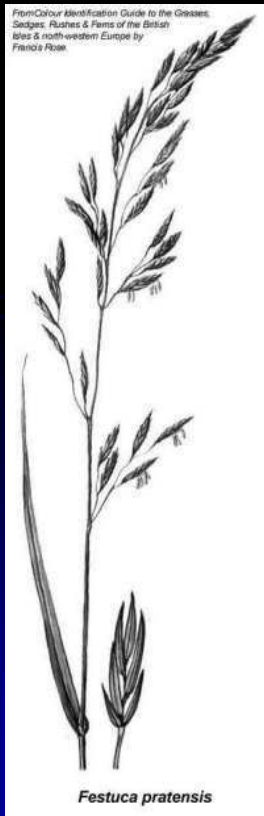


# Genome mapping within the *Festuca-Lolium* complex



David Kopecký

Laboratory of Molecular Cytogenetics and Cytometry  
Institute of Experimental Botany  
Academy of Sciences of Czech Republic

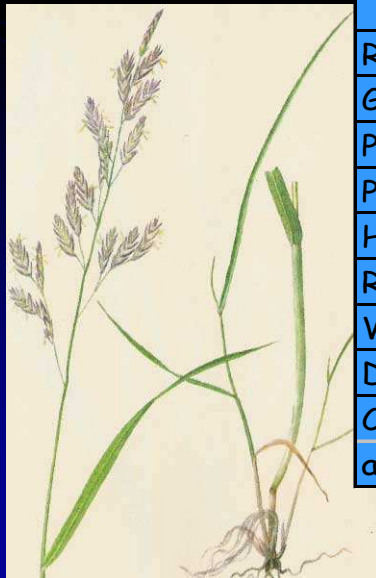
# Outline

## DArTFest

- Development of the array
- Diversity study
- Genetic mapping
- Physical mapping
- Integration of the maps
- Characterization of Festulolium hybrids
- Conclusions and future work

# FESTUCA - LOLIUM COMPLEX

- Ones of the most important forage and turf species
- Cross of the species of both genera results in fertile hybrids
- Hybrids display agronomically important attributes of both genera
- Chromosomes of both genera frequently pair and recombined with each other
- Chromosomes of both genera can be distinguished by GISH



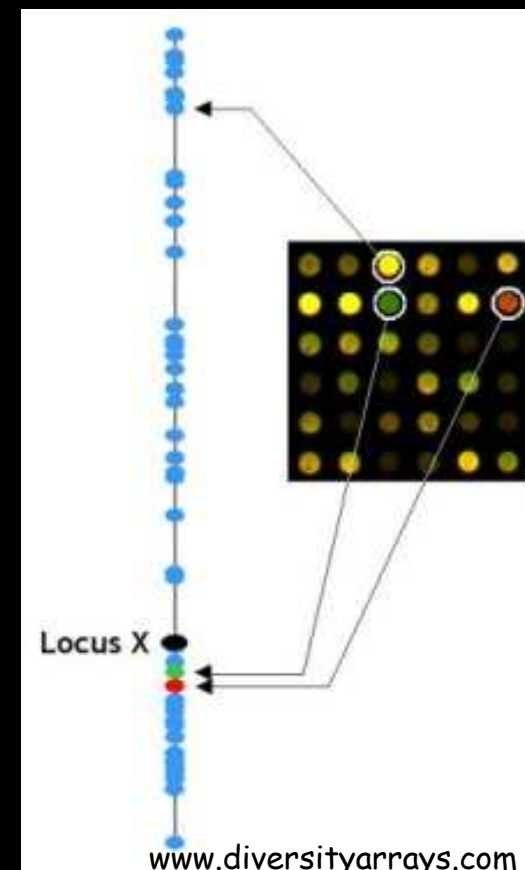
	Lm	Lp	Fp	Fa
Rapid establishment from seed	+	+/-	-	-
Good production in seeding year	+	+/-	-	-
Palatability	+	+/-	+	-
Persistency	-	+	+	+
High tillering density	-	+	-	-
Resistance to treading	-	+	-	-
Winter hardiness	-	+/-	++	+
Drought tolerance	-	-	+	++
Continued high production after second harvest year	-	-	+	+



# DArT technology

## Diversity Array Technology

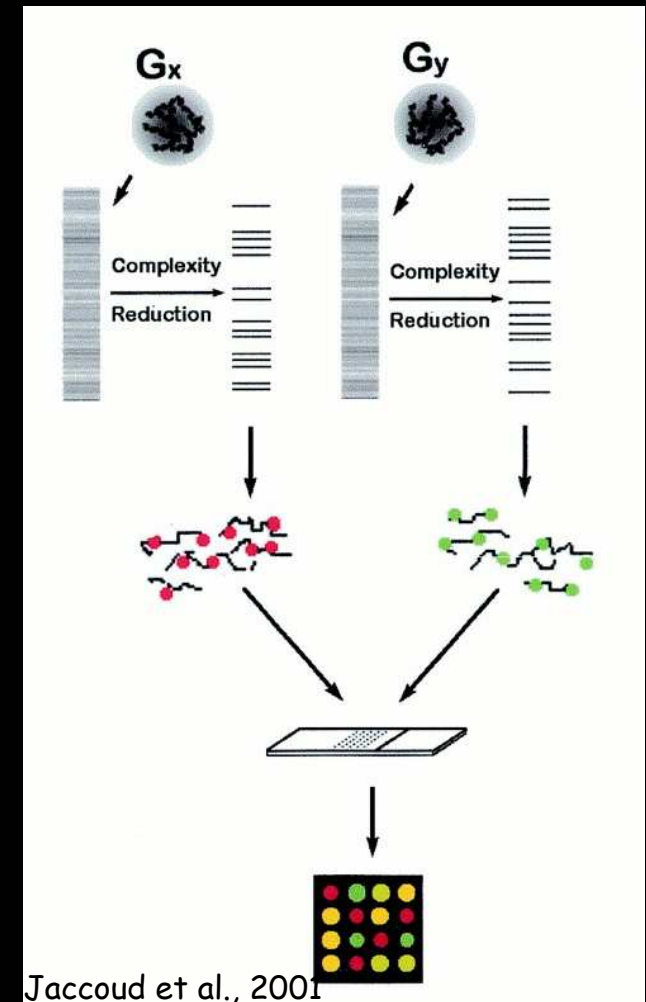
- Developed by Dr. Andrzej Kilian (Canberra, Australia) (Jaccoud et al., 2001)
- microarray technology
- dominant markers
- positives: high number of markers, anonymous markers (can be sequenced)
- negatives: low polymorphism in some species



# DArT technology

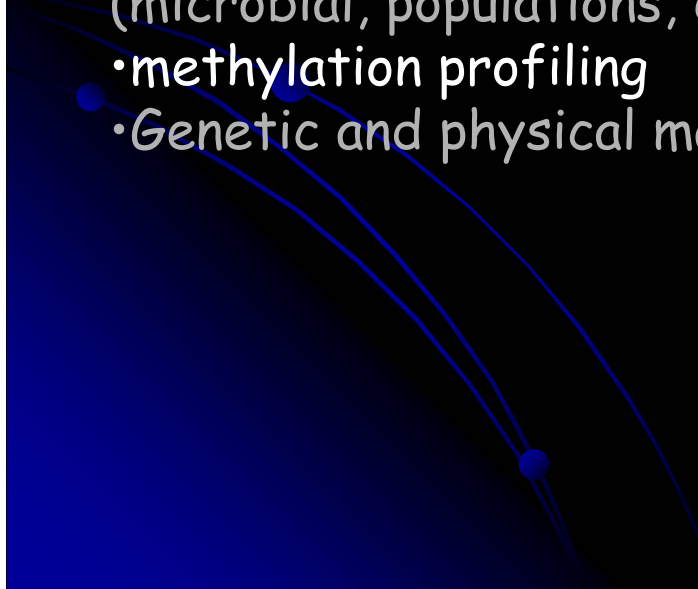
DArT technology consists of several steps:

- Complexity reduction of DNA
- Library creation
- Microarray from libraries
- Hybridization of fluoro-labelled DNA
- Scanning of slides for hybridization signal (confocal laser)
- Data extraction and analysis (DArT software)



# DArT technology

Main use:

- identification of QTL
  - simultaneous marker-assisted selection for several traits
  - evaluation of genetic diversity
  - varietal identification of crops
  - monitoring the composition of complex DNA samples (microbial, populations, etc.) - also in hybrids
  - methylation profiling
  - Genetic and physical mapping
- 

# DArTFest

A first DArT array for the *Festuca-Lolium* complex

Development of array:

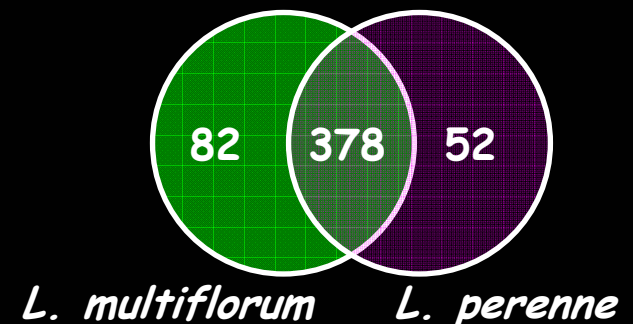
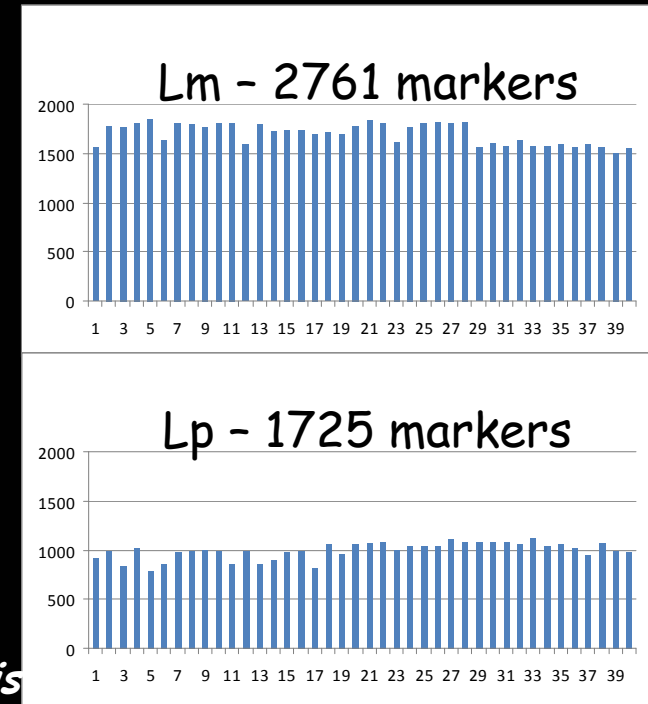
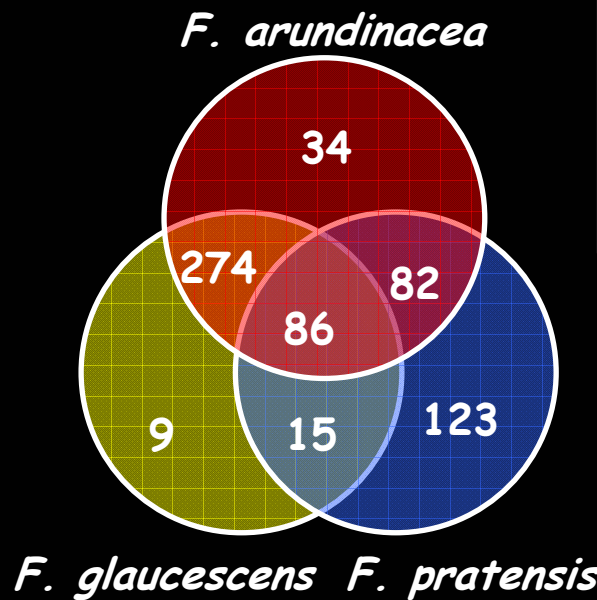
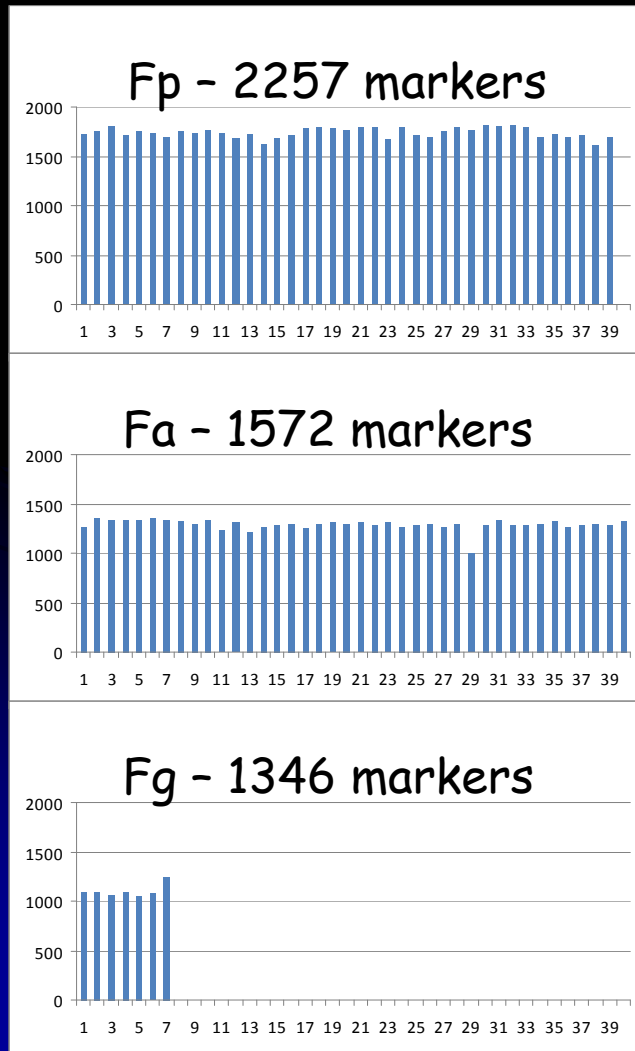
Material: 5 species

- 40 accessions of *Lolium perenne* (2x, 4x)
- 40 accessions of *L. multiflorum* (2x, 4x)
- 40 accessions of *Festuca pratensis* (2x, 4x)
- 40 accessions of *F. arundinacea* (6x)
- 7 accessions of *F. glaucescens* (4x)

Array: 7680 probes

Number of polymorphic markers: 3884

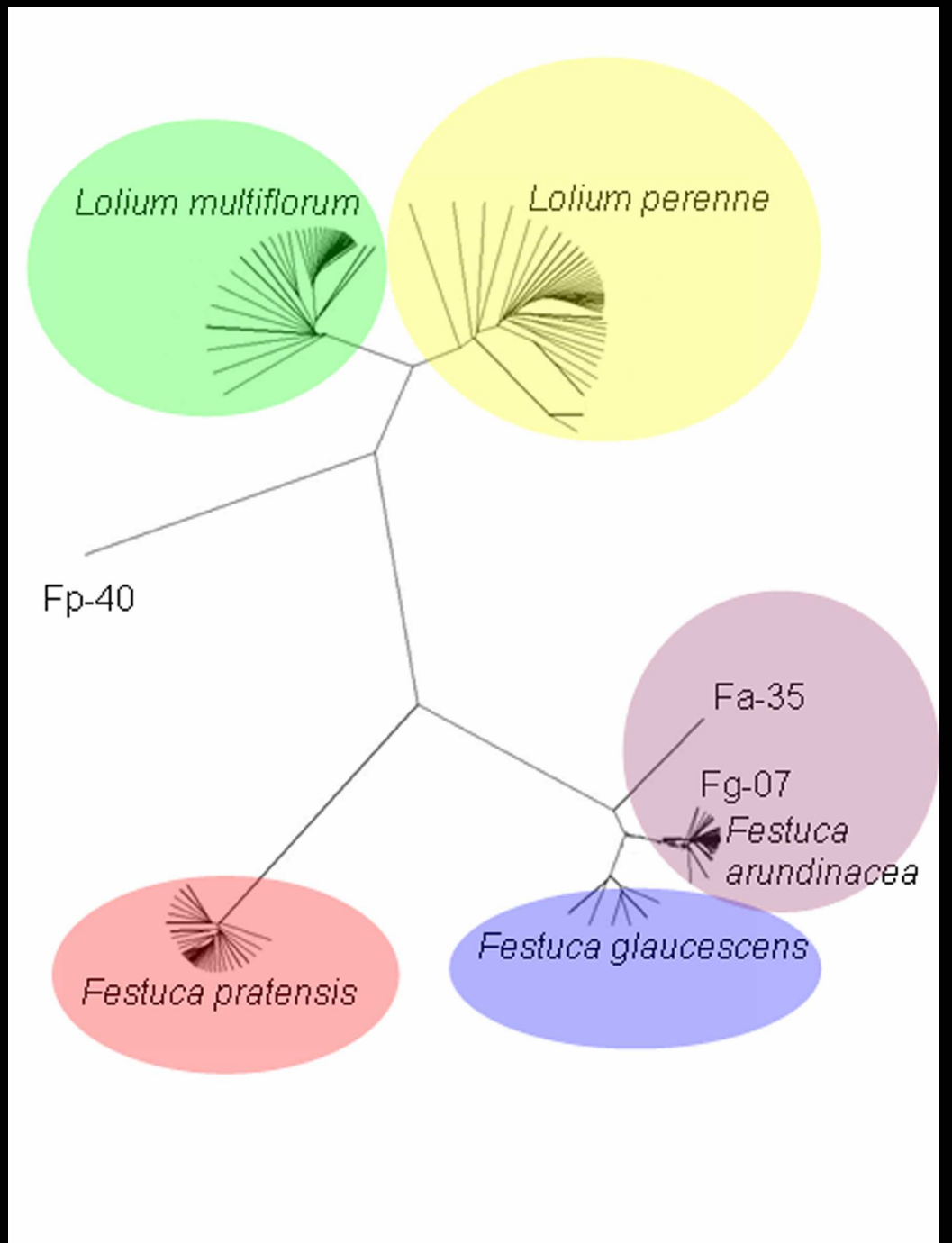
# DArTFest: Genus- and species-specific markers



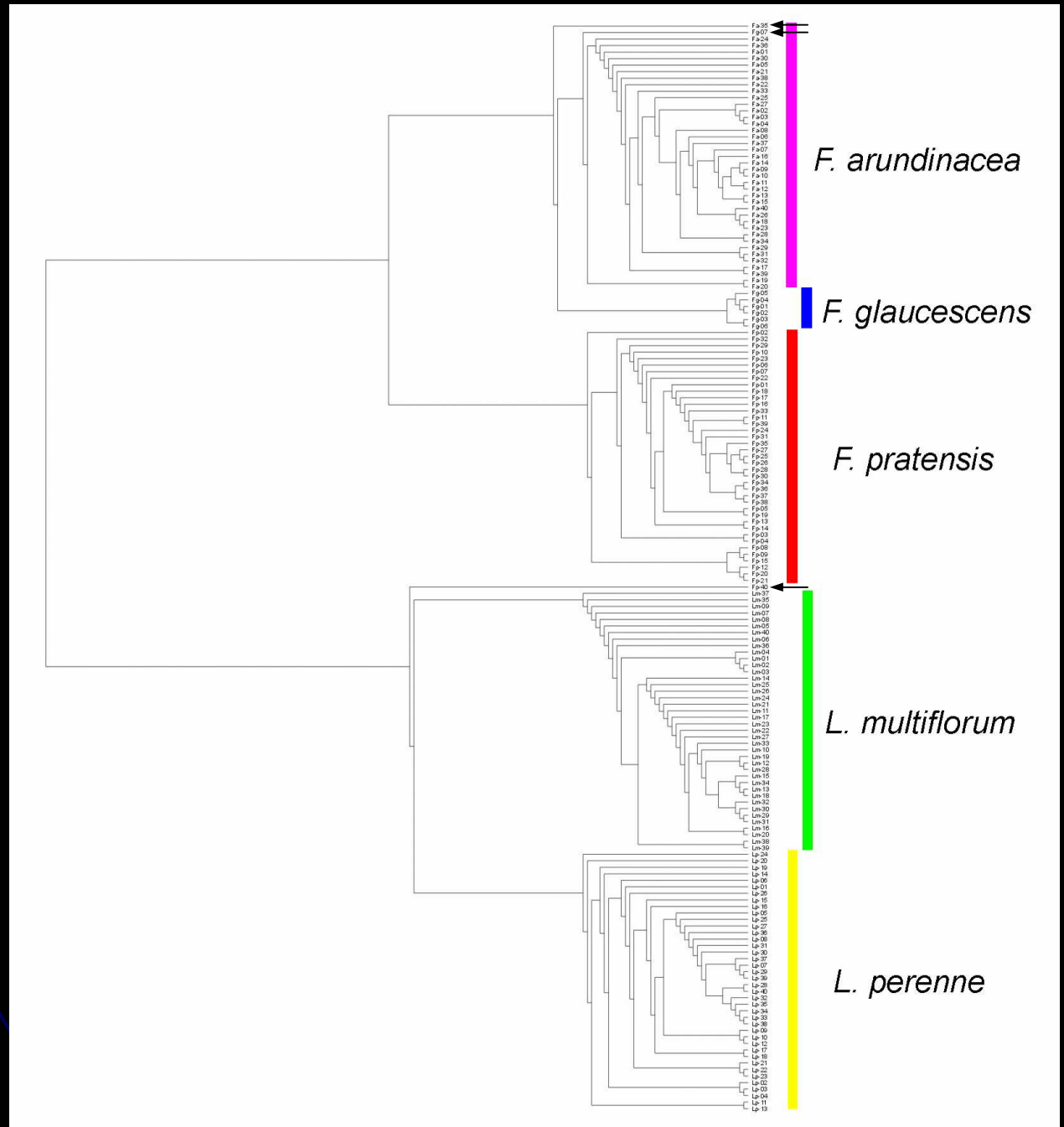


# DArT*Fest*: Genetic diversity

Based on the analysis of 2637  
DArT markers



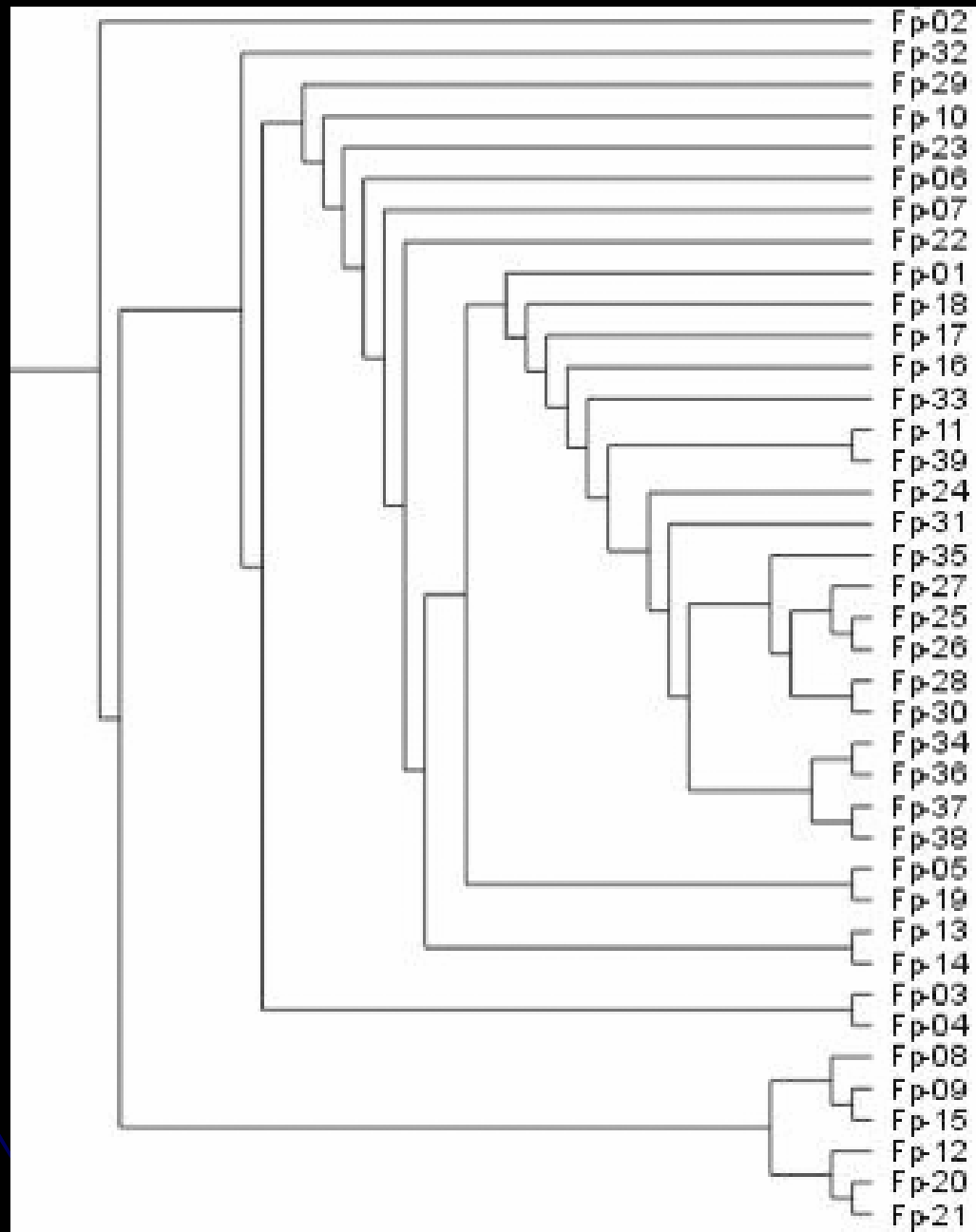
# DArTFest: Genetic diversity





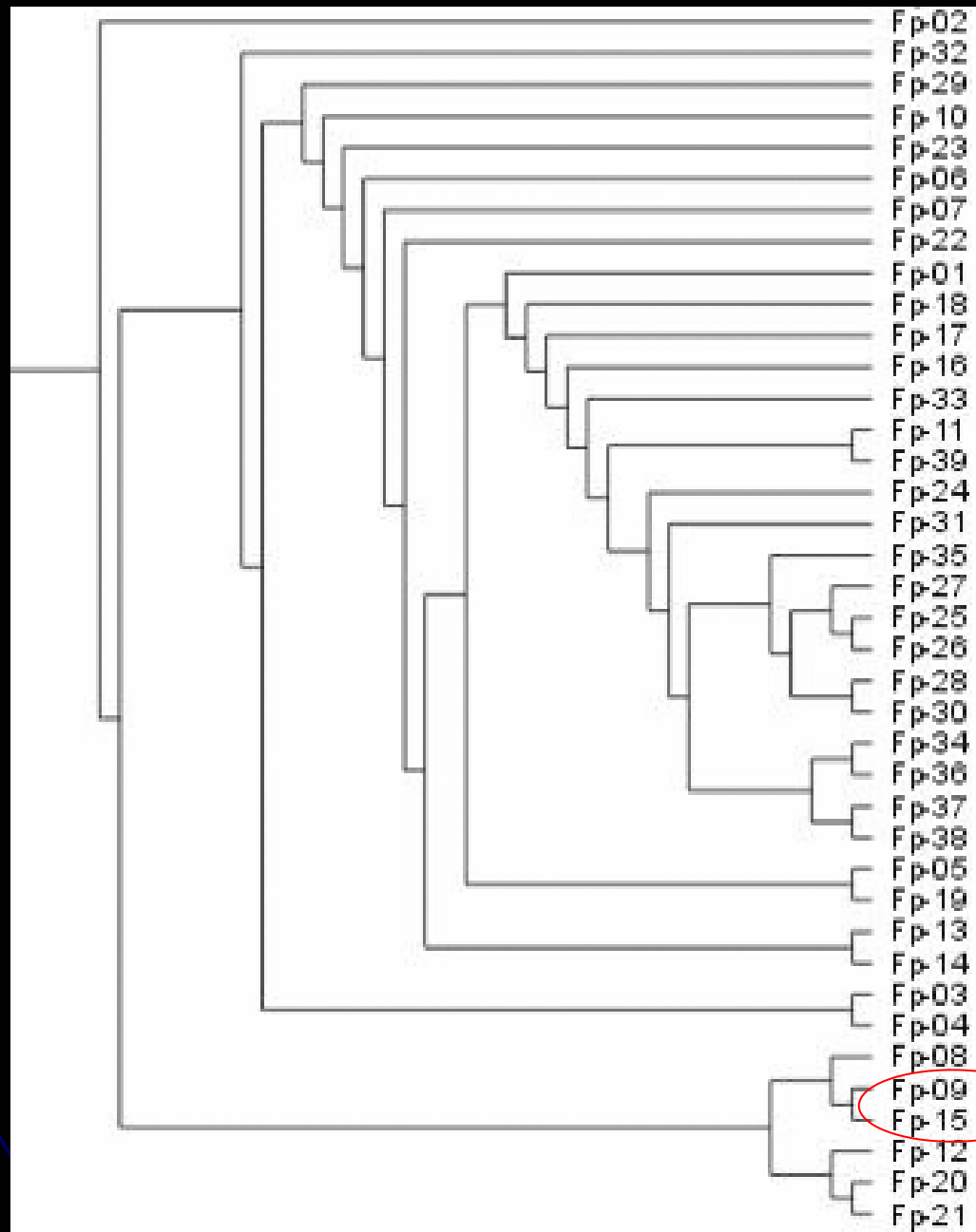
# DArTFest: Genetic diversity

(*F. pratensis*)



# DArT*Fest*: Genetic diversity

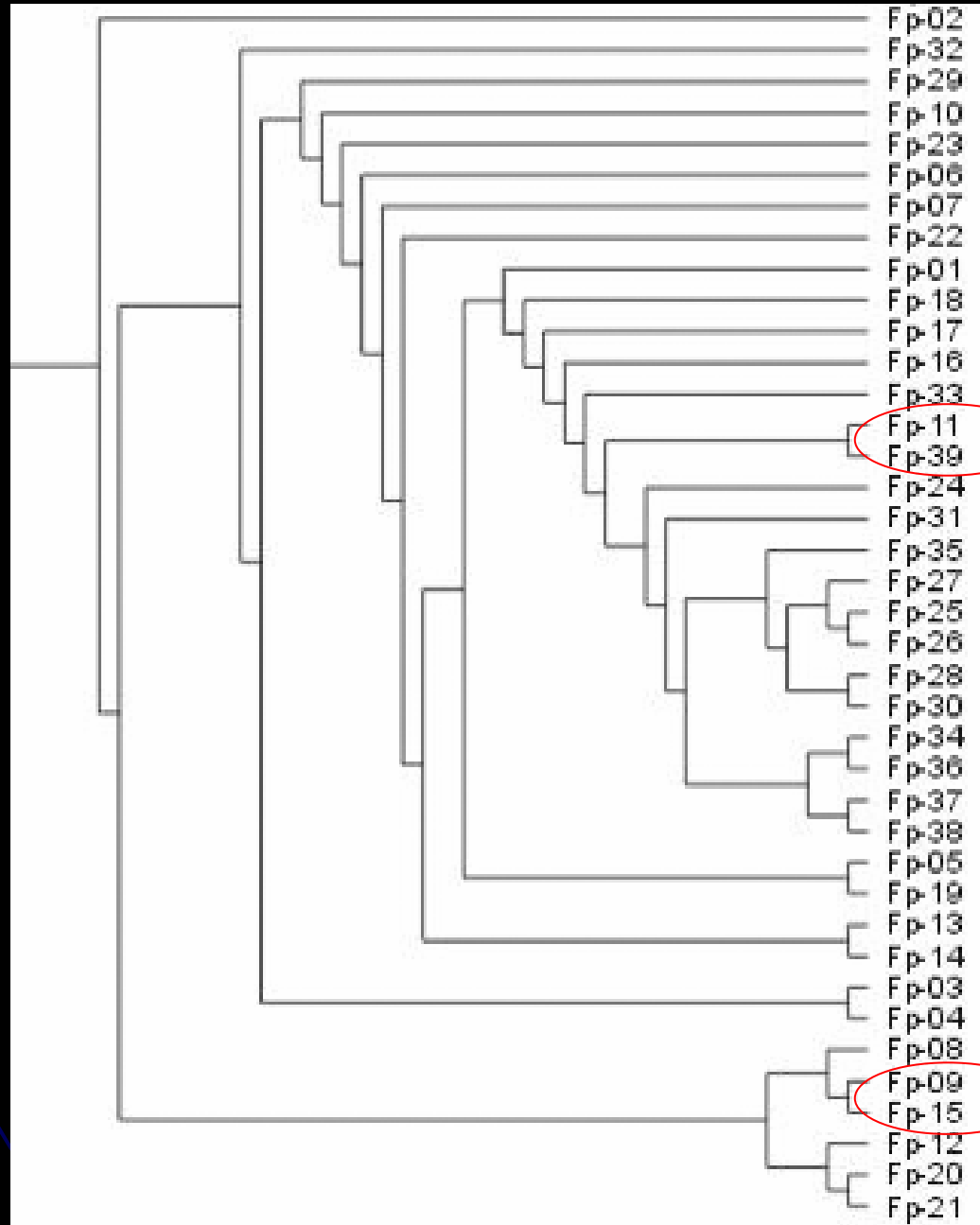
(*F. pratensis*)



Ecotypes  
from  
Lithuania  
(Acc. 3982  
and 3985)

# DArTFest: Genetic diversity

(*F. pratensis*)

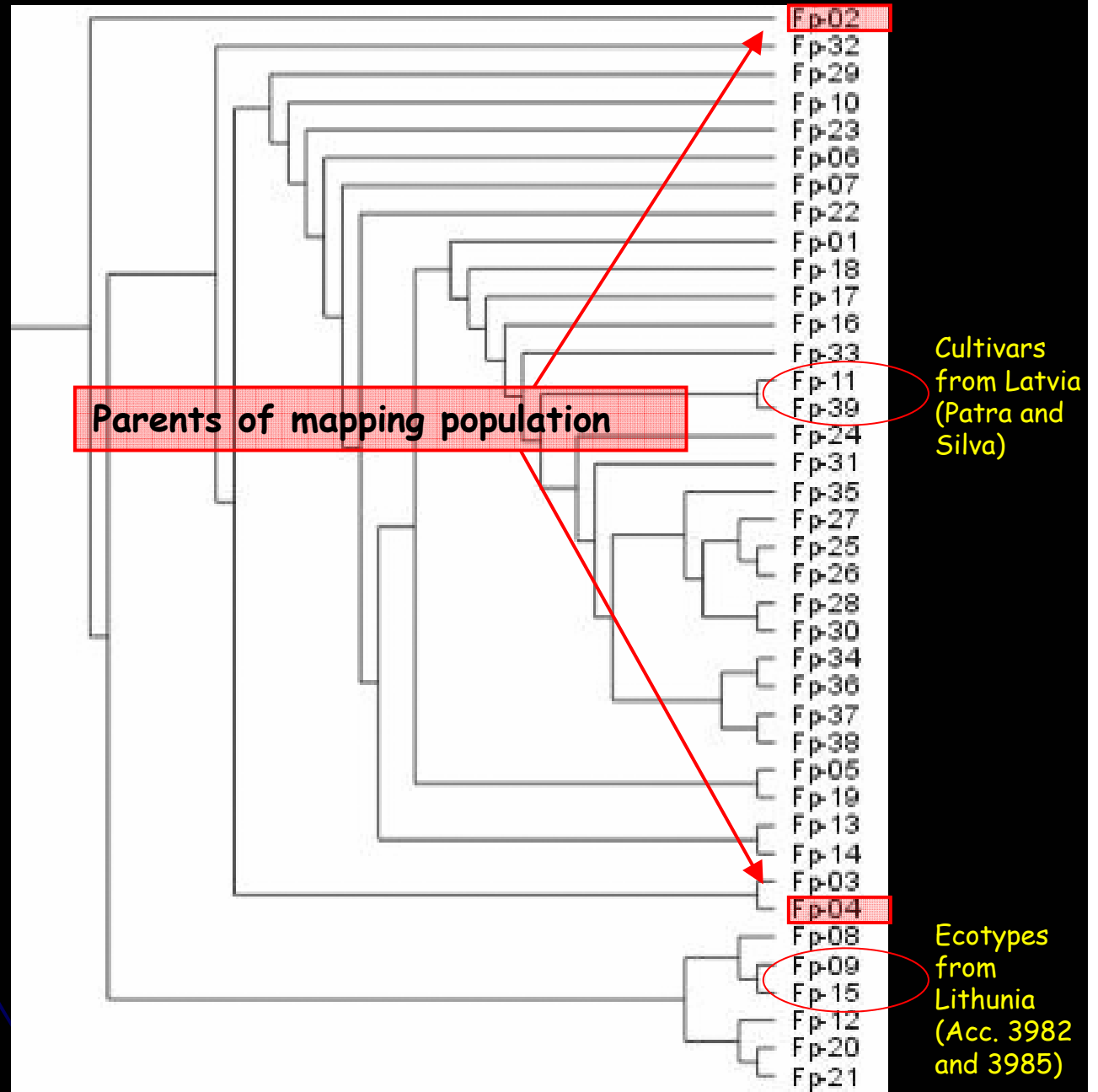


Cultivars  
from Latvia  
(Patra and  
Silva)

Ecotypes  
from  
Lithuania  
(Acc. 3982  
and 3985)

# DArTFest: Genetic diversity

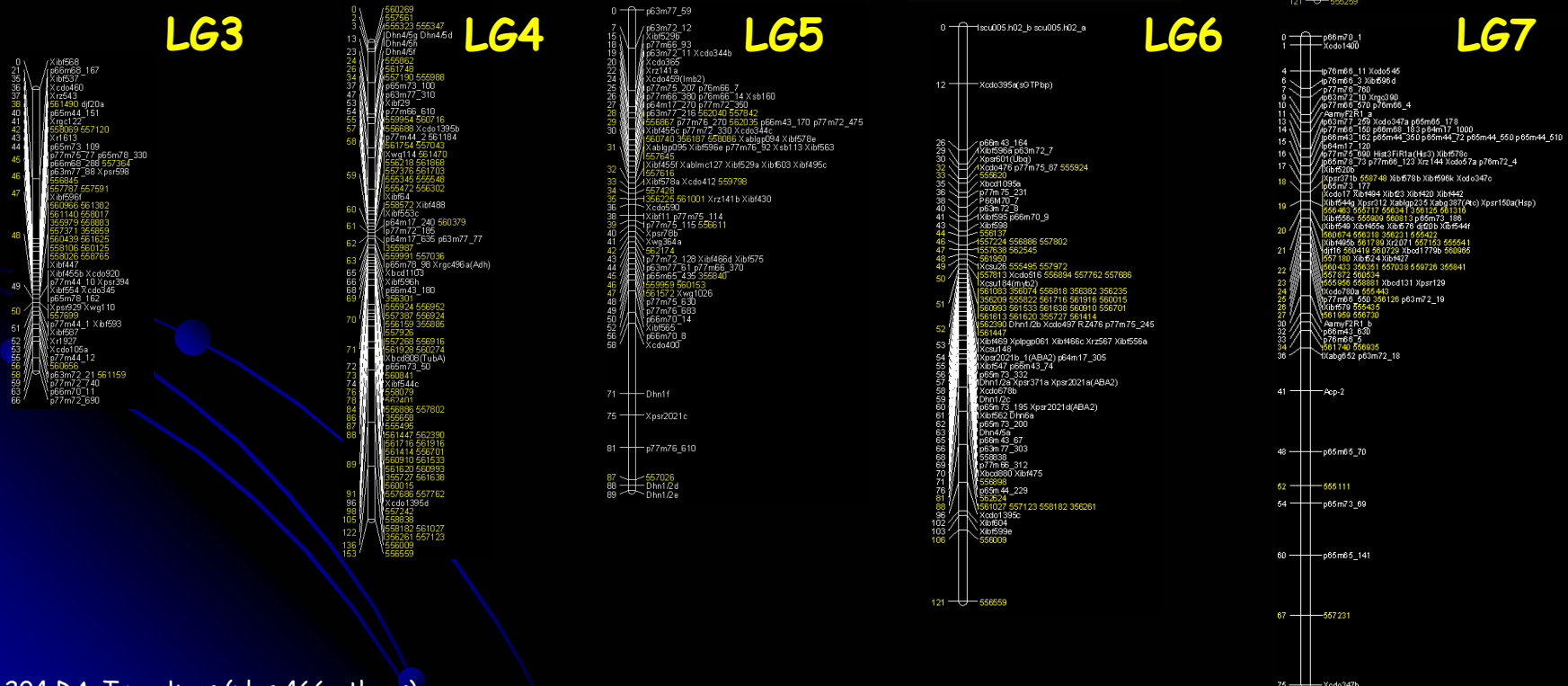
(*F. pratensis*)



# DArTFest:

## Genetic mapping

(*Festuca pratensis* - Odd Arne Rognli & Simen Rød Sandve)



- 204 DArT markers (plus 466 others)
- The total length of genetic map 775cM (DArT genetic map by itself - 507cM; the original map - 605cM)

DArT markers  
other markers



# DArTFest: Genetic mapping

(*Lolium multiflorum* - Roland Kölliker)

In progress



# DArTFest: physical mapping

Development of materials for the mapping

$$F_p (2n=2x=14) \times L_m (2n=4x=28)$$



$$F_1: (2n=3x=21, F_p L_m L_m) \times L_m (2n=4x=28)$$



$$F_2: (2n=4x=28, F_p L_m L_m L_m) \times L_m (2n=4x=28)$$

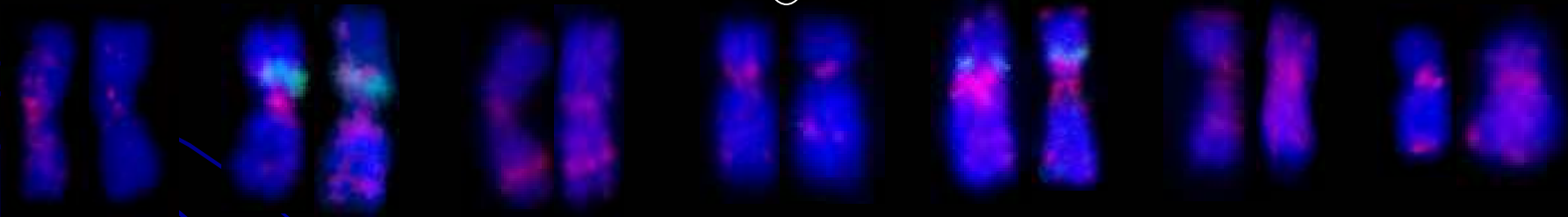
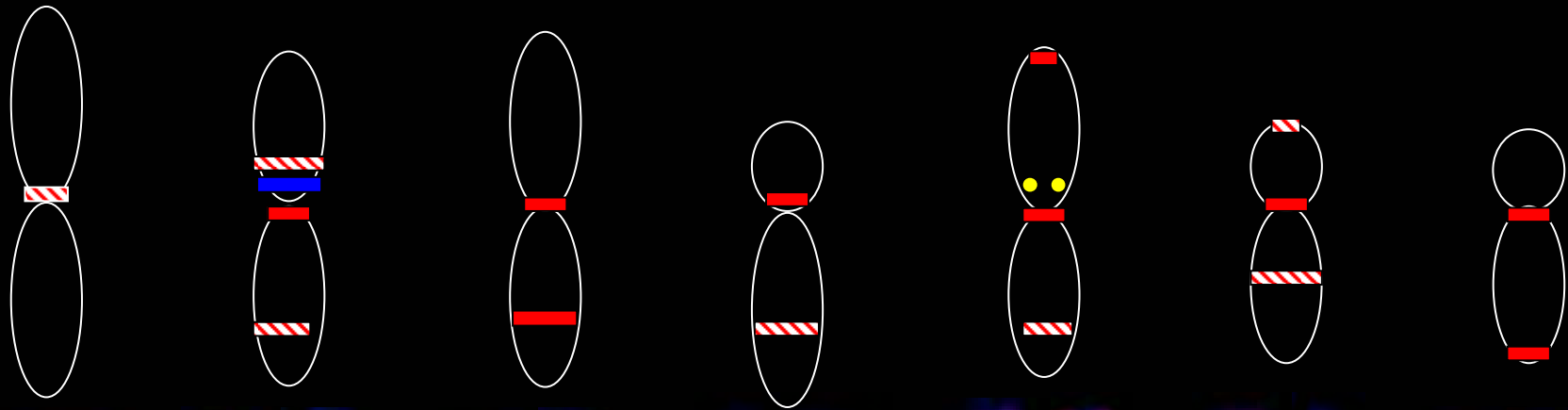


$$F_3 \text{ etc. } (2n=4x=28; 27L_m+1F_p)$$

L<sub>m</sub> L<sub>m</sub> L<sub>m</sub> F<sub>p</sub>



# Cytogenetic mapping in *Festuca pratensis*



Chromosome 4	Chromosome 3	Chromosome 7	Chromosome 5	Chromosome 2	Chromosome 6	Chromosome 1
Chromosome A	Chromosome N	Chromosome C	Chromosome D	Chromosome B	Chromosome F	Chromosome G

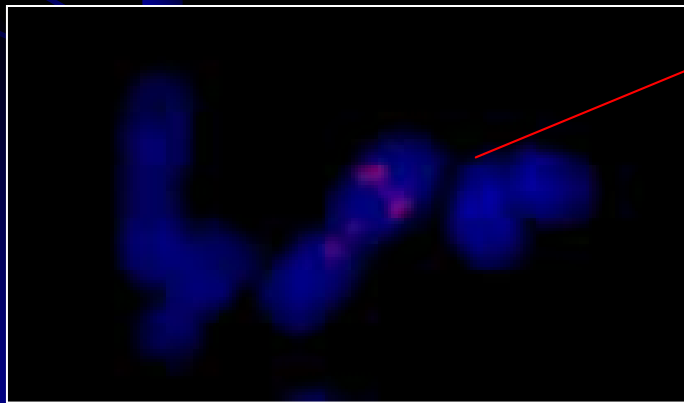
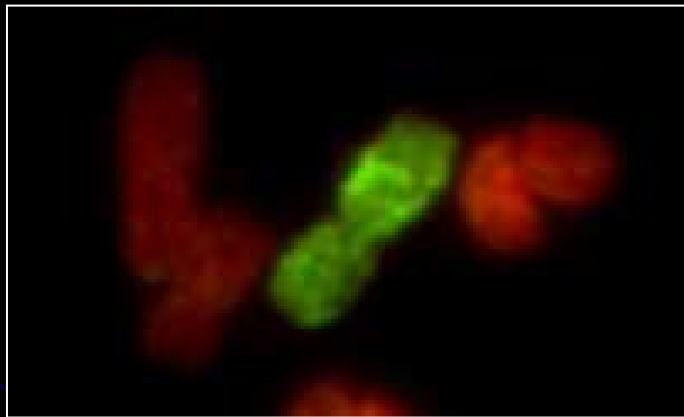
— BAC clone 1G18

— BAC clone 1G18 (weak signal)

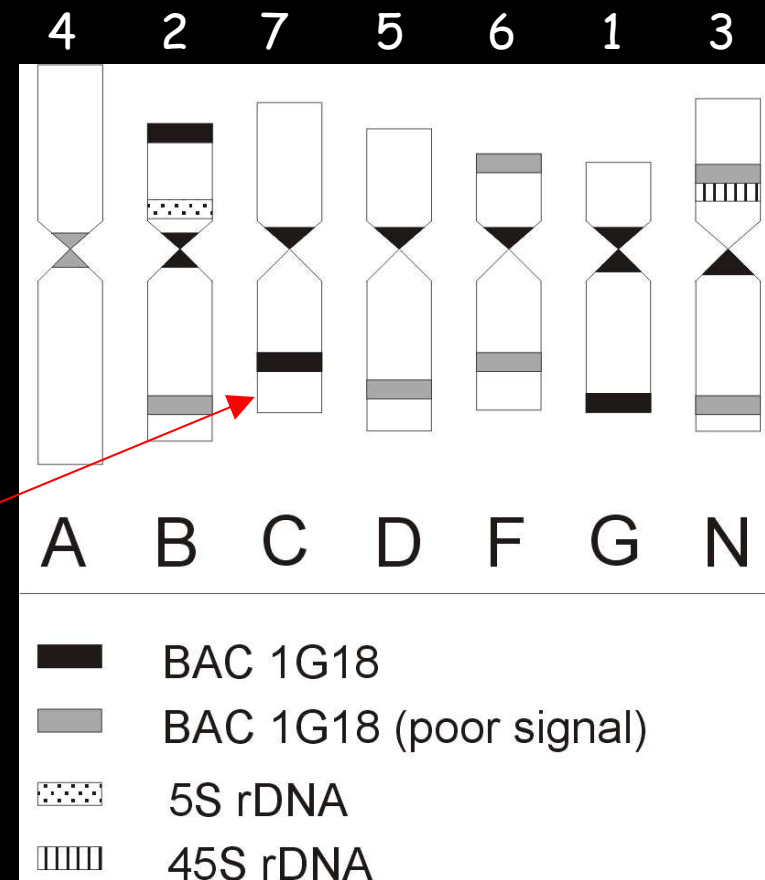
— 45S rDNA

•• 5S rDNA

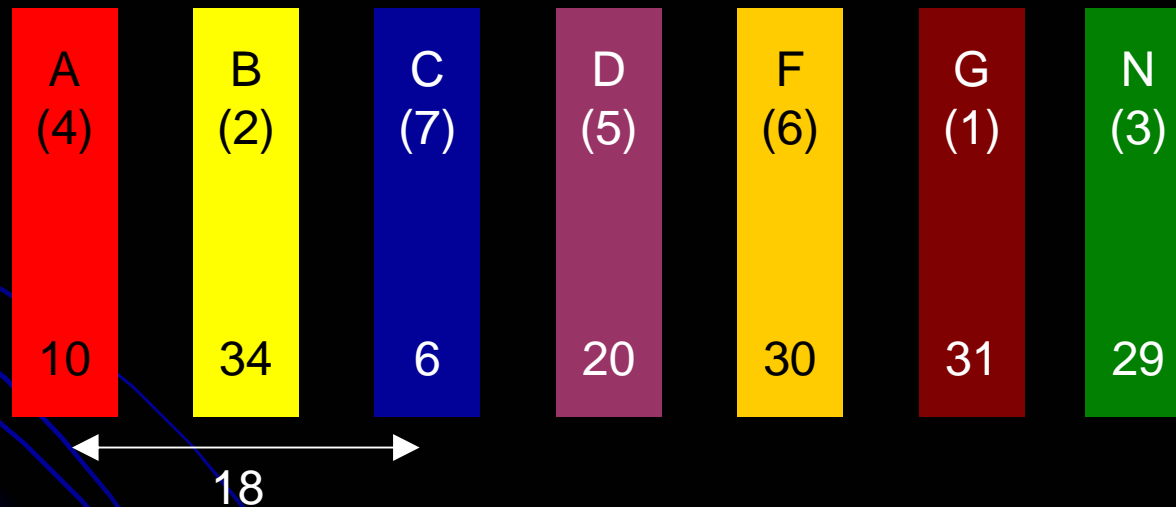
# Cytogenetic mapping in *Festuca pratensis*



Chromosome C



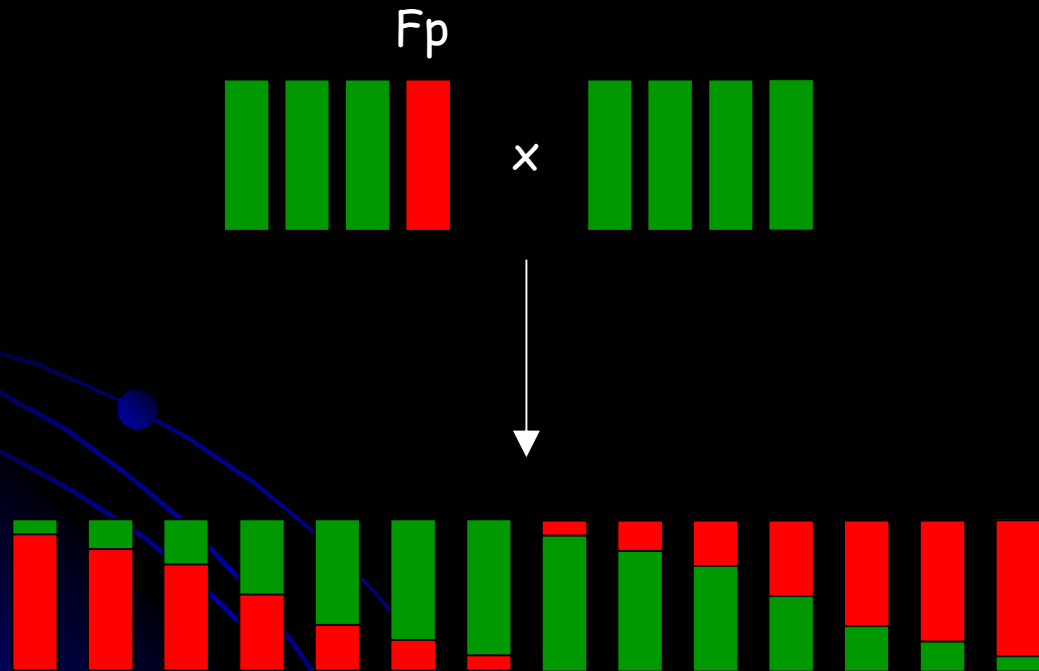
# DArT*Fest*: Anchoring DArT markers to individual Fp chromosomes



=160 markers (out of 288 *F. pratensis* positive and *Lolium* negative)

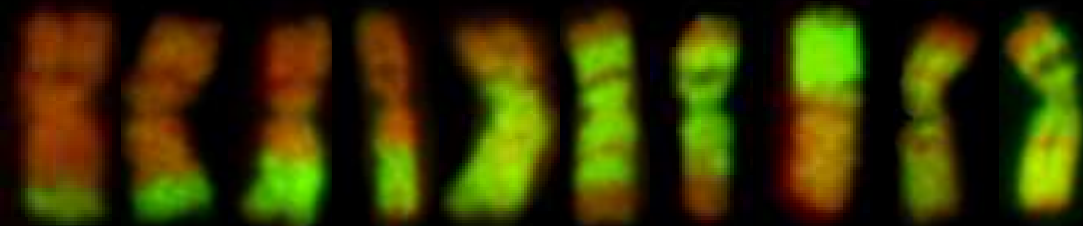
# DArT*Fest*: physical mapping

- Mapping to subchromosomal regions (bins)
- Development of recombination lines



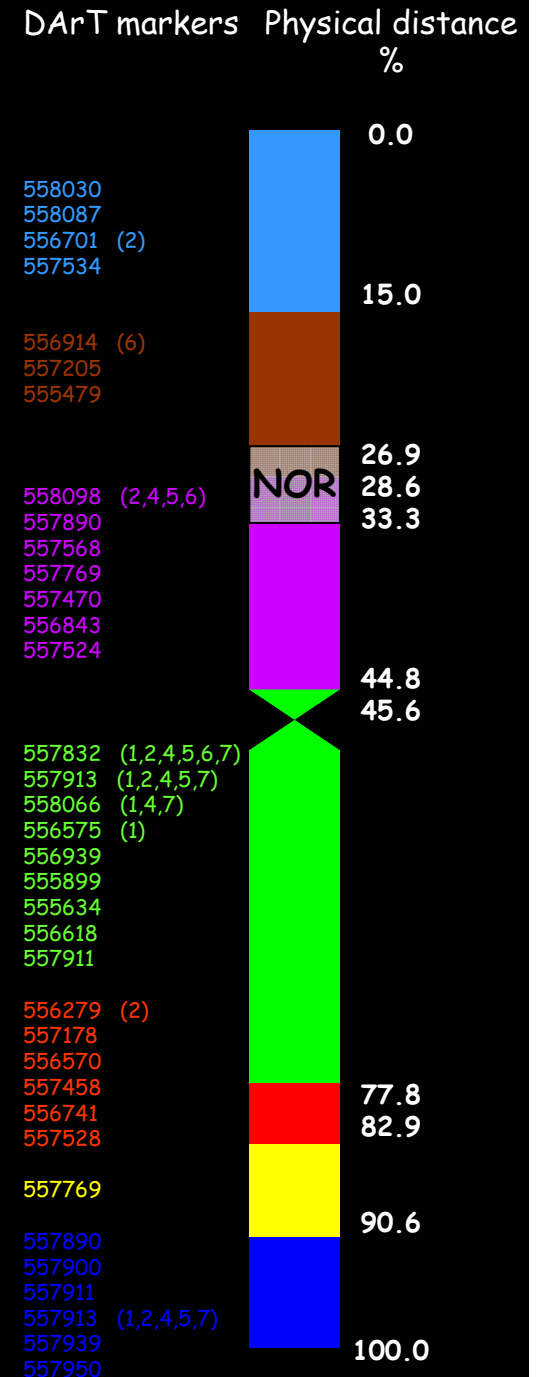
- More than 30 lines have been developed for each  $F_p$  chromosome

# DArT*Fest*: Anchoring DArT markers to Fp chromosome bins



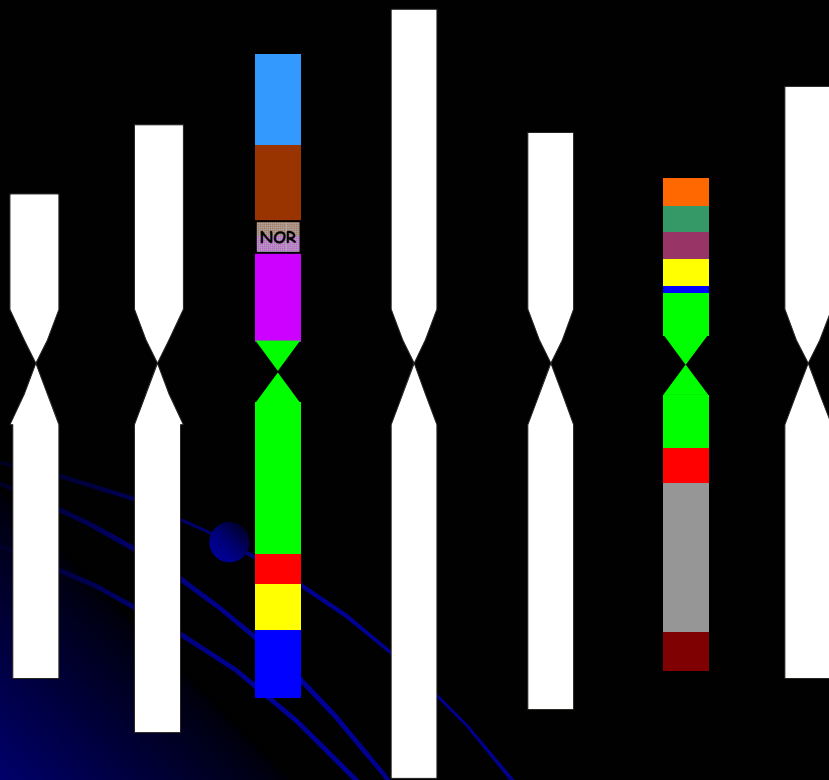
Chromosome 3 of *F. pratensis*

- L. multiflorum*
- F. pratensis*



\*) detected in other chromosomes

# DArT*Fest*: Anchoring DArT markers to Fp chromosome bins

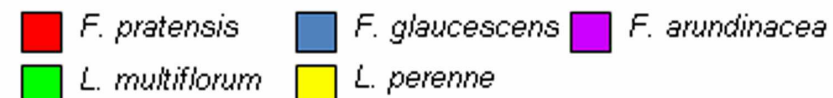
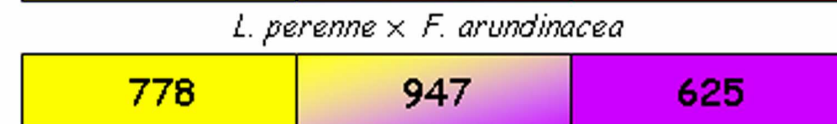
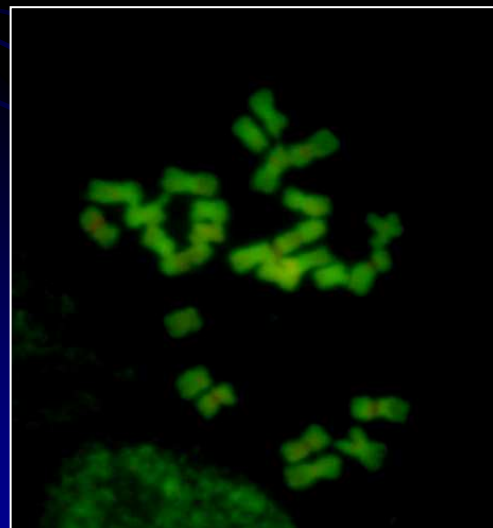
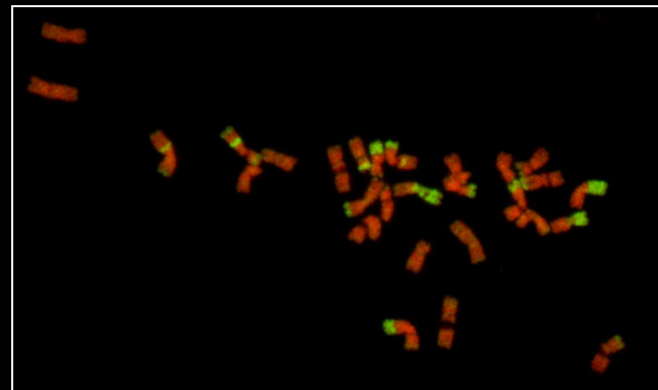


We expect to have about 10 bins per chromosome = dissect *F. pratensis* genome to about 70 bins



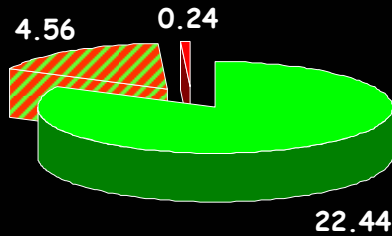


# DArTFest in hybrids

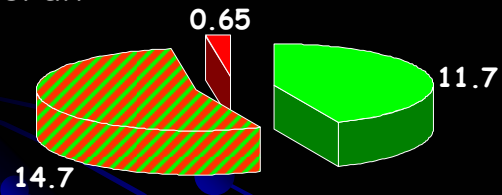


# DArT*Fest* in hybrids

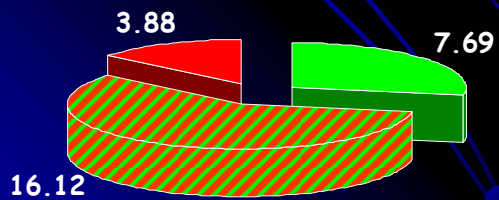
Spring Green



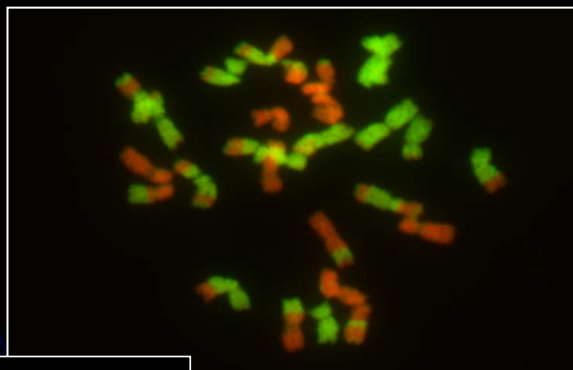
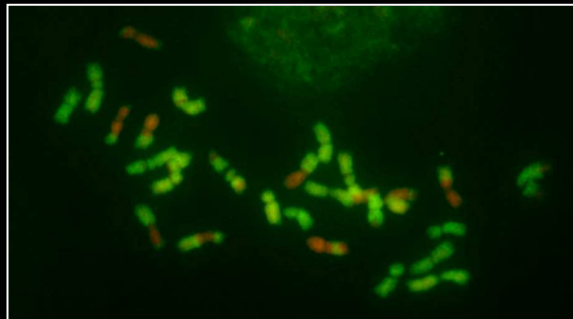
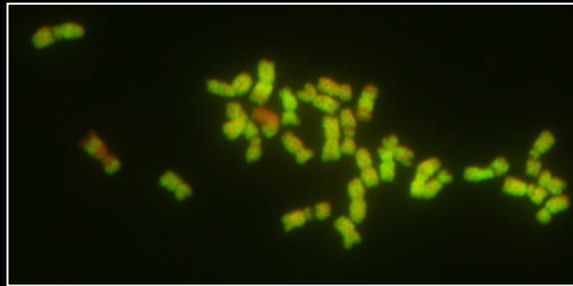
Perun



Elmet



GISH



DArT

Lm

Fp

468

65

470

342

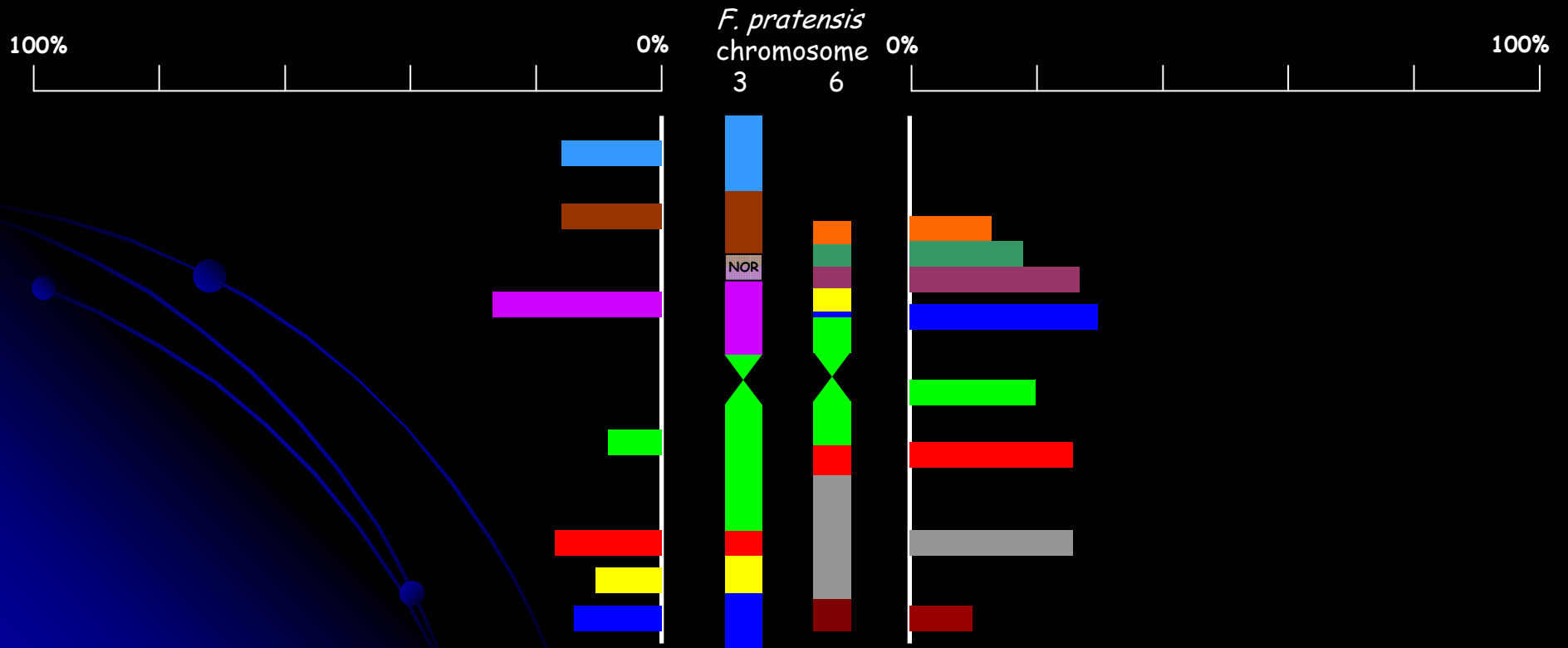
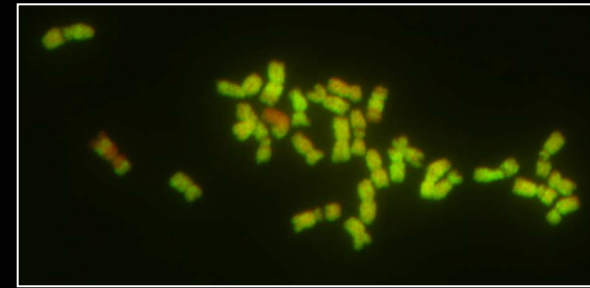
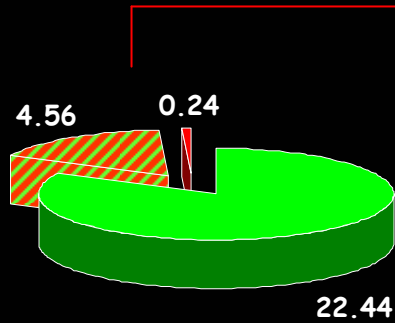
359

410

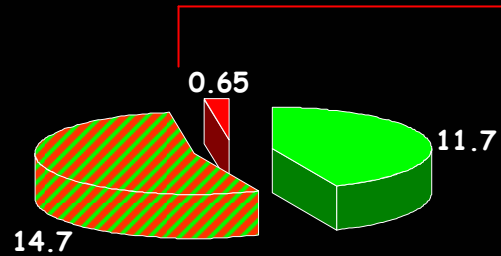
■ *L. multiflorum* ■ Recombination LmxFp ■ *F. pratensis*

# DArTFest in hybrids

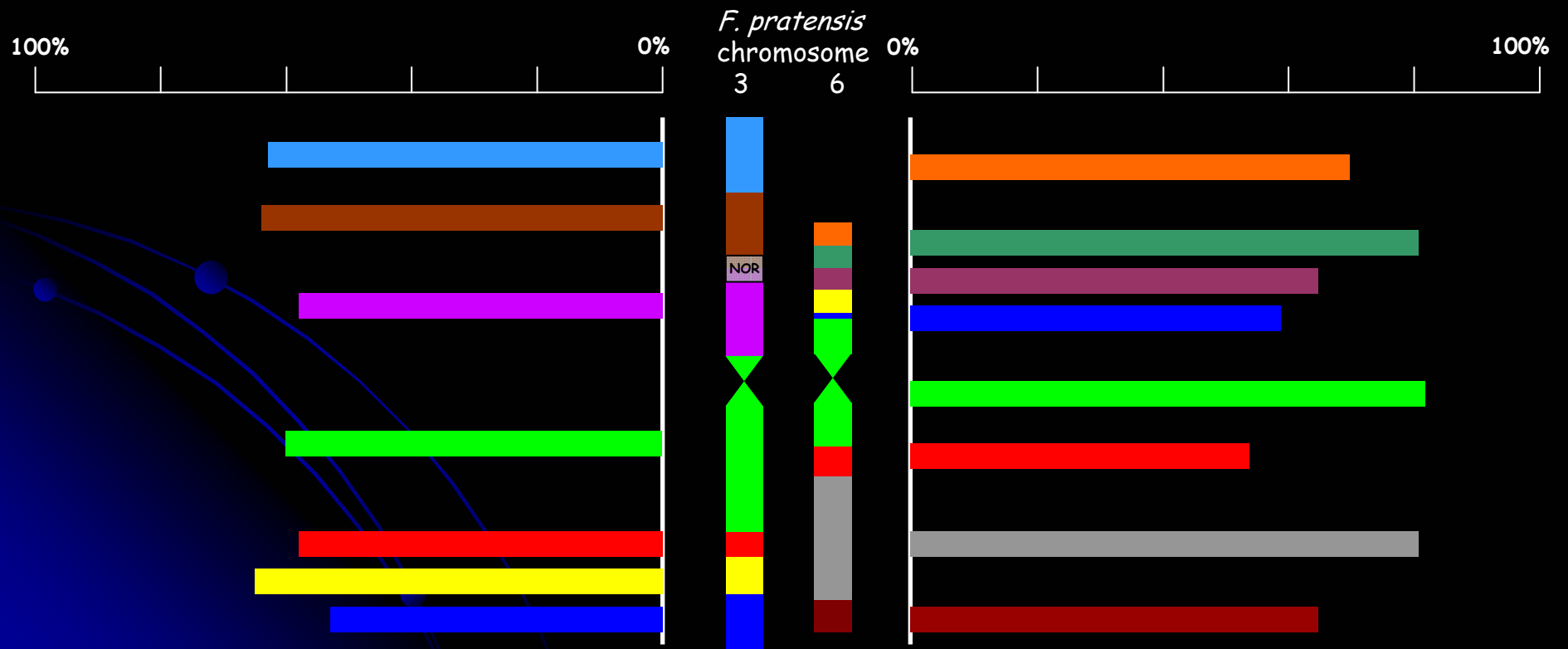
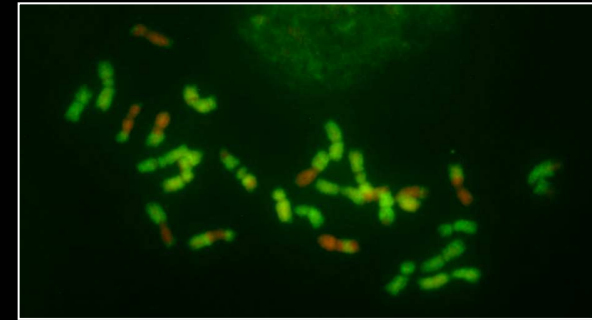
cv. Spring Green



# DArTFest in hybrids

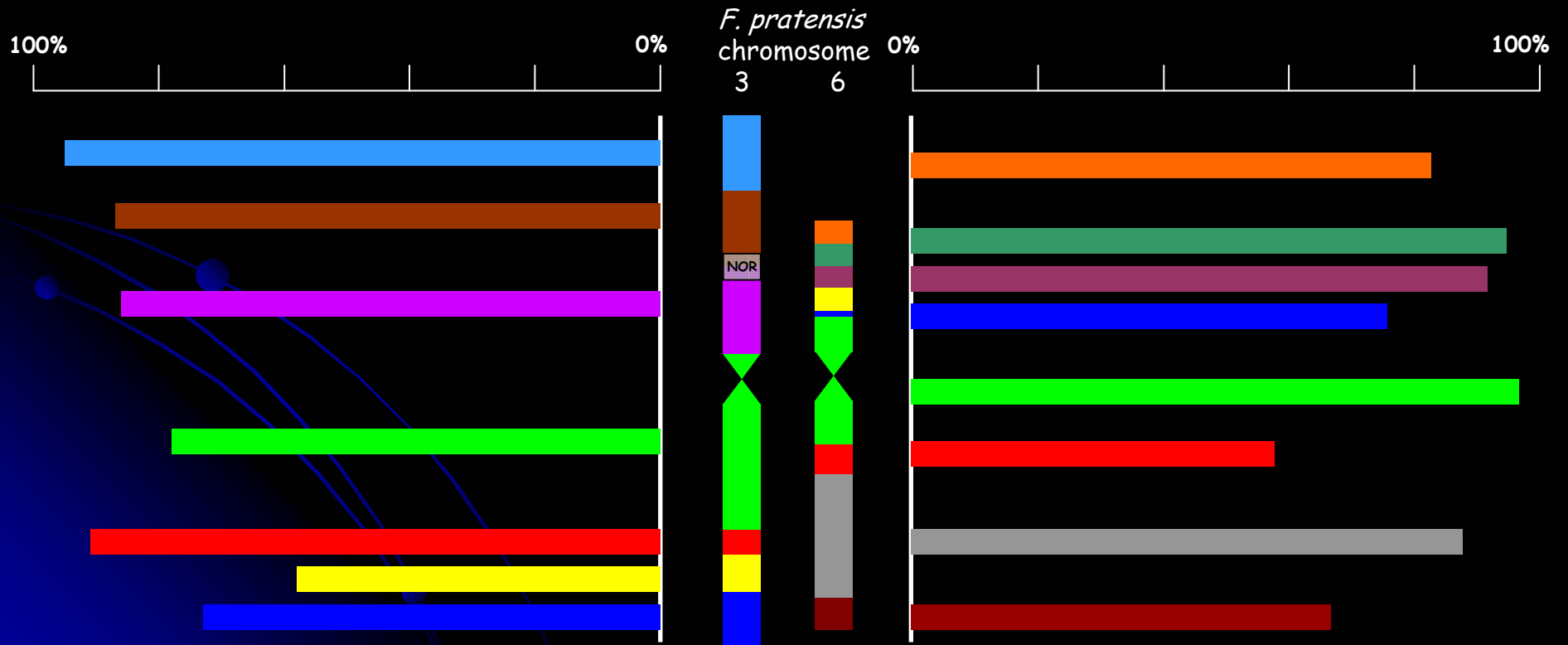
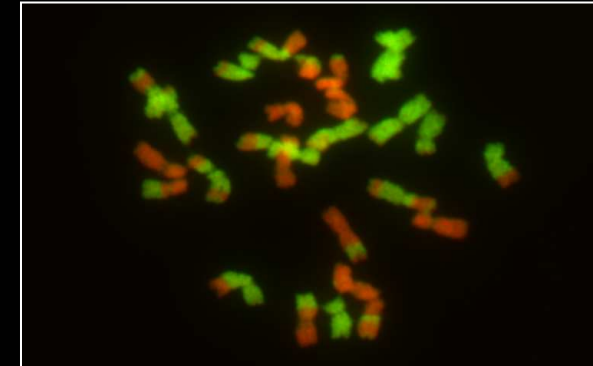
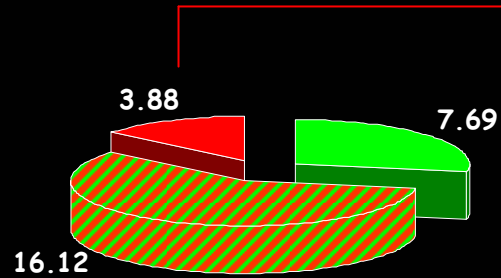


cv. Perun



# DArTFest in hybrids

cv. Elmet



# Conclusions

We have developed the first DArT array (*DArTFest*) for *Festuca-Lolium* complex

The array contains 3884 markers and can be used for:

- Analysis of genetic diversity
- Genotyping
- Analysis of genomic constitution of hybrids
- Genetic and physical mapping

# Future work

- (1) Establishment of genetic maps with DArT markers
- (2) Integration of genetic and bin maps
- (3) Comparative analysis of species within the *Festuca-Lolium* complex
- (4) Association of markers with agronomically important traits
- (5) Sequencing of already mapped DArT markers and conversion to PCR markers



# Acknowledgements

Institute of Experimental Botany

Prof. Jaroslav Doležel

Dr. Jan Bartoš

Diversity Arrays Technology

Canberra, Australia

Dr. Andrzej Kilian

Dr. Helen Blois

Dr. Vanessa Caig

Department of Botany and Plant Sciences

University of California, Riverside, USA

Prof. Adam J. Lukaszewski

Dr. James H. Baird

Šlechtitelská stanice Hladké Žitovice

Dr. Vladimír Černoč

Agroscope Reckenholz Tanikon Research

Station ART

Zurich, Switzerland

Dr. Roland Kölliker

Norwegian University of Life Sciences, Aas,

Norway

Prof. Odd-Arne Rognli, Simen Rød Sandve

IBERS, Aberystwyth University, Wales

Prof. Ian P. King, Dr. Leif Skot

ILVO, Merelbeke, Belgium

Dr. Isabel Roldan-Ruiz, Dr. Hilde Muyllé

TEAGASC, Oak Park, Carlow, Ireland

Dr. Susanne Barth

Wageningen UR Plant Breeding, The Netherlands

Dr. Carole Boucoiran, Dr. Oene Dolstra

Aarhus University, Slagelse, Denmark

Dr. Bruno Studer, Dr. Stephan Hentrup

DLF-Trifolium, Roskilde, Denmark

Dr. Niels Roulund

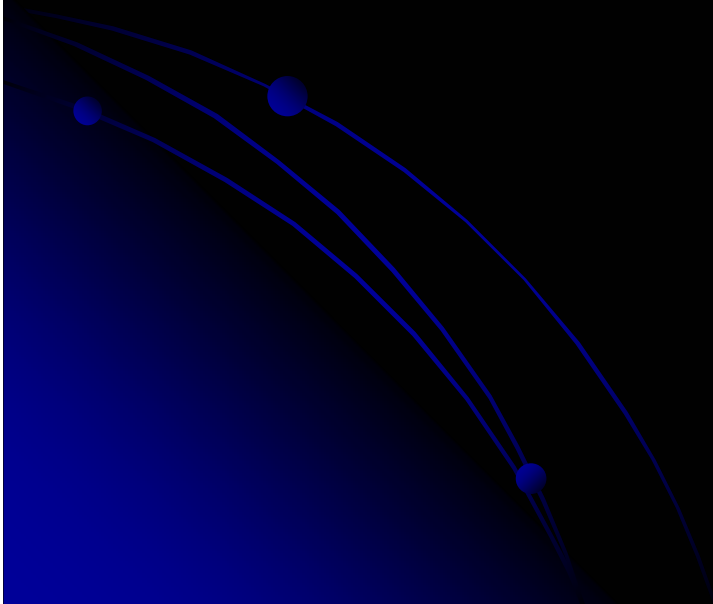
INRA, Lusignan, France

Marc Ghesquiere, Philippe Barre

# Acknowledgements

Ministry of Agriculture of the Czech Republic (grant award NAZV QH71267): The development and use of DArT array for xFestulolium breeding

Czech Science Foundation (grant award 521/07/P479): Cytogenetic mapping of genome of meadow fescue (*Festuca pratensis* Huds.)



Thank you for your attention

