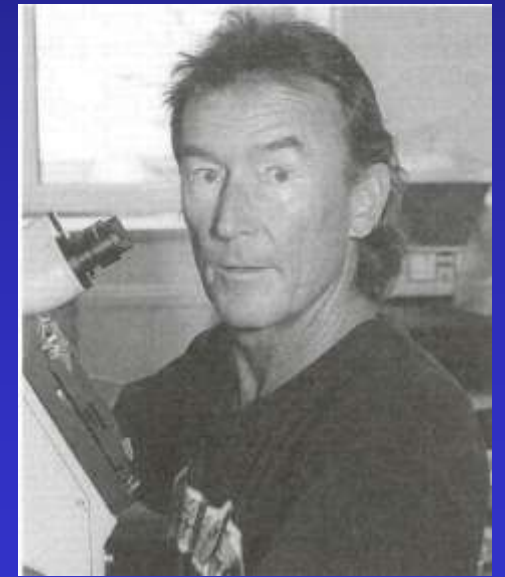


1966 – J. Gurdon publikuje klonování žáby *Xenopus laevis* z buněk střevního epitelu



1981 - K. Illmensee tvrdí, že klonoval tři myši z buněk embryoblastu

1983 - J. McGrath a D. Solter vyvinuli novou mikromanipulační metodu

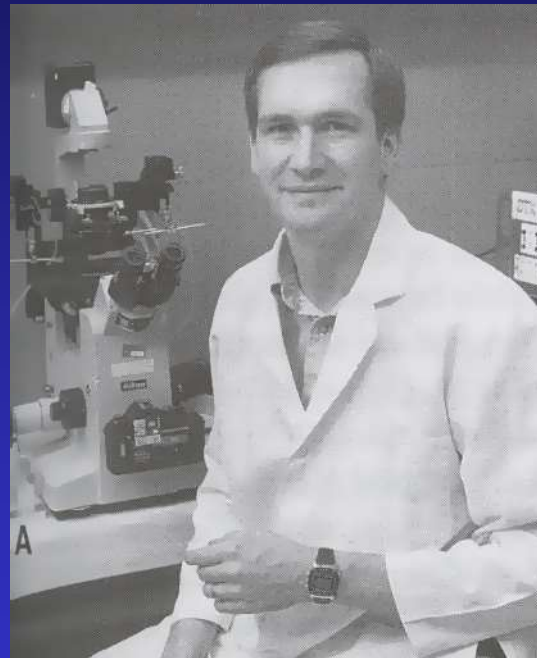


1986 - S. Willadsen klonuje ovci z embryonálních buněk

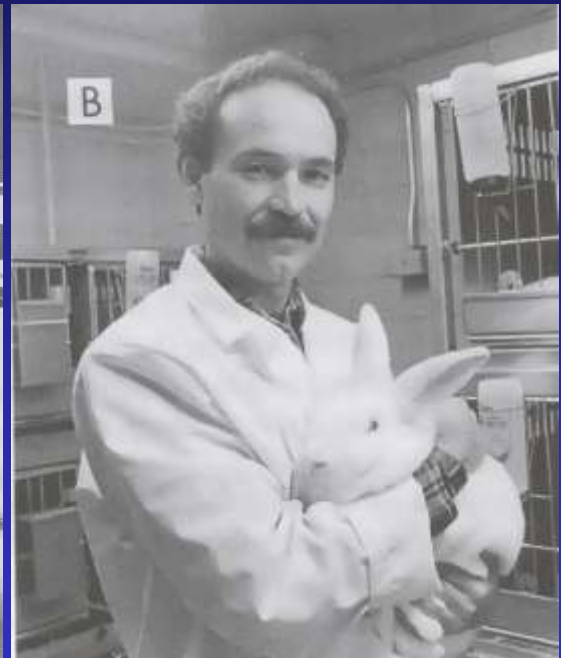
1986 - N. First, R.S. Prather a W. Eystone klonují skot z embryonálních buněk.



Steen Willadsen

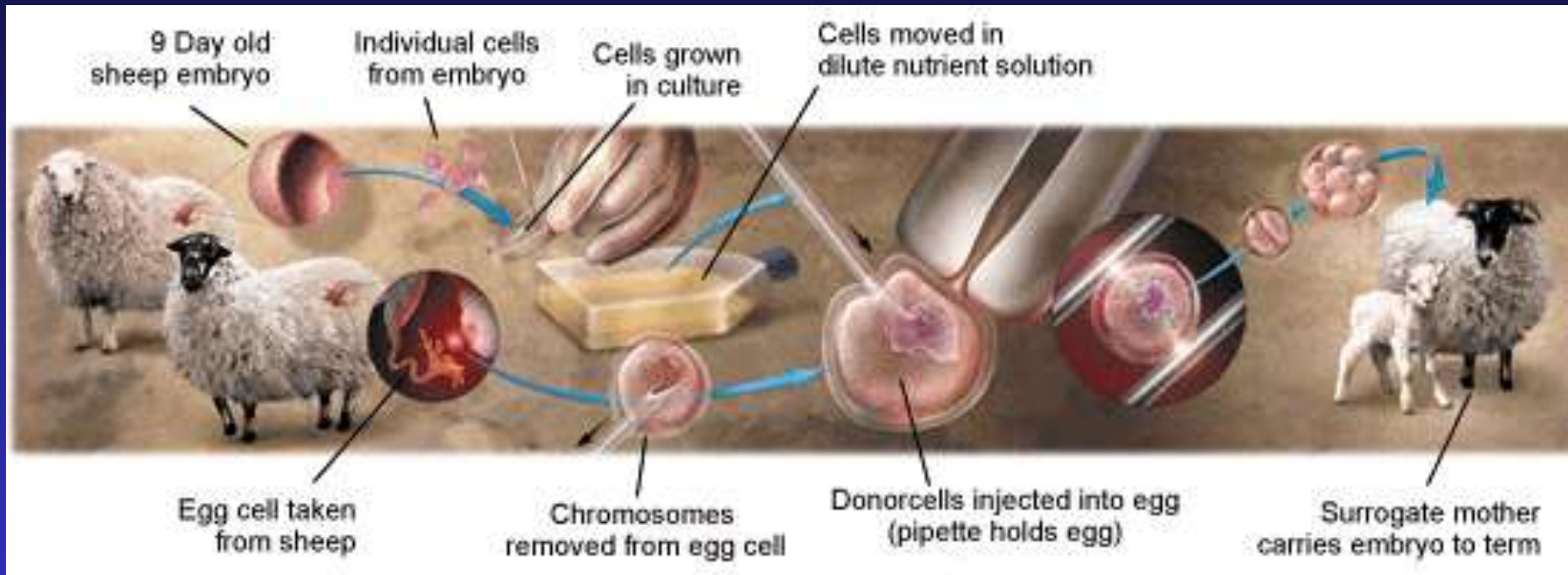


Randy Prather



Jim Robl

1995 - I. Wilmut a K. Campbell klonují ovce z diferencovaných fibroblastů kultivovaných in vitro (Campbell a kol., 1996a).

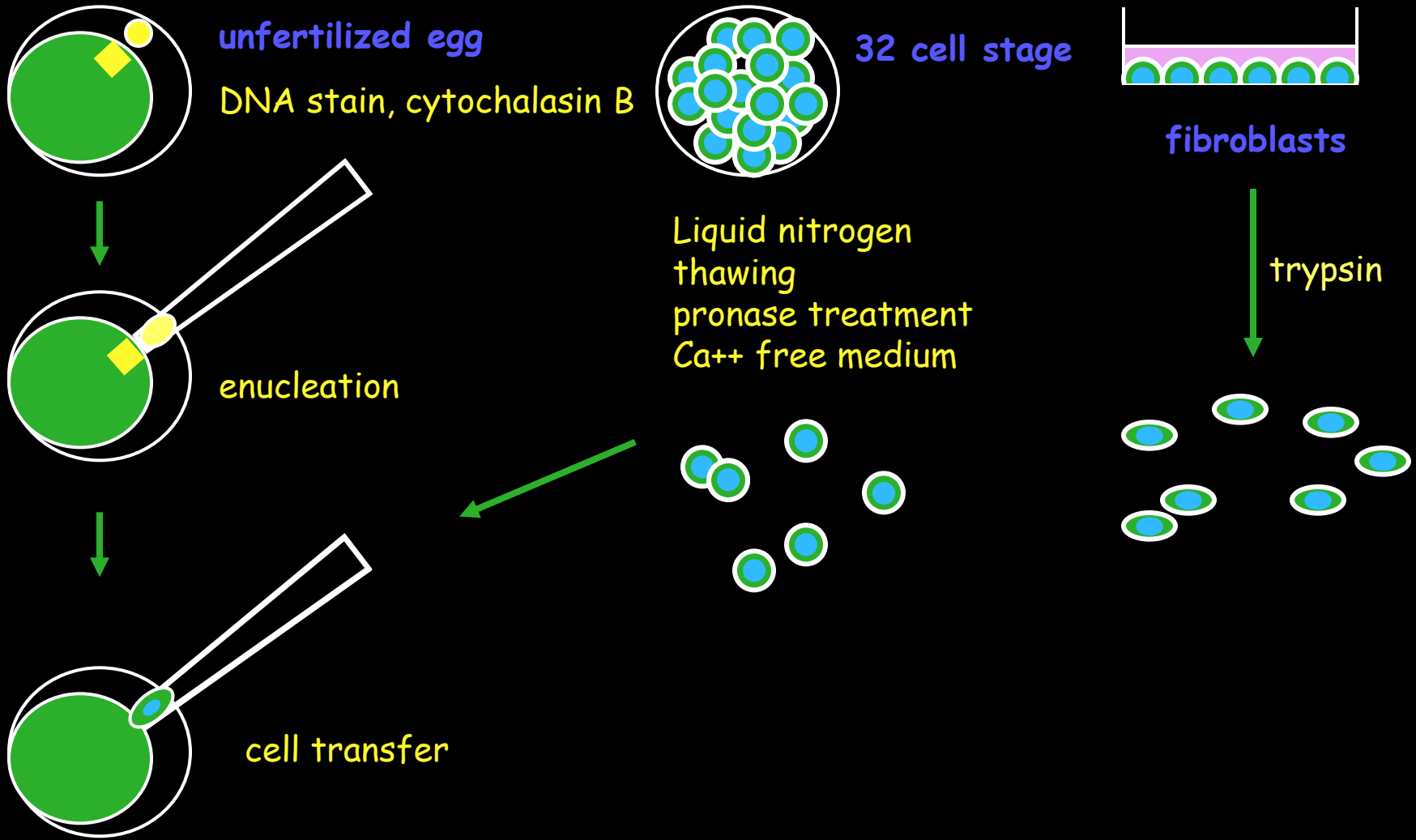


1996 - narodila se ovce Dolly, první savec klonovaný ze somatických buněk dospělého jedince

Wilmut, I./Schnieke, A.E./McWhir, J./Kind, A.J./Campbell, K.H.S.,
Nature, 1997



NUCLEAR TRANSFER IN RABBIT



Electrofusion, in vitro cultivation



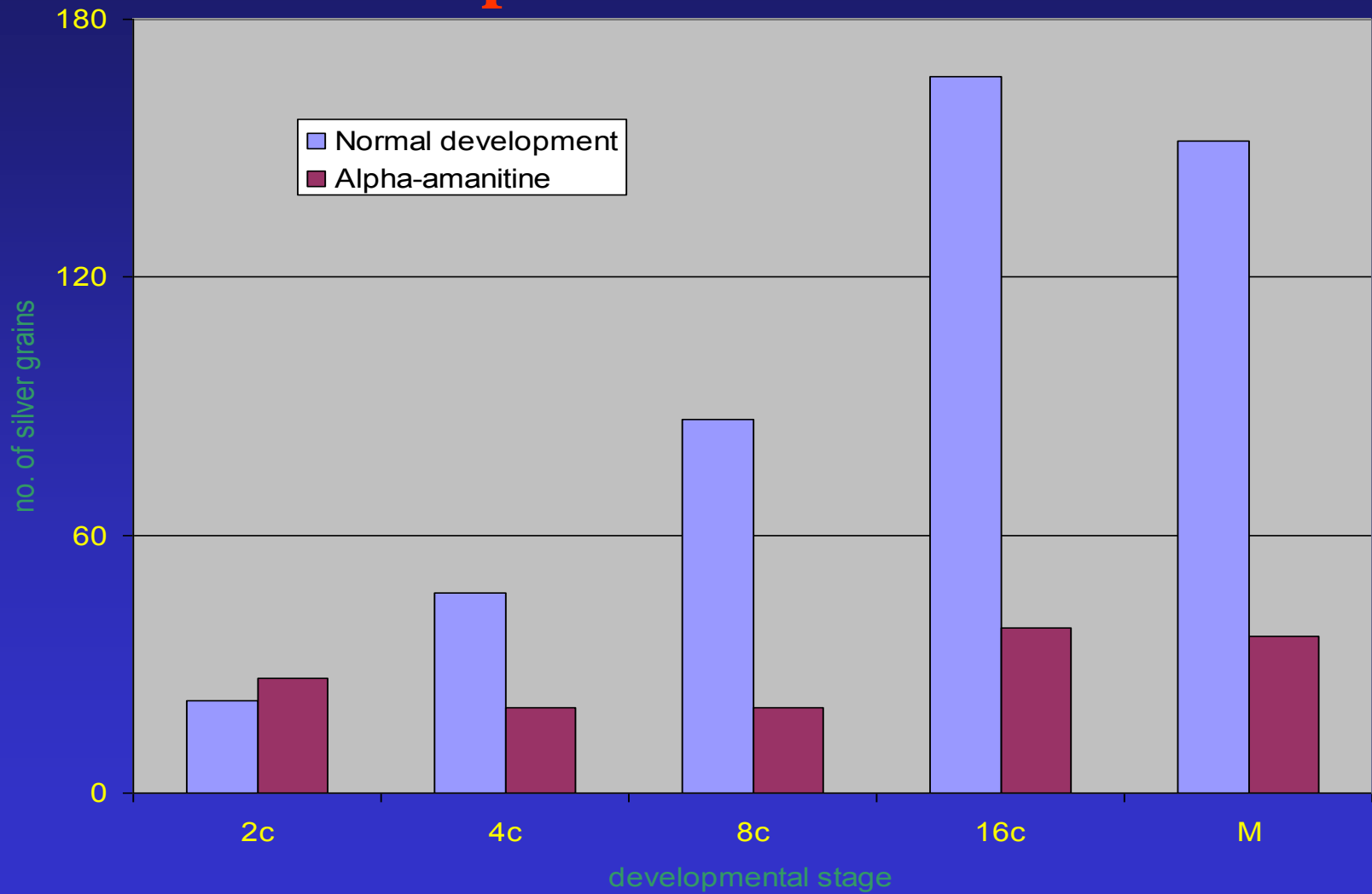


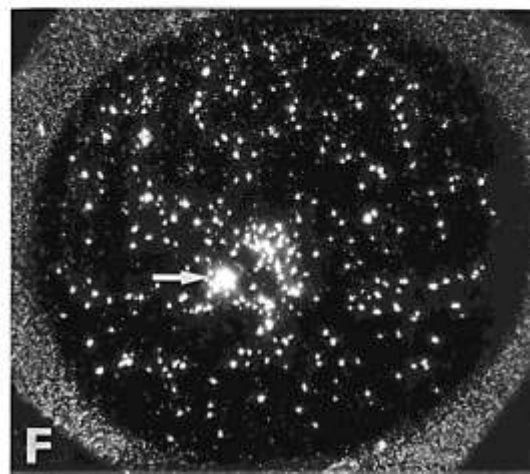
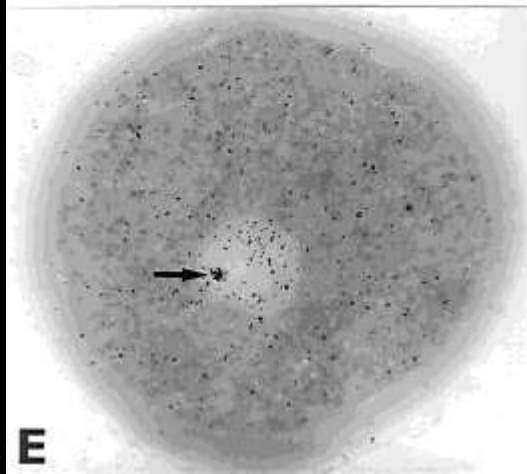
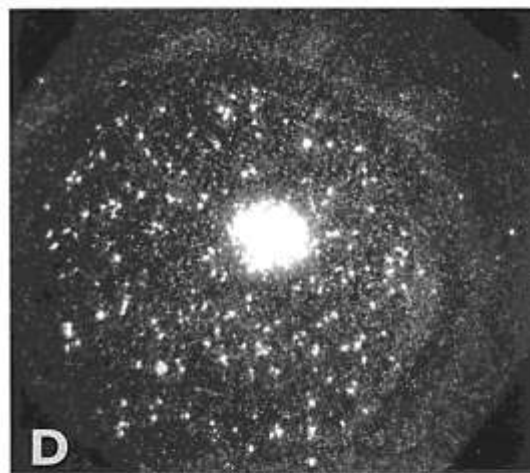
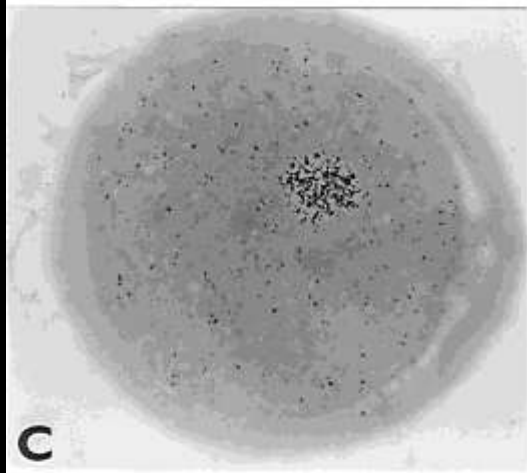
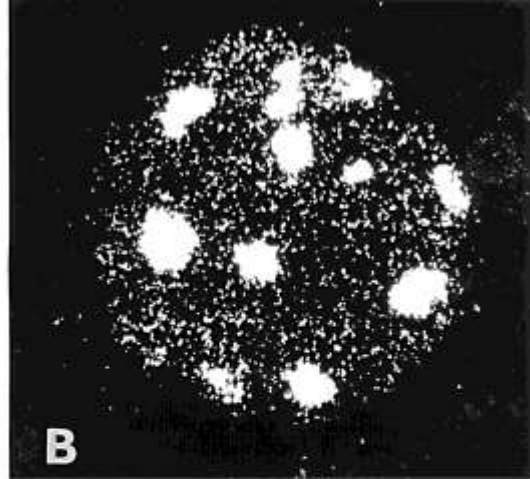
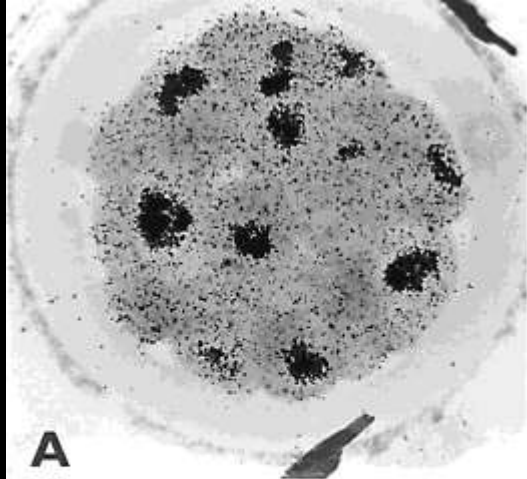
METODY

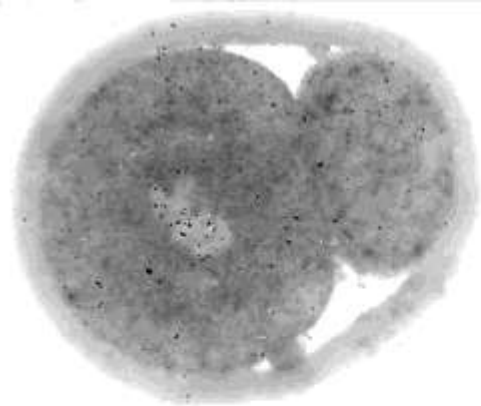
V následujících experimentech byla RNA syntéza u normálních a klonovaných embryí sledována pomocí:

- Autoradiografie (ARG)
- Elektronová mikroskopie
- mRNA differential display a následná RT-PCR

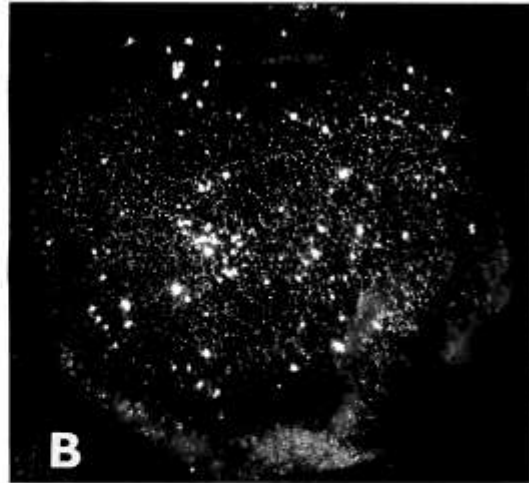
Incorporation of ^3H -uridine



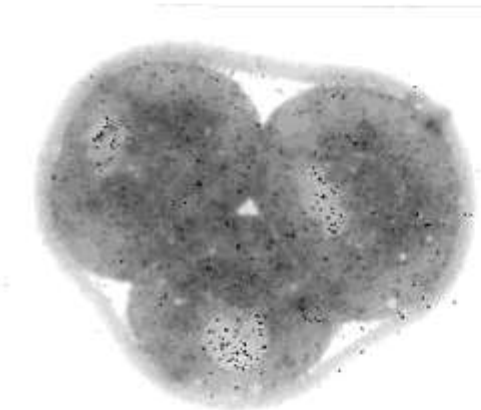




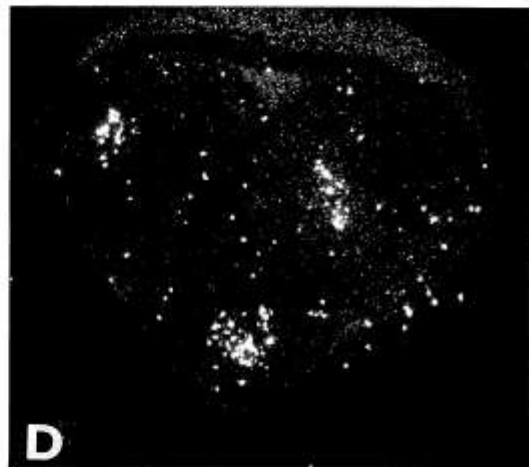
A



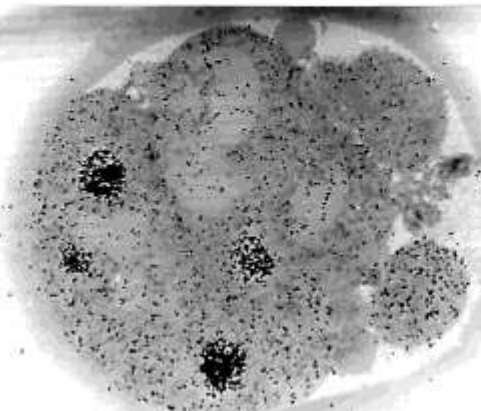
B



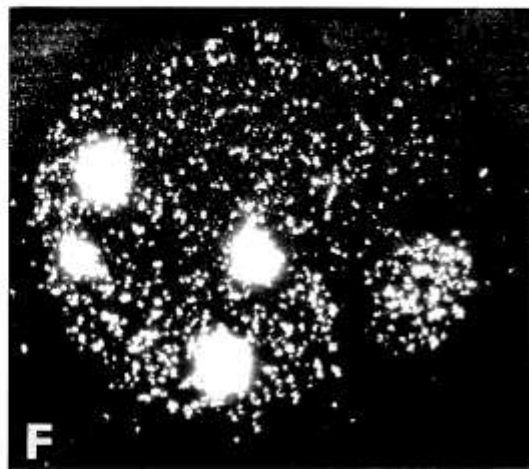
C



D

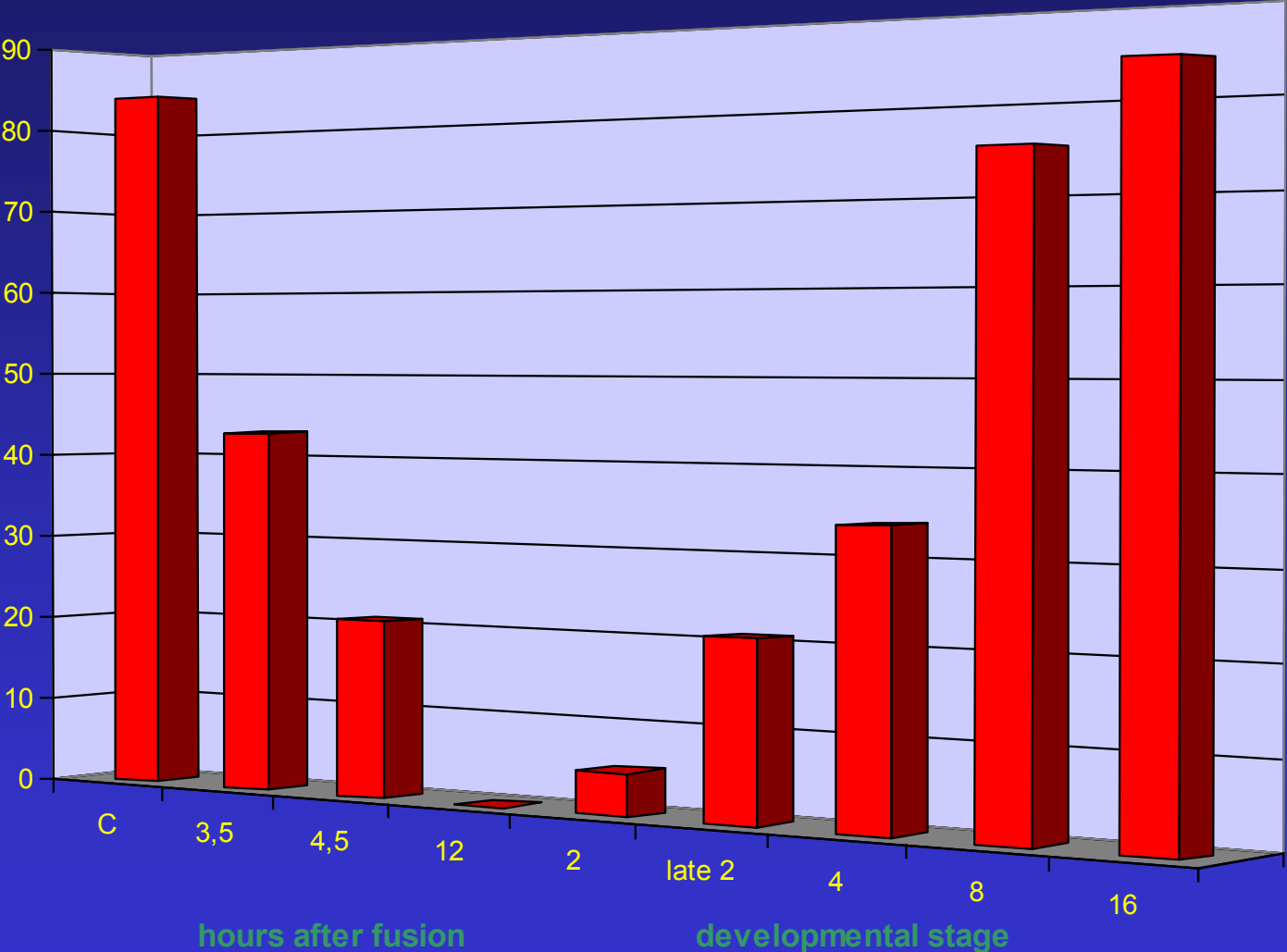


E

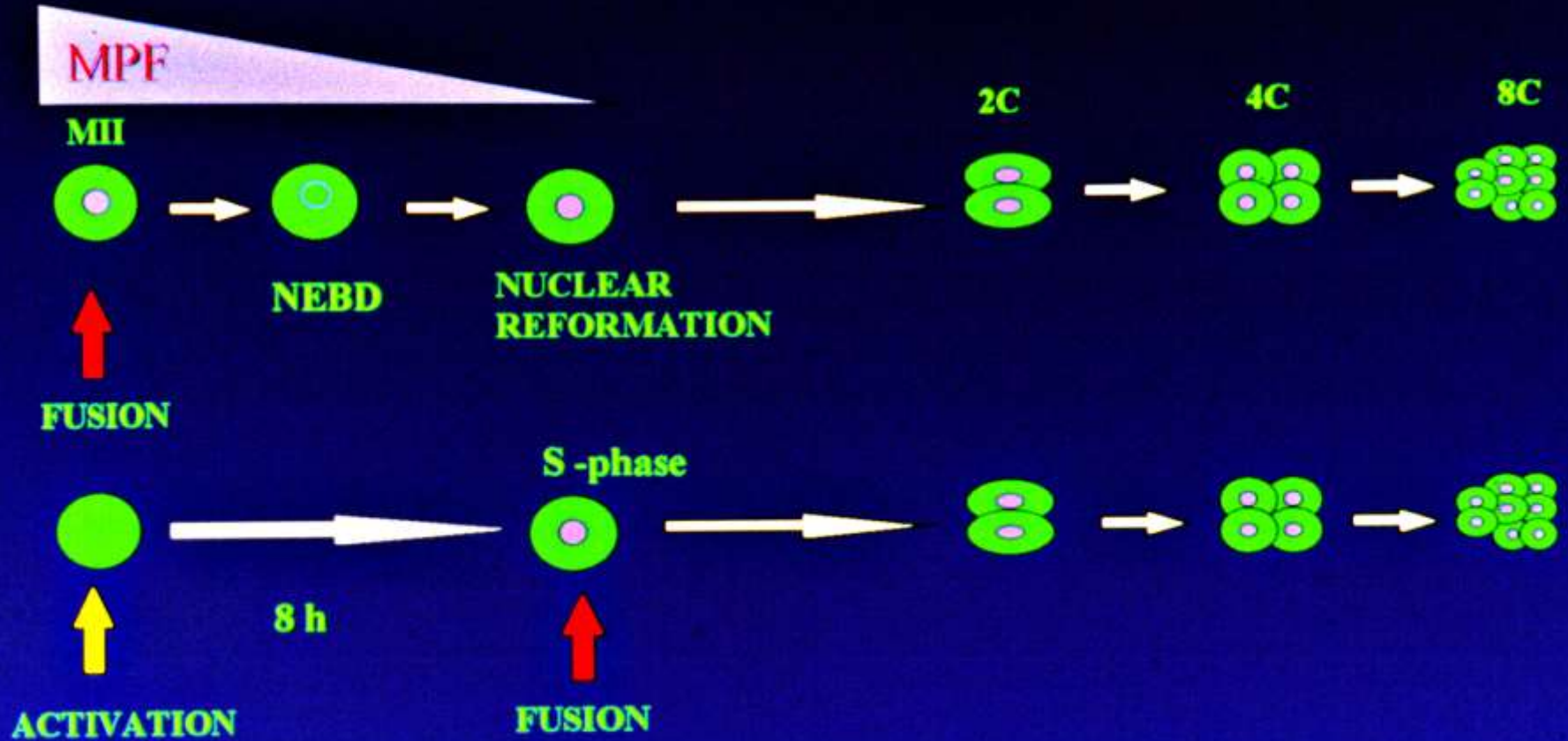


F

RNA synthesis in the nuclei of reconstructed embryos

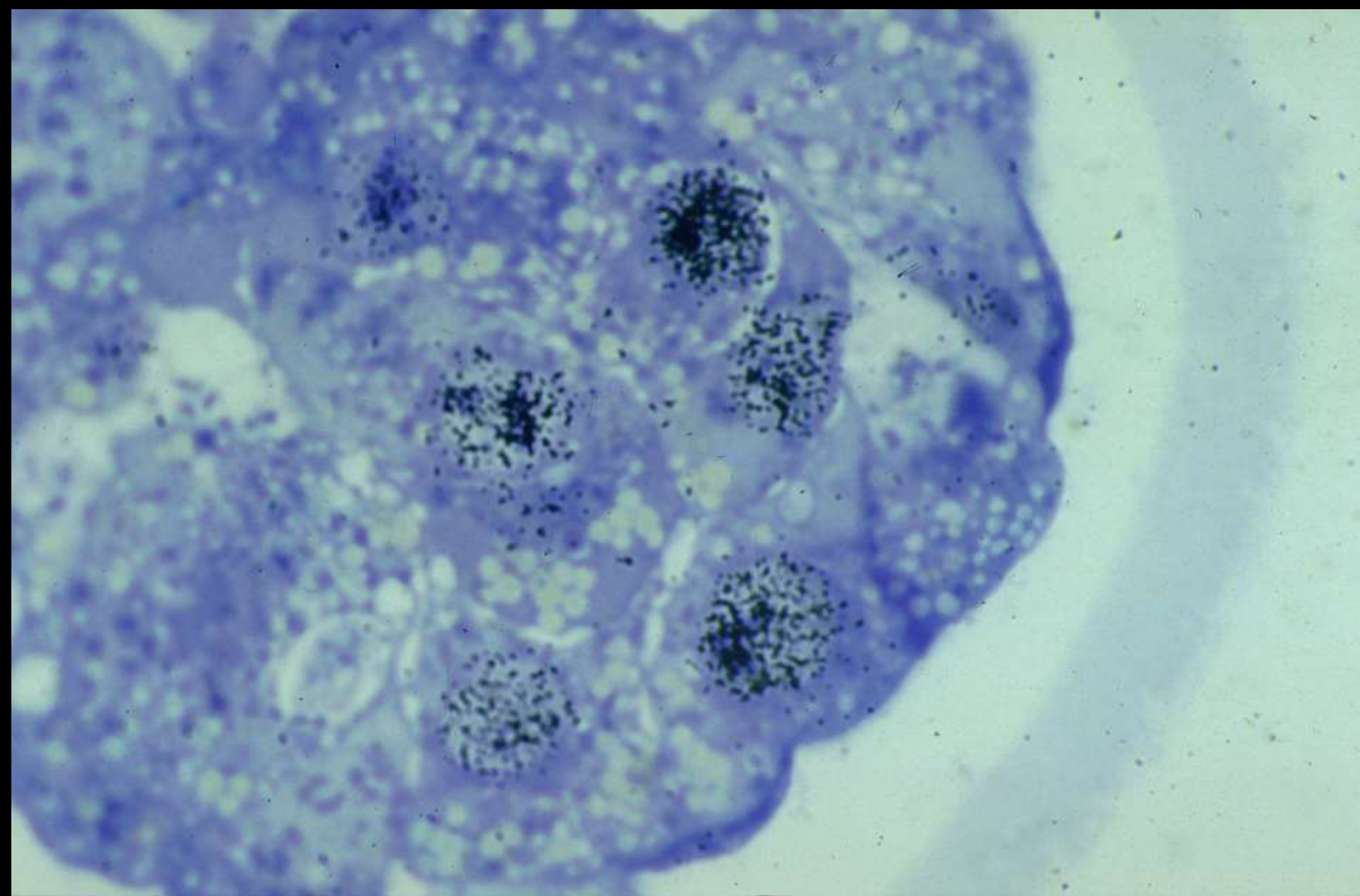


METHODS

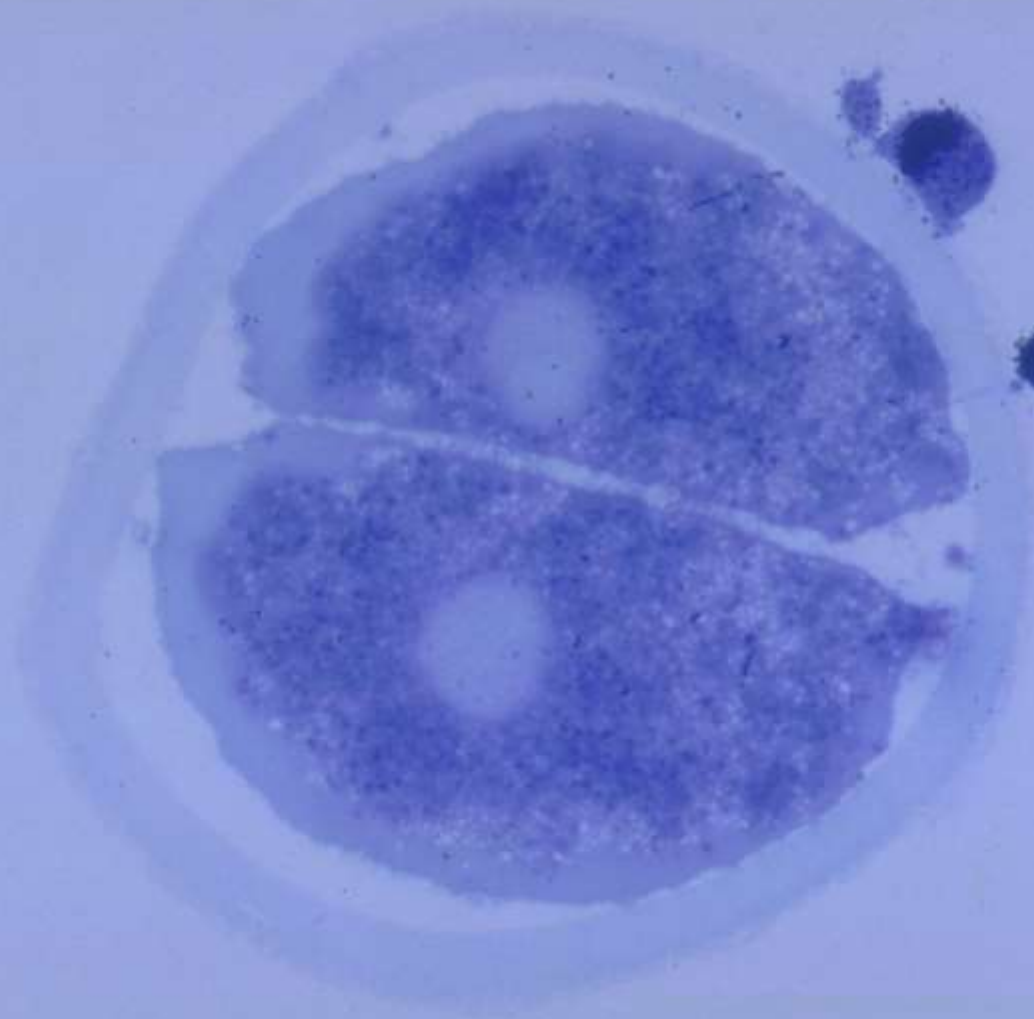


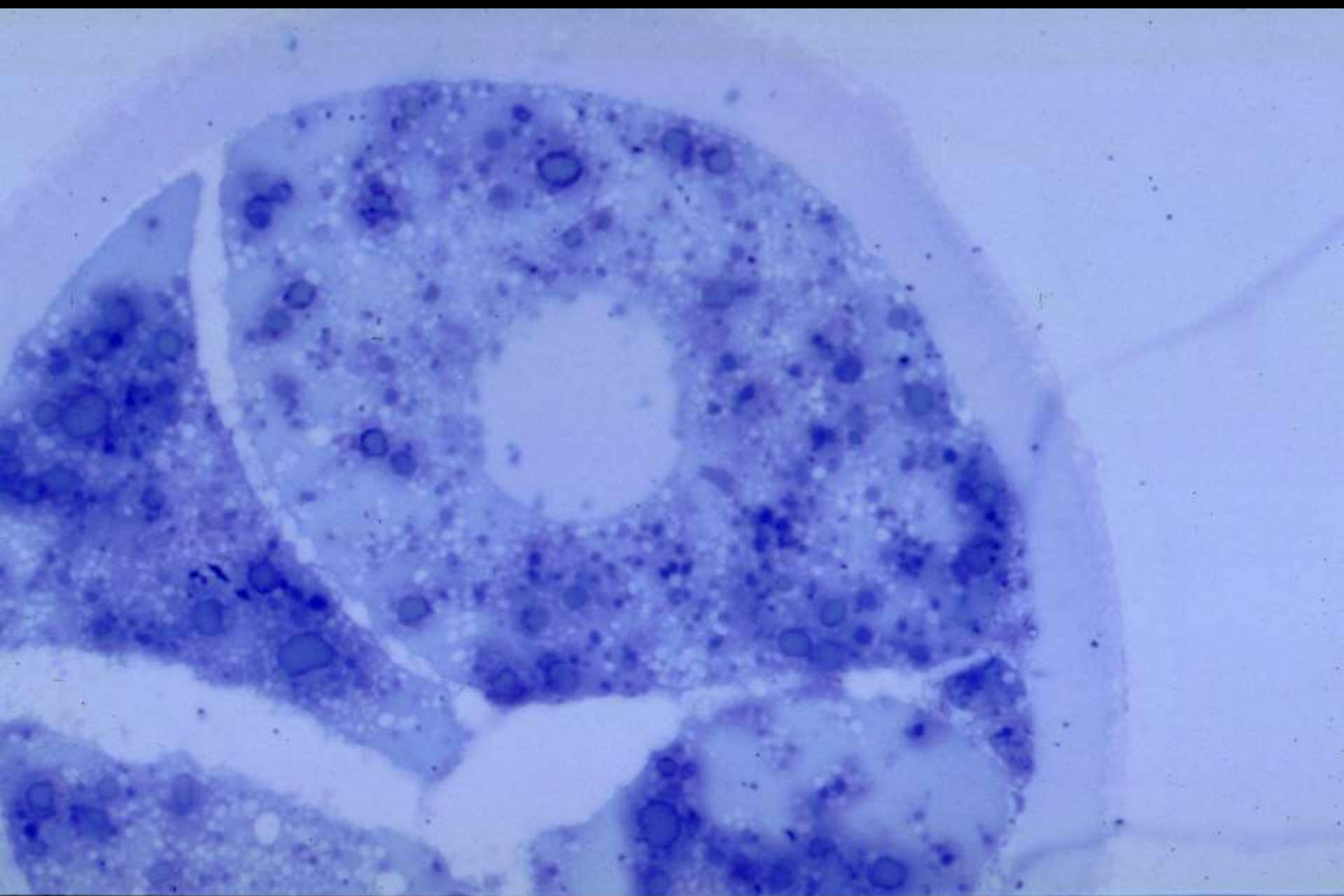
NTE were produced using either a MII phase (nonactivated) cytoplasts or S phase (activated) cytoplasts activated with calcium ionophore A23187 and cycloheximide treatment 8 h prior to fusion with a blastomere from an *in vitro* produced morula stage embryo. Control *in vitro* produced embryos were ³H-uridine labelled at the 2-, 4-, early 8- and late 8-cell stages.

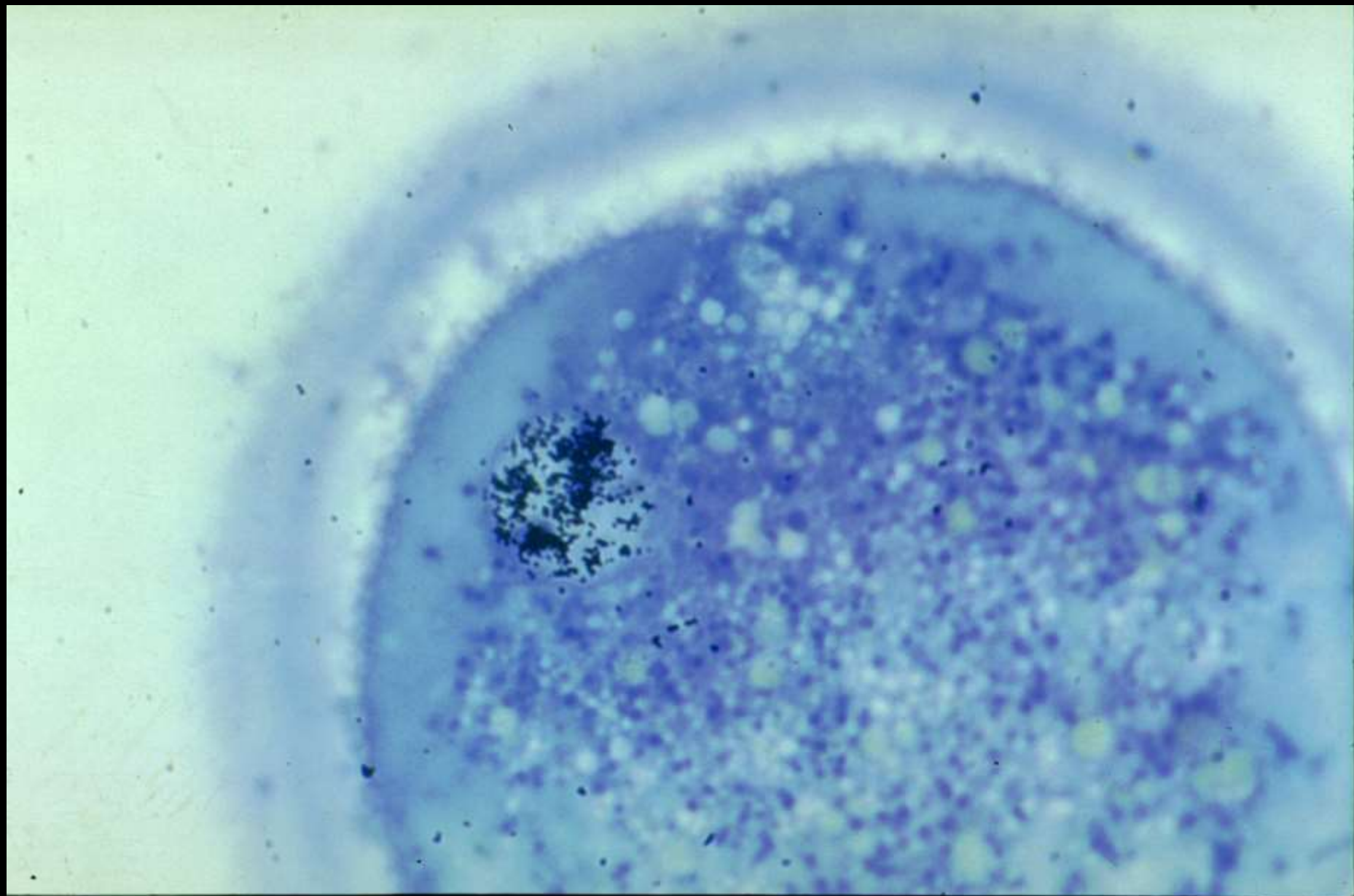
MPF – maturation promoting factor, p34^{cdc2} catalytic subunit +cyclin B regulatory component

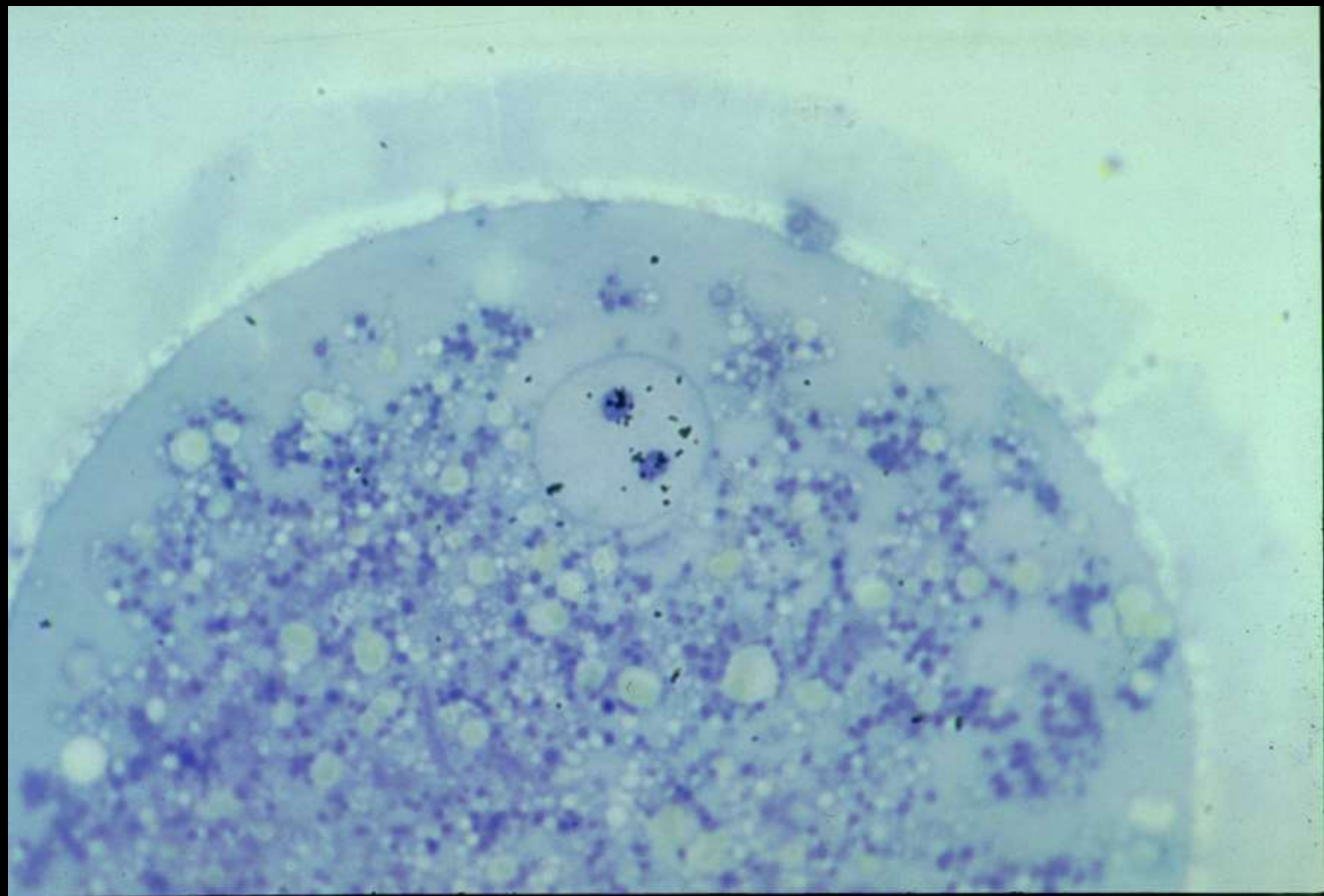


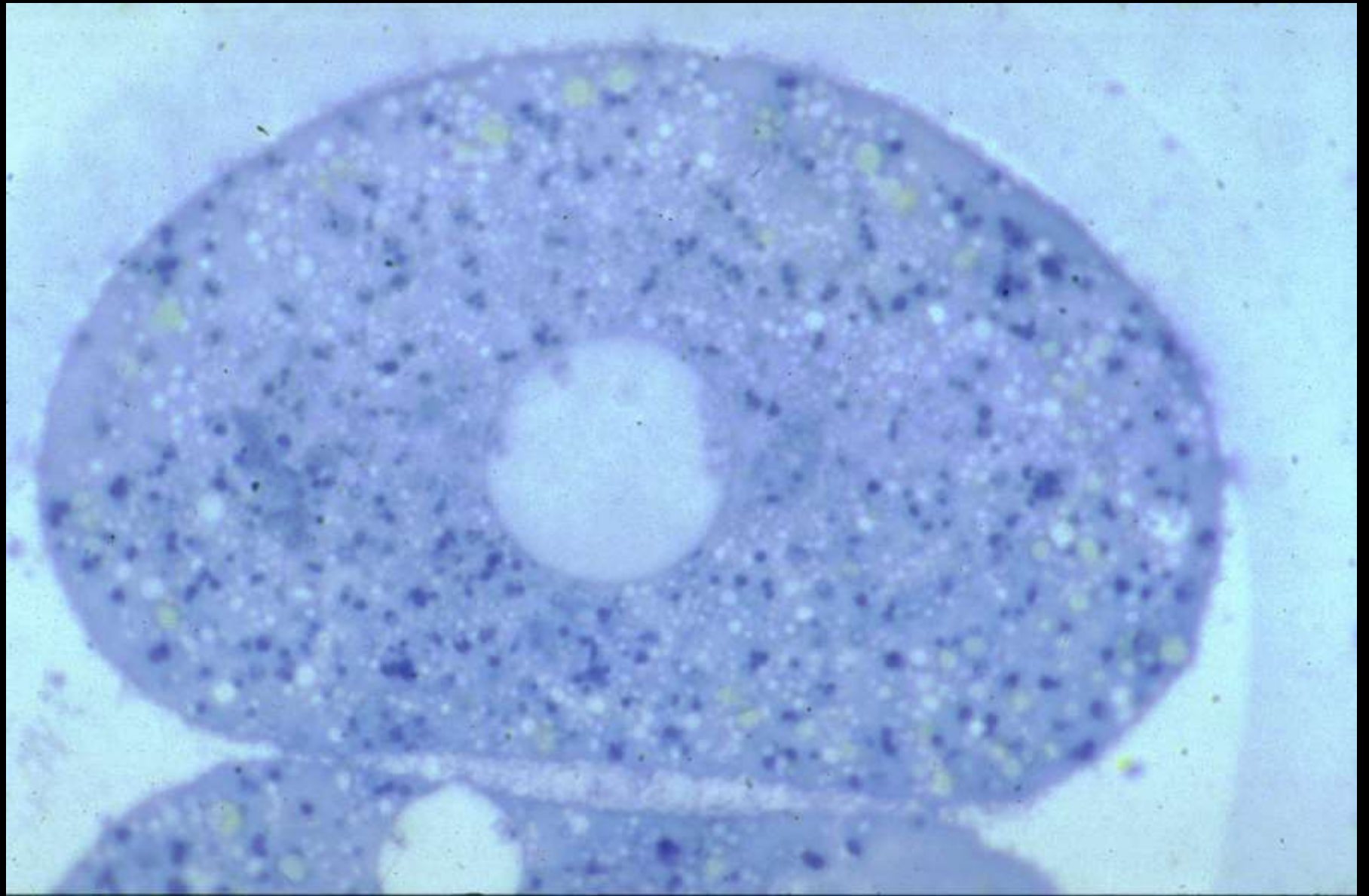




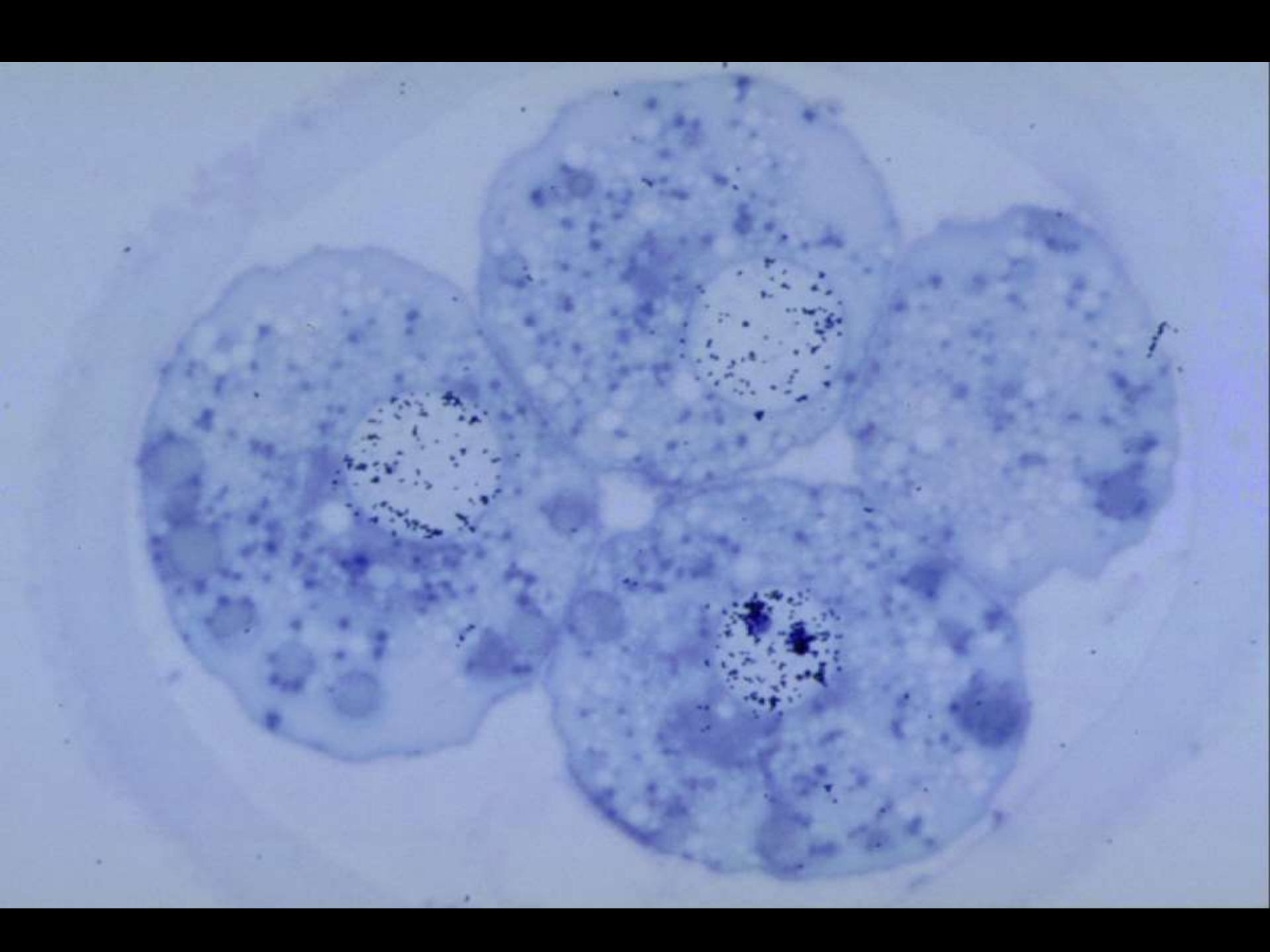












Výsledky

V kontrolních embryích skotu RNA syntéza začíná na stadiu pozdních 8 buněk, v časnějších stadiích není RNA syntéza detekovatelná

Embrya rekonstruovaná z neaktivovaných cytoplasm (MII) :

- rychlý pokles syntézy RNA 1 a 3 hod. po fúzi, absence syntézy 20 hod. po fúzi
- RNA syntéza není přítomna na 2, 4 ani 8 buněčném stadiu














Výsledky

Embrya rekonstruovaná z aktivovaných cytoplastů (S fáze) :

- pozvolný pokles RNA syntézy po fúzi, absence syntézy RNA 20 hod. po fúzi a v 2 buněčném stadiu
- počátek syntézy RNA ve 4 buněčném stadiu, tedy o jeden buněčný cyklus dříve než u kontrolních embryí

NUCLEAR TRANSFER IN BOVINE EMBRYOS

TABLE 1. Nucleolar Ultrastructure and Nuclear RNA Synthesis

	CONTROL IN VITRO	NTE NONACTIVATED CTP	NTE ACTIVATED CTP
1 CELL	 -	 -/+	 ++
2 CELL	 -	 -  -	 -  -
4 CELL	 -	DEGENERATION	 +
8 CELL	 -  +		 ++

+ POUL HYSTEL '98 +



Calf n°8803 «MARGUERITE»

Born February. 20, 1998

Dead April 4, 1998

Calf n°7810 «NARCISSE»

Born March 6, 1998



Celkové shrnutí

Určitý stupeň reprogramace jádra po přenosu je nezbytný k úspěšnému vývoji

Naprosto přesná časová posloupnost a kvantita exprese jednotlivých genů po přenosu jader není nezbytně nutná

Sledování exprese jednotlivých genů v budoucnu poskytne užitečné informace o vývoji normálního i klonovaného embrya savců

Nucleolar proteins in bovine embryos

Single nuclei from bovine embryos

1-cell

2-cell

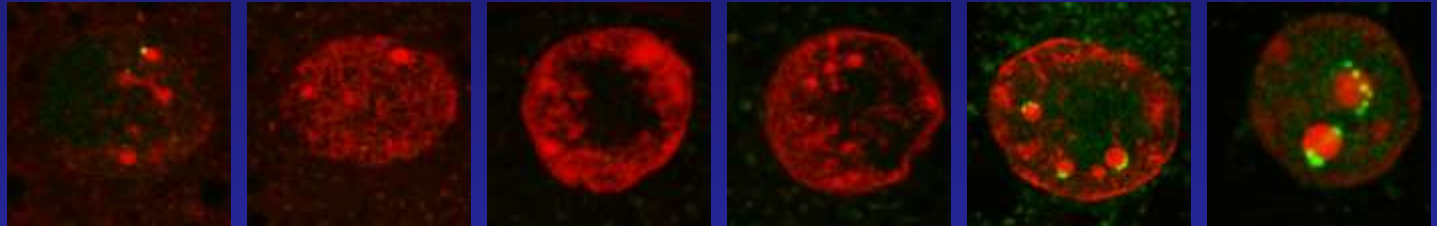
4-cell

8-cell-e

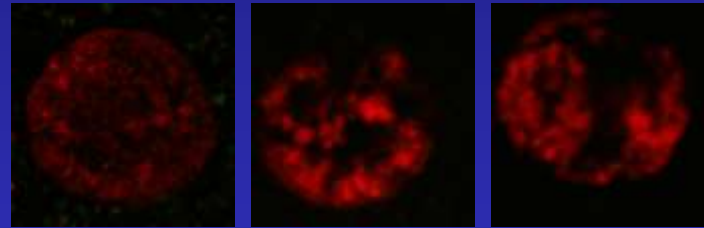
8-cell-l

16-cell

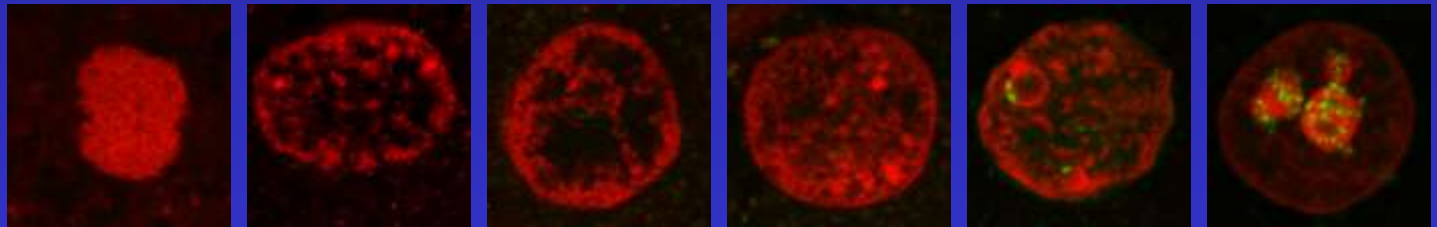
RNA pol I



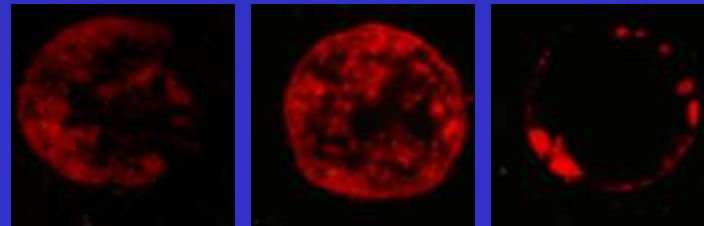
RNA pol I (amanitine)



UBF



UBF (amanitine)



Nucleolar proteins in bovine embryos

Single nuclei from bovine embryos

1-cell

2-cell

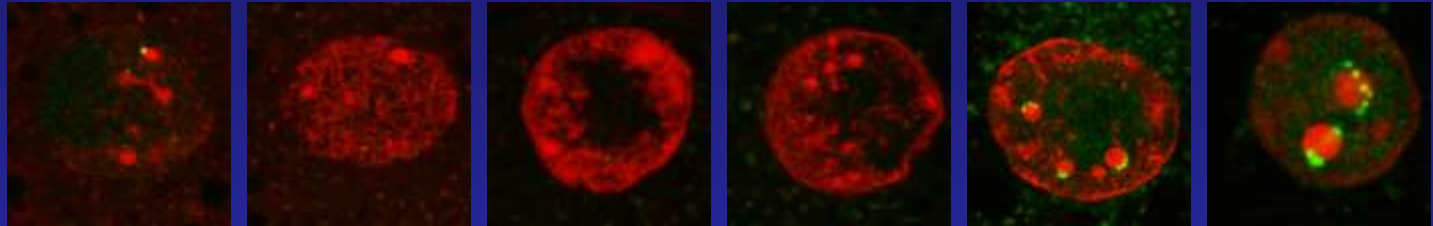
4-cell

8-cell-e

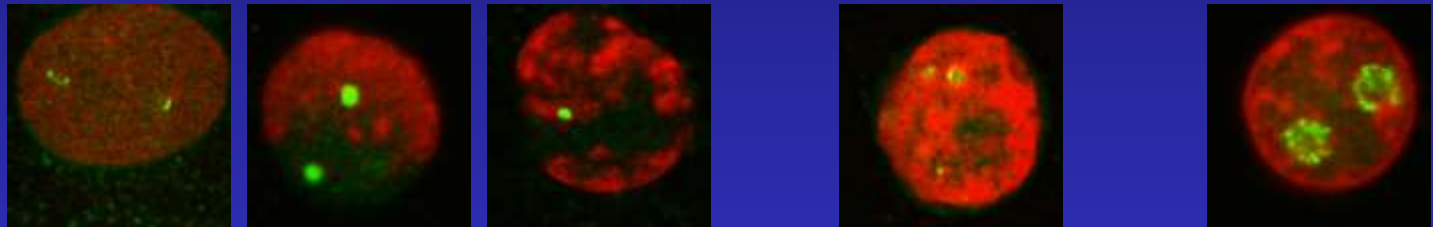
8-cell-l

16-cell

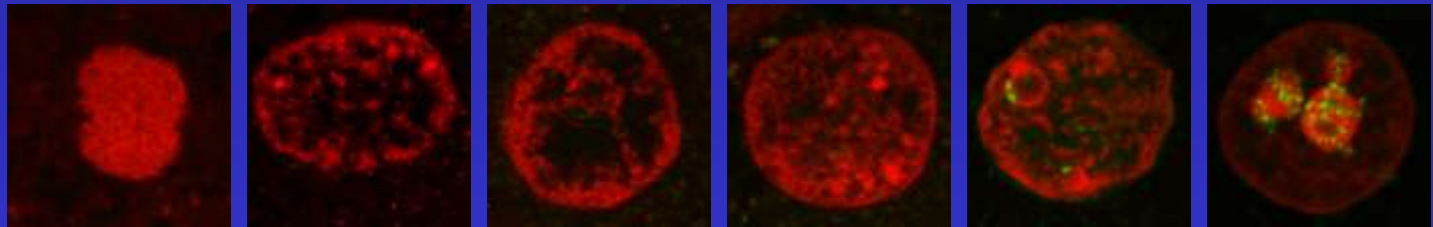
RNA pol I
IVP



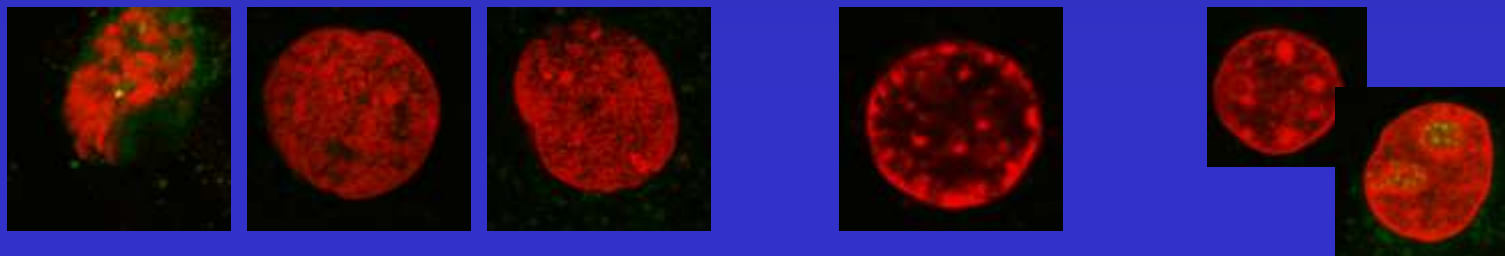
RNA pol I
NT



UBF
IVP

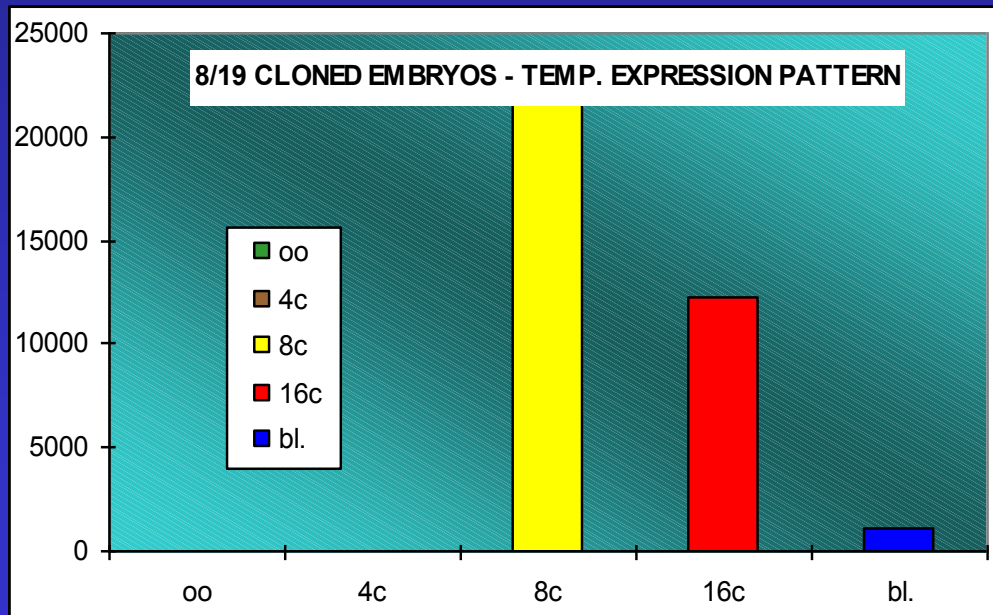
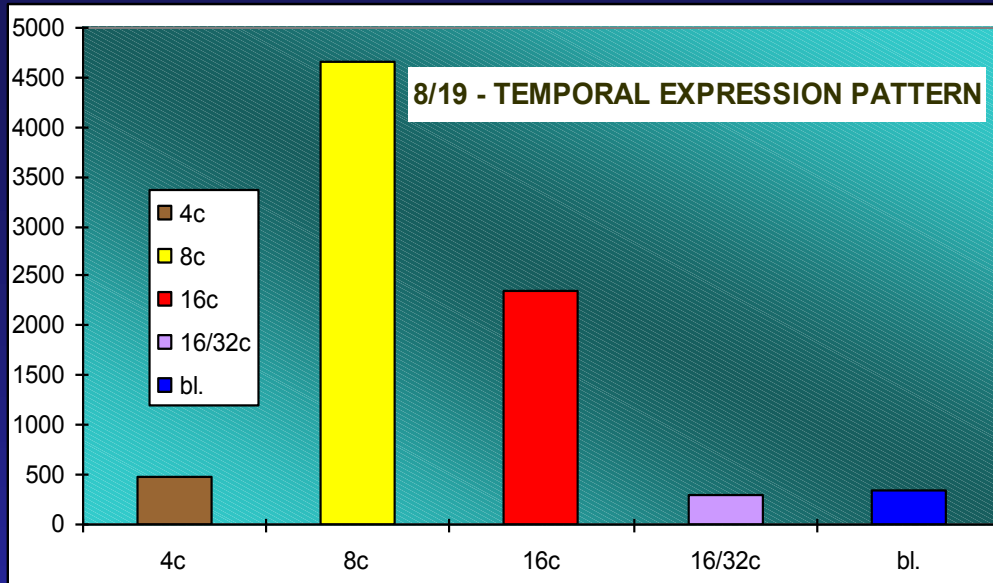


UBF
NT

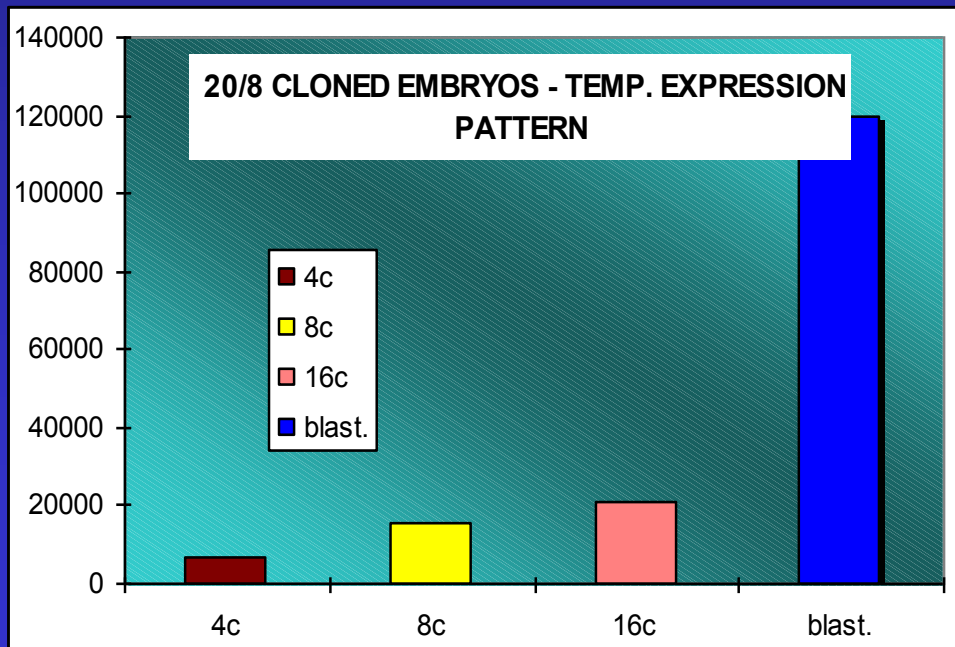
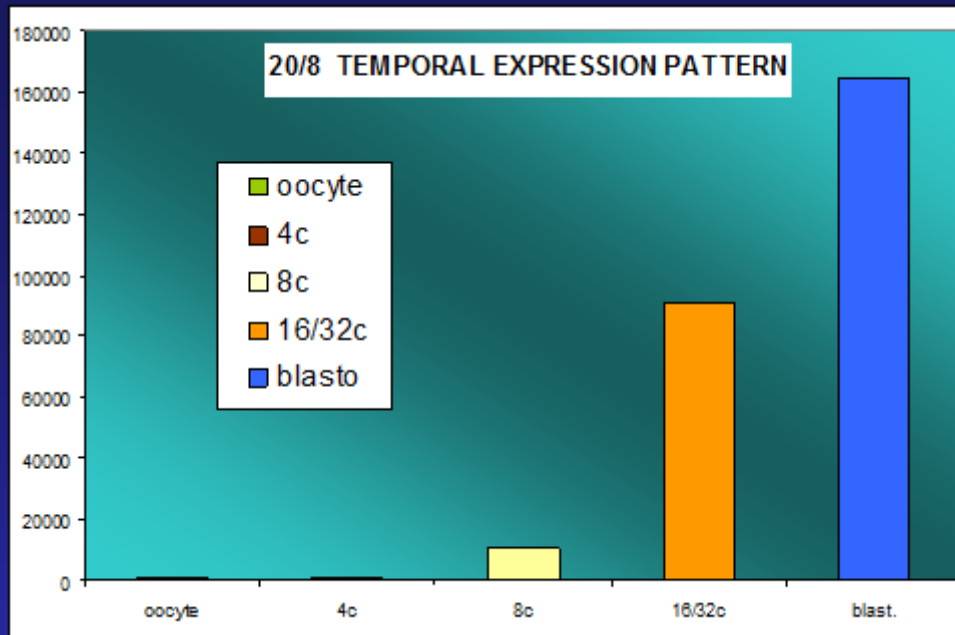


No. 8/19

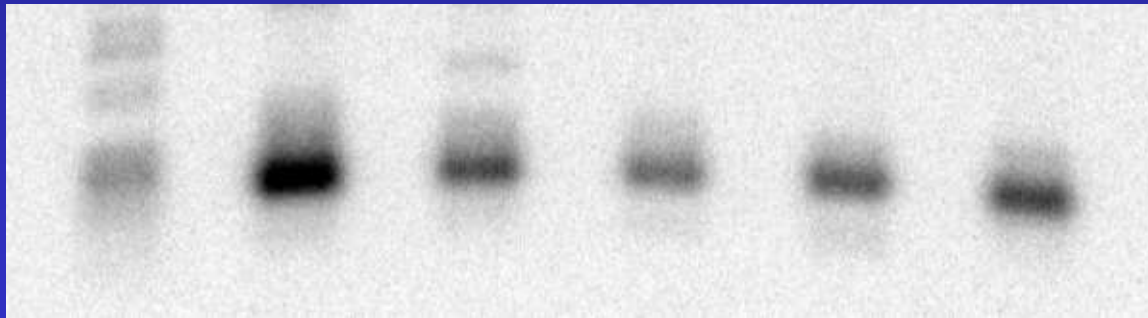
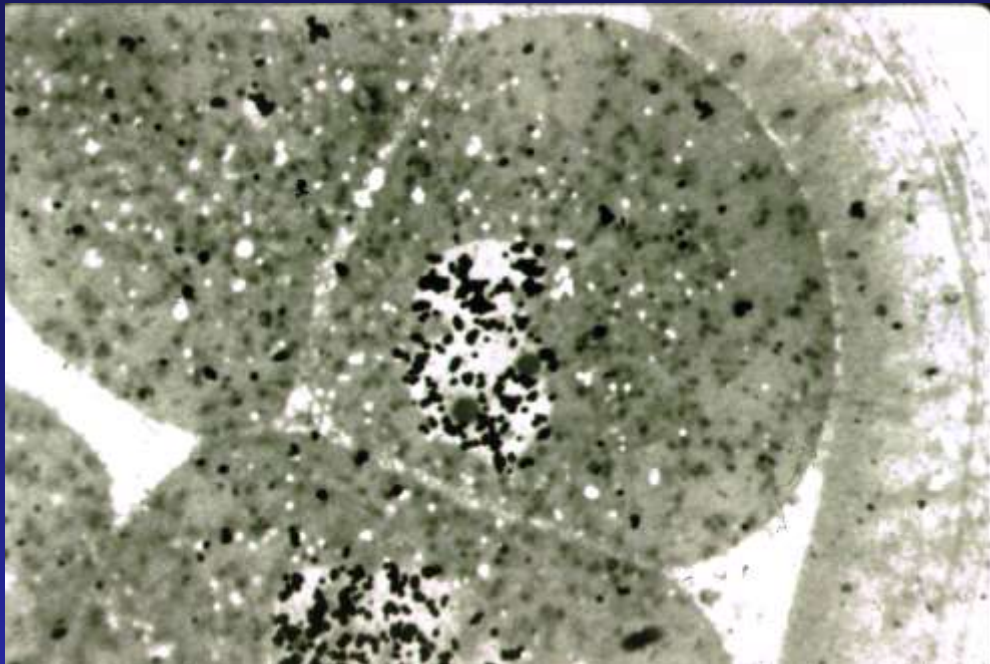
**mRNA for
proline rich
proteine**



No. 20/8
mRNA for
S3A ribosomal
protein



Srovnání ARG a semikvantitativní RT-PCR



4c 8c 16c 16-32c morula blasto.

1998 - T. Wakayama (skupina R. Yamagimashiho) vytvořil více než 50 klonovaných myší z kumulárních buněk

1999 - T. Wakayama a R. Yamagimashi klonují samce myši ze somatických buněk



- D. Wells klonuje s vysokou úspěšností telata z buněk granulocy



1998 - J.A. Thomson (Univ. of Wisconsin) - embryonální kmenové buňky člověka

J.A. Gearhart (Johns Hopkins Univ., Baltimore) – primordiální embryonální kmenové buňky člověka

1999 - Genzyme Transgenics oznamuje narození kozy, klonované ze somatických buněk, transgenních pro recombinant human antithrombin III.



Březen 2000 - PPL Therapeutics oznamuje narození 5 selat, klonovaných ze somatických buněk.

Millie, Christa, Alexis, Carrel and Dotcom, the first cloned pigs



August 22, 2002

PPL Therapeutics

Double knock-out pigs

Alpha 1,3 galactose



Noah

Advanced Cell Technology (ACT) in
Massachusetts, January 2001



Genetic rescue of an endangered mammal by
cross-species nuclear transfer using post-
mortem somatic cells

Loi P, Ptak G, Barboni B, Fulka J, Cappai P,
Clinton M

NATURE BIOTECHNOLOGY 19: (10) 962-964

OCT 2001



Science. 2003 Aug 22;301(5636):1063.

A mule cloned from fetal cells by nuclear transfer.

**Woods GL, White KL, Vanderwall DK, Li GP,
Aston KI, Bunch TD, Meerdo LN, Pate BJ.**

Northwest Equine Reproduction Laboratory,
Department of Animal and Veterinary Science,
University of Idaho, Moscow, ID 83844, USA.
gwoods@uidaho.edu

IDAHO GEM



A cloned horse born to its dam twin

(vol 424, pg 635, 2003)

**Galli C, Lagutina I, Crotti G, Colleoni S, Turini P,
Ponderato N, Duchi R, Lazzari G**

NATURE

425 (6959): 680-680 OCT 16 2003

PROMETHEA



Science. 2003 Sep 25 _
**Generation of Fertile Cloned Rats by Regulating Oocyte
Activation.**

**Zhou Q, Renard JP, Le Friec G, Brochard V, Beaujean N,
Cherifi Y, Fraichard A, Cozzi J.**

Developmental Biology and Reproduction Unit, INRA, Jouy en
Josas, France; Institute of Zoology, CAS, Beijing, RP of China.

RALPH



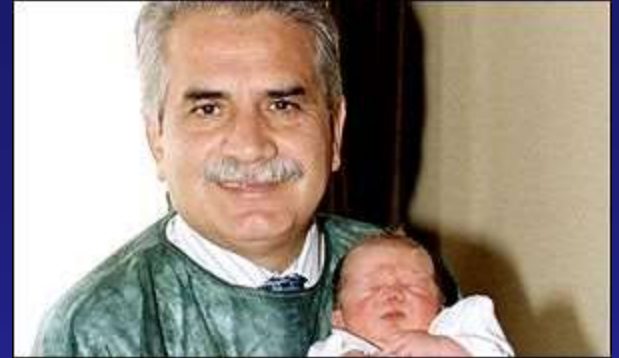
Efficiency of nuclear transfer

Species	Donor cell type	Morulae or blastocysts per fused or injected oocytes	Offspring per transferred embryos	Reference
Cattle	Cumulus cells	18/47 (38%)	5/6 (83%)	Kato et al., 1998
	Oviductal cells	20/94 (21%)	3/4 (75%)	Kato et al., 1998
	Granulosa cells	383/552 (69%)	10/100 (10%)	Wells et al., 1999
	Mammary epithelial cells	36/140 (26%)	1/4 (25%)	Zakhartchenko et al., 2000
		48/149 (32%)	2/45 (4%)	Kishi et al., 2000
	Fetal fibroblasts	24/92 (26%)	7/8 (87%)	Urukawa et al., 2001
	Calf ear skin cells	n.d. (n.d.)	1/6 (17%)	Renard et al., 1999
	Adult ear fibroblasts	49/82 (60%)	1/16 (6%)	Zakhartchenko et al., 1999
		80/197 (41%)	2/43 (4,7%)	Kishi et al., 2000
	Adult fibroblasts (17 yr)	131/440 (30%)	6/54 (11%)	Kubota et al., 2000
	Transfected fetal fibroblasts	33/276 (12%)	4/28 (14%)	Cibelli et al., 1998
		87/1896 (5%)	6/79 (8%)	Lanza et al., 2000
	Fetal, newborn, adult cells	980/2529 (39%)	24/172 (14%)	Kato et al., 2000
Overall efficiency		21-69%	4-83%	

Efficiency of nuclear transfer

Mouse	Cumulus cells	1385/2468 (56%)	16/1385 (1%)	Wakayama et al., 1998
	Fetal ovary cells	108/191 (56%)	4/108 (3,7%)	Wakayama and Yanagimachi, 2001
	Fetal testis cells	114/212 (54%)	2/112 (1,8%)	Wakayama and Yanagimachi, 2001
	Immature Sertoli cells	210/646 (33%)	7/215 (3,3%)	Ogura et al., 2000
	Fetal fibroblasts	278/932 (30%)	5/272 (1,8%)	Ono et al., 2001
	Tail tip fibroblasts	341/633 (54%)	3 (0,5%)	Wakayama and Yanagimachi 1999
	Transfected tail tip fibroblasts	281/832 (34%)	8/301 (2,7%)	Ogura et al., 2000
Overall efficiency		30-59%	0,5-3,7%	
Rabbit	Cumulus cells	371/612 (61%)	6/371 (1,6%)	Chesne et al., 2002
	Cumulus cells	n.d. (n.d.)	1/87 (1,1%)	Shin et al., 2002

Dr. Severino Antinori



Dr. Panos Zavos



November 26, 2001

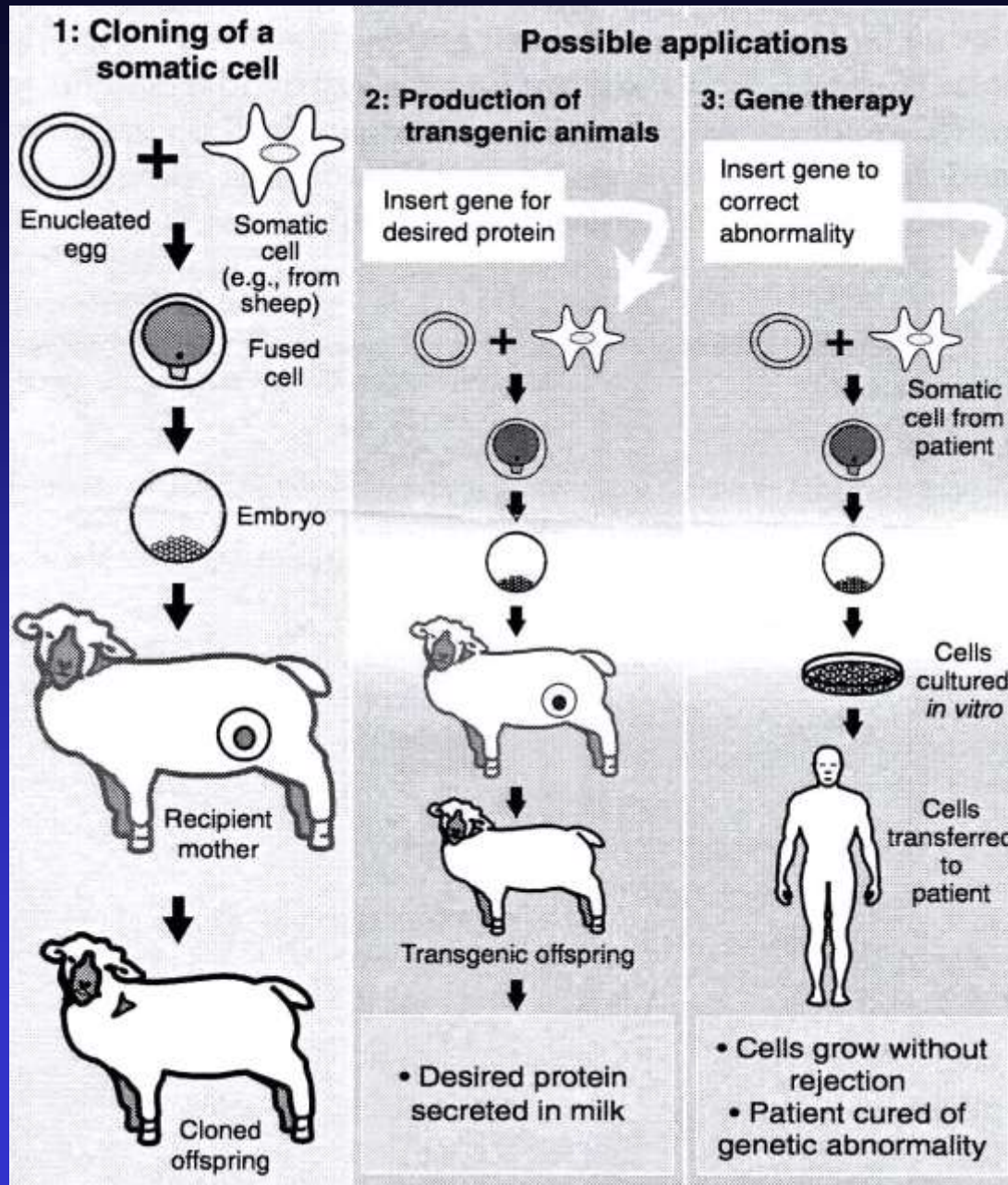
**Advanced Cell Technology Inc.
(ACT), Worcester, Massachusetts**

Jose B. Cibelli, Ph.D., D.V.M., Vice-
President of Research at ACT



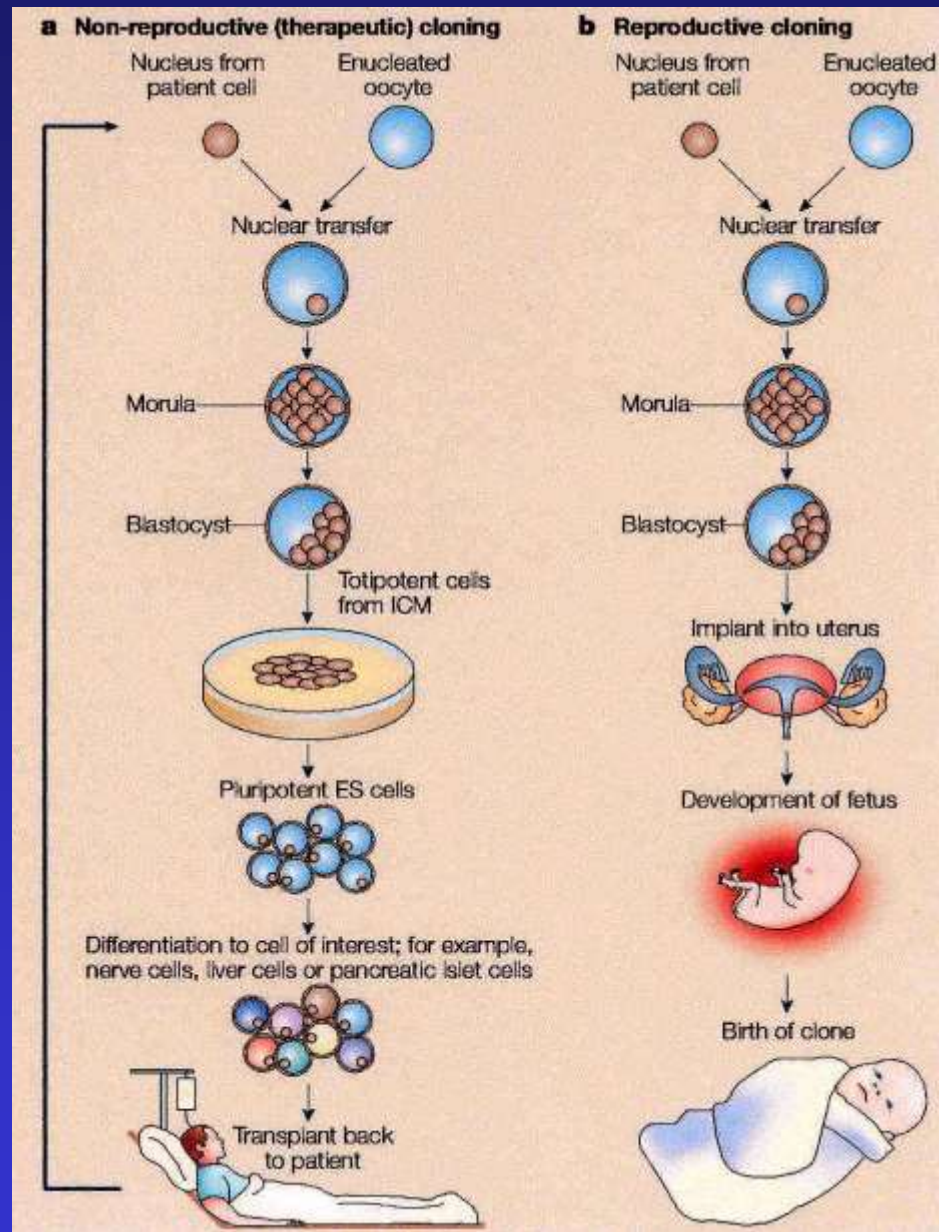
ACT President Michael West

CLONING - POTENTIAL BENEFITS

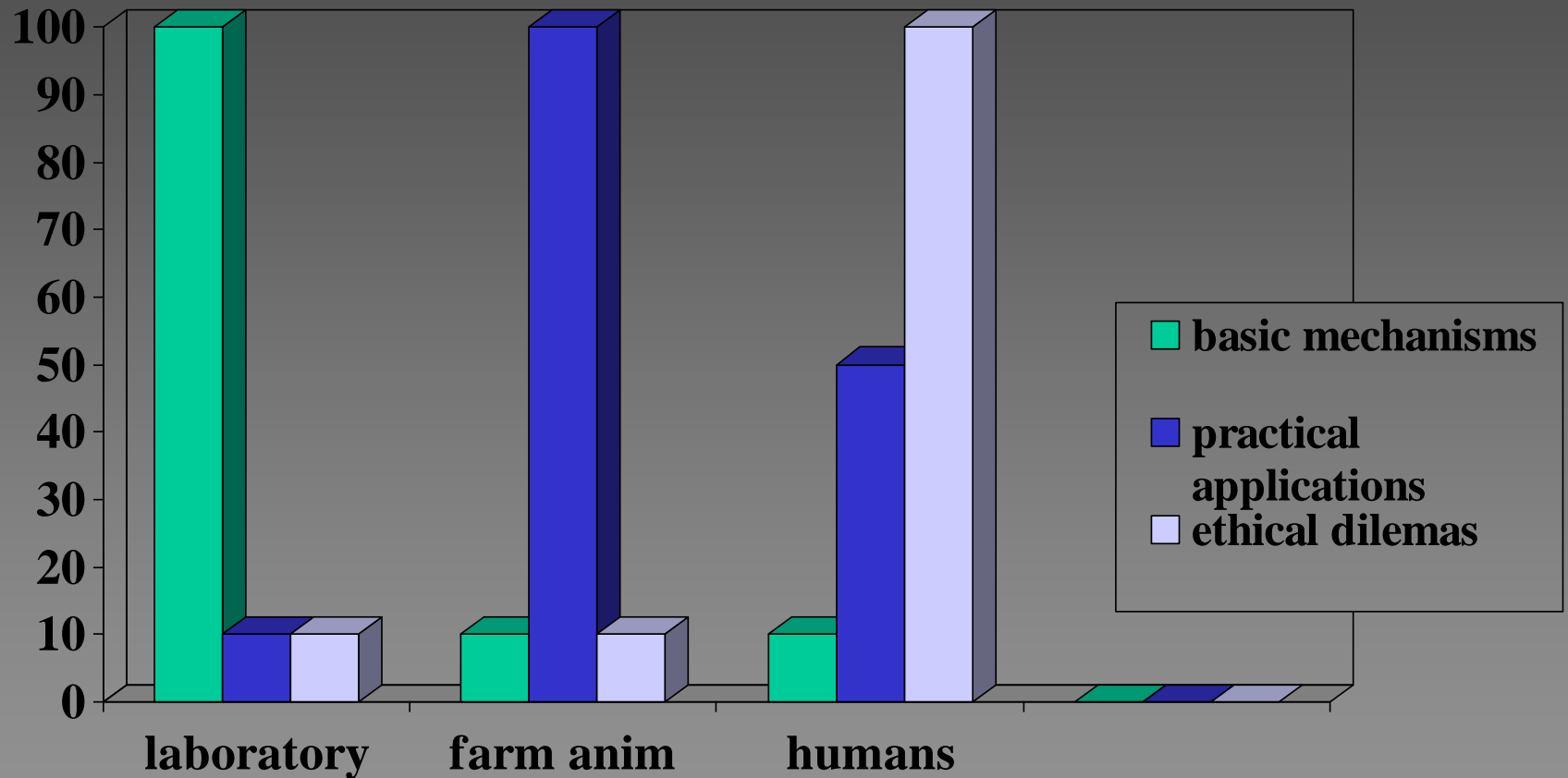


Trounson, A. ;
MJA 167:568-569
; 1997

Reproductive versus non-reproductive cloning



CLONING AND EMBRYONIC STEM CELLS



Solter, D. ; Croatian Medical
Journal 40(3):309-318 ; 1999

J. Fulka Jr.

Inst. of Animal Production, Prague, Czech Rep.

Heyman Y., Chesne P., Renard J.-P., Fléchon J.-F., Duranthon V.

**Biologie Cellulaire et Moléculaire, INRA, 78352 Jouy-en-Josas Cedex,
France**

H. Callesen, P. Holm, S. D. Smith, E. Soloy

**Embryo Technology Center, Danish Institute of
Animal Science, DK-8830 Tjele, Denmark**

P. Hyttel

**Royal Veterinary and Agricultural University, Section for Reproduction,
DK-1870 Frederiksberg C, Copenhagen, Denmark**

Ouhibi N., Moor R. M.

**Laboratory of Protein Function, The Babraham Inst., Cambridge CB2 4AT,
UK**