

Emerging Role of Interleukin-1 in Cardiovascular Diseases

B. VICENOVA¹, V. VOPÁLENSKÝ¹, L. BURÝŠEK², M. POSPÍŠEK^{1,3}

¹Department of Genetics and Microbiology, Faculty of Science, Charles University, Prague, ²Gen-Trend, s.r.o., České Budějovice, ³Centre of Biomedical Informatics, Institute of Computer Science AS CR, Prague, Czech Republic

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Summary

There is an increasing evidence linking dysbalance between various proinflammatory mediators and higher risk of cardiovascular events and pathologies. Likewise, some of the cardiovascular diseases lately appeared to have an autoimmune component. Interleukin-1 (IL-1), a master regulator of diverse inflammatory processes in higher eukaryotes and the key player in numerous autoimmune disorders including rheumatoid arthritis, diabetes mellitus or systemic sclerosis, has recently been proved to be involved in development of several cardiovascular diseases as well. This report aims to give a summary on current knowledge about the IL-1 signaling pathways and about the implication of IL-1 and the IL-1 receptor antagonist (IL-1Ra) in some of the diseases of the cardiovascular system.

Key words

Interleukin-1 • Interleukin-1 receptor antagonist protein • Signal pathways • Cardiovascular diseases

Corresponding author

M. Pospíšek, Department of Genetics and Microbiology, Faculty of Science, Charles University, Prague, Viničná 5, 128 44 Prague 2, Czech Republic. Fax: +420 221951724. E-mail: martin@natur.cuni.cz

Introduction

Nowadays, inflammation is well established as a crucial component of diverse chronic diseases and interleukin-1 (IL-1), a proinflammatory cytokine with pleiotropic biological effects, appears to be one of the key

players of various inflammation-linked disorders. Currently, the role of interleukin-1 in rheumatoid arthritis, Alzheimer disease (Nicoll *et al.* 2000, Rainero *et al.* 2004) and associated vascular dementia (Yucesoy *et al.* 2006), diabetes mellitus, periodontitis (Lopez *et al.* 2005), systemic sclerosis (Kawaguchi 1994, Kawaguchi *et al.* 2006), autoimmune encephalomyelitis (Sutton *et al.* 2006) and cerebral infarction (Um *et al.* 2005) is well accepted.

Interleukin-1 was first described in 1972 as a lymphocyte-activating factor (Gery and Waksman 1972) and later was shown to exert a variety of effects including induction of inflammation, body temperature increase, stimulation of proliferation of T and B cells, induction of acute phase proteins and prostaglandins or regulation of hematopoiesis. Its activities are not restricted to the immune system; interleukin-1 is also involved in the regulation of blood calcium levels, stimulation of proliferation of various cells, regulation of blood pressure or modulation of sleep. However, IL-1 represents one of the most important mediators of the inflammatory response that induces a cascade of proinflammatory effector molecules. It is thus not surprising that reports of evidence are accumulating that interleukin-1 is involved in pathogenesis of certain cardiovascular diseases, mainly due to its proinflammatory potential. Multiple studies revealed that effects of IL-1 on cardiac myocytes include structural and functional alterations like hypertrophy (Thaik *et al.* 1995, Isoda *et al.* 2001), regulation of MAP kinase pathway and induction of expression of inducible nitric oxide synthase (iNOS) (Singh *et al.* 1996, Fichera *et al.* 2004), induction of the vasodilator peptide

adrenomedullin (Horio *et al.* 1998) or vascular endothelial growth factor (VEGF) induction (Tanaka *et al.* 2000). Certain IL-1 gene variations have been associated with overexpression of inflammatory mediators and also with increased risk of cardiovascular events (Francis *et al.* 1999, Kornman *et al.* 1999, Momiyama *et al.* 2001, Kornman *et al.* 2004, Iacoviello *et al.* 2005, Florez *et al.* 2006, Kornman 2006, Lai *et al.* 2006). Current knowledge about the involvement of interleukin-1 proteins in various cardiovascular diseases is reviewed throughout this text.

The biology of IL-1

Interleukin-1 is a term for two distinct but related proteins, interleukin-1 α and interleukin-1 β , encoded by two separate genes. Both of them are produced in consequence of stress or cell injury as 31-kDa precursors which undergo proteolytical cleavage by specific proteases during the process of maturation. The major interleukin-1 producing cells are macrophages, but many other cells like neutrophils, lymphocytes, dendritic cells, keratinocytes, endothelial cells, hepatocytes, fibroblasts or muscle cells have been shown to synthesize IL-1. Main target cells of IL-1 actions are primarily cells of the immune system such as monocytes, lymphocytes, granulocytes, dendritic cells, but this cytokine can affect many other cells like epithelial cells, fibroblasts, endothelial cells or smooth muscle cells.

The three-dimensional structure of IL-1 α and IL-1 β is composed of 12 antiparallel beta-strands arranged into a beta-trefoil. Both of the IL-1 proteins lack the signal peptide and if secreted, the secretion proceeds along a non-conventional secretory pathway differing from the classical endoplasmic reticulum-Golgi route (Rubartelli *et al.* 1990). IL-1 α and IL-1 β use the same membrane receptor (IL-1R) to trigger the signal transduction pathway leading to activation of NF- κ B or AP-1.

Despite all of the structural and functional similarities these two forms of IL-1 possess different features and play partly different roles in the organism. Unlike IL-1 β both the IL-1 α precursor molecule (amino acids 1-271) and the mature protein (amino acids 113-271) generated by calpain cleavage are biologically active and able to bind to membrane receptors (Mosley *et al.* 1987). IL-1 α is constitutively expressed by many cell types under physiological conditions and its synthesis is stimulated during inflammation. On the contrary, IL-1 β is

not produced unless the cell receives an inflammatory signal and also its cleavage by caspase-1 is a tightly regulated process. Moreover, while mature IL-1 β is released from cells *via* an ATP-dependent non-classical secretory pathway including P2X₇ receptors and pannexin-1 (Ferrari *et al.* 1997, Pelegrin and Surprenant 2006), the release of IL-1 α seems to employ a different pathway dependent on copper ions and protein product of the S100A13 gene (Mandinova *et al.* 2003). Nonetheless, the secretion of mature IL-1 α from human cells is a rare event and the protein is usually not detected in body fluids of healthy individuals. To exert its biological activity and interact with the IL-1 receptor, the IL-1 α precursor needs to be inserted into the plasma membrane due to myristoylation on specific lysine residues (Stevenson *et al.* 1993). It is now accepted that the major part of the IL-1 α precursor remains localized intracellularly. A nuclear localization sequence is present within the precursor molecule and this points toward a specific function of IL-1 α in the cell nucleus (Wessendorf *et al.* 1993, Maier *et al.* 1994). Conversely, mature IL-1 β is the secreted form of IL-1.

It seems that some of the biological functions are shared by both IL-1 forms and some of them are not. In particular, the striking difference in regulation, processing, tissue-specific expression, subcellular localization and compartmentalization of IL-1 α and IL-1 β isoforms suggests the differential function and physiological role of these molecules (Kavita and Mizel 1995, Beissert *et al.* 1998, Nakae *et al.* 2001, Hacham *et al.* 2002, Song *et al.* 2003). However, only few studies have evaluated the relationship of IL-1 α and IL-1 β in parallel with pathophysiology of various diseases. Therefore, the precise differences in the role of these two forms of IL-1 have not been clarified yet and many questions concerning the possible redundancy of IL-1 remain open.

IL-1-mediated inflammatory effects can be abolished by an effective natural inhibitor, IL-1 receptor antagonist (IL-1Ra). IL-1Ra joins both IL-1 proteins in the so-called IL-1 family of proteins cognate in the terms of structural and evolutionary relationship. There are both soluble and intracellular isoforms of IL-1Ra. The soluble IL-1 receptor antagonist molecule binds the IL-1 receptor, but this interaction does not lead to the activation of the signaling pathway. It is synthesized primarily in monocytes and macrophages, but might be produced in various other cell types. The function of the intracellular IL-1Ra (icIL-1Ra) isoforms remains unclear,

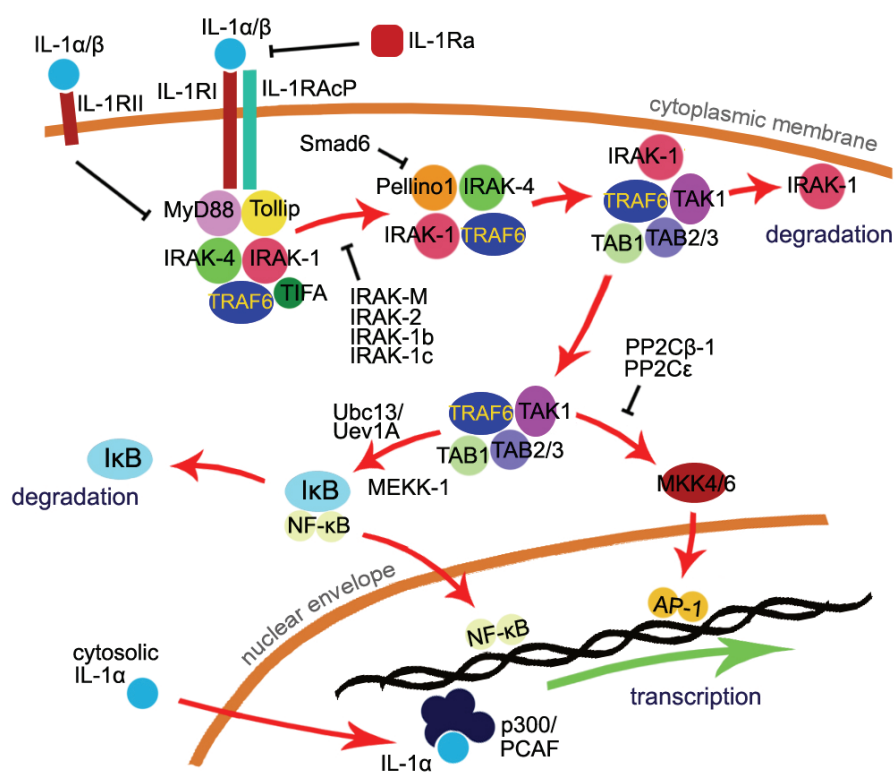


Fig. 1. The IL-1 signaling and its negative regulation.

but type I icIL-1Ra has been demonstrated to interact with the third component of the COP9 signalosome CSN3 and inhibit activity of CSN-associated kinases as well as IL-1 α -mediated release of IL-6 and IL-8, displaying thus unique anti-inflammatory activities inside the cells (Banda *et al.* 2005). Recently, Dewberry *et al.* (2008) reported that type I icIL-1Ra promotes endothelial cell proliferation by modulating CDK2 activity and retinoblastoma (Rb) protein phosphorylation.

Nowadays, the recombinant non-glycosylated form of IL-1Ra, anakinra, is used for treatment of rheumatoid arthritis and is tested also for IL-1 inhibition during acute gout (So *et al.* 2007), diabetes mellitus (Tellez *et al.* 2005) or familial cold autoinflammatory syndrome (Metyas and Hoffman 2006).

The IL-1 signaling pathways

Interleukin-1 triggers a complex network of signaling pathways which are influenced by a variety of regulatory proteins and target expression of a significant number of genes (Fig. 1).

Extracellular IL-1 is recognized and bound by the type I transmembrane IL-1 receptor which forms a complex with the IL-1 receptor accessory protein (IL-1RAcP) (Greenfeder *et al.* 1995), adaptor MyD88 (Muzio *et al.* 1997), Tollip (Burns *et al.* 2000) and kinases

IRAK-1 (Croston *et al.* 1995) and IRAK-4 (Li *et al.* 2002, Lye *et al.* 2004). Upon IL-1 binding to the receptor, IRAK-1 becomes phosphorylated on Thr-209 and Thr-387 (supposedly by IRAK-4) and autophosphorylated within the so-called ProST region (proline-, serine-, threonine-rich) (Croston *et al.* 1995, Yamin and Miller 1997, Kollwe *et al.* 2004), interacts with tumor necrosis factor receptor-associated factor (TRAF6) (Cao *et al.* 1996) presumably through TIFA protein (TRAF-interacting protein with a forkhead-associated domain) (Takatsuna *et al.* 2003, Ea *et al.* 2004) and leaves the receptor. Pellino1, a homologue of *Drosophila* adaptor protein Pellino, is recruited (Jiang *et al.* 2003) and thus a new intermediate complex IRAK-1-IRAK-4-TRAF6-Pellino1 is formed. Subsequently, IRAK-4 and Pellino1 leave the complex and TRAF6 undergoes polyubiquitinylation (Wang *et al.* 2001) that brings about its activation. At the cytoplasmic membrane, another complex is formed that consists of IRAK-1, TRAF6, transforming growth factor- β -activated kinase 1 (TAK1) and two TAK1-binding proteins TAB1 and TAB2 (Ninomiya-Tsuji *et al.* 1999, Takaesu *et al.* 2001) or alternatively TAK1, TAB1 and TAB3 (Cheung *et al.* 2004). Afterwards, phosphorylated IRAK-1 is ubiquitinylated and degraded in proteasomes, TAB2 becomes phosphorylated and TAB1 promotes autophosphorylation of TAK1 and thus its activation

(Kishimoto *et al.* 2000). Phosphorylation of TAK1 and TAB2 then facilitates the translocation of this complex from the membrane to the cytosol (Jiang *et al.* 2002).

As a result of all these events, NF- κ B inhibitor (I κ B) becomes phosphorylated on serine residues -32 and -36 by MEKK1 kinase (Lee *et al.* 1997), covalently modified by TRAF6-mediated and Ubc13/Uev1A-dependent ubiquitinylation at lysine-63 (Deng *et al.* 2000) and targeted to proteasome for degradation. NF- κ B transcription factor is thus made available and enters the nucleus where it activates transcription of target genes. Alternatively, the signal triggered by IL-1 is mediated by MAP kinases and AP-1 since the TAK1 kinase is able to activate both MKK4-JNK and MKK6-p38 pathway as well (Yamaguchi *et al.* 1995).

Among the proteins whose synthesis is induced by IL-1 there are proinflammatory cytokines (IL-6, TNF) and mediators (cyclooxygenase, inducible nitric oxide synthase), acute phase proteins (C-reactive protein, complement factors) or growth factors (FGF; reviewed in Dinarello 1996).

The IL-1 signaling pathway is very similar to that triggered by the Toll-like receptors (TLR) involved in the innate immunity. Also both IL-1R and TLR contain the evolutionary conserved intracellular TIR domain that is crucial for interactions of the receptors with adaptors mediating the signal from outside.

Aside from the signaling pathways triggered by IL-1 from the cell surface, there has been an evidence of IL-1 α activity in the cell nucleus. As mentioned above, IL-1 α can be found in the nucleus since the precursor (but not the mature protein) contains a nuclear localization sequence (Wessendorf *et al.* 1993). This sequence is situated within the so-called N-terminal peptide of IL-1 α (IL-1 α NTP) that is cleaved from the precursor by calpain in the process of IL-1 α maturation. However, IL-1 α NTP is evolutionarily well conserved among vertebrates, suggesting that it is more than a byproduct of IL-1 α cleavage. Both IL-1 α precursor protein and IL-1 α NTP are commonly found in the cell nucleus and we previously demonstrated that IL-1 α interacts with nuclear histone acetyltransferase complex p300/PCAF *via* its N-terminal part (Burýšková *et al.* 2004). Results confirming the role of IL-1 α precursor and NTP in transcriptional activation were observed by others as well (Werman *et al.* 2004). IL-1 α NTP was also shown to be associated with elements of the RNA splicing apparatus and to induce apoptosis in malignant cells (Pollock *et al.* 2003). The translocation of IL-1 α to the nucleus is mediated by HAX-1 protein which

appears to interact with IL-1 α by three different segments (Kawaguchi *et al.* 2006). Moreover, the IL-1 α precursor, but not the mature protein, can regulate the human endothelial cell migration *in vitro* (McMahon *et al.* 1997). All these results suggest that the IL-1 α precursor or its N-terminal peptide might have a specific function in the cell nucleus. However, this “intracrine” activity of IL-1 α is still poorly understood.

Interestingly, a small group of „dual-function“ proteins that are active both extracellularly and within the cell nucleus has been described recently (Carriere *et al.* 2007). From this point of view, interleukin-1 α resembles to the high-mobility group box 1 (HMGB1) protein that has been found associated with nuclear chromatin (Agresti and Bianchi 2003) but acts also as a proinflammatory cytokine (Wang *et al.* 1999a,b), and interleukin-33, a cytokine from the IL-1 family promoting the Th2 response but acting also as a nuclear transcriptional repressor (Carriere *et al.* 2007, Gadina and Jefferies 2007). The latter molecule has been recently found to be involved in cardiovascular disease as well (Kakkar and Lee 2008).

Negative regulation of the IL-1 pathway can occur at multiple levels. Outside the cytoplasmic membrane, the functional concentration of IL-1 can be decreased by action of an inactive IL-1 receptor type II which lacks the TIR domain and does act as a kind of decoy target for IL-1, preventing its binding to IL-1R type I. Furthermore, IL-1Ra that is unable to trigger the IL-1R-mediated pathway can competitively occupy the IL-1 receptors and therefore is able to abrogate the signaling. Inside the cell, two naturally occurring variants of IRAK but lacking the kinase activity, IRAK-M and IRAK-2, can also inhibit the IL-1/TLR signaling (Kobayashi *et al.* 2002, Janssens and Beyaert 2003). Regulation of the IRAK kinase activity is further provided by the mechanism of alternative splicing. IRAK-1b, an alternatively spliced version of IRAK-1, is not efficient in NF- κ B activation and does not become phosphorylated and degraded in response to IL-1. This prolonged stability of IRAK-1b may lead to the replacement of IRAK-1 by IRAK-1b in the signaling complex and attenuate the IL-1 signaling pathway (Jensen and Whitehead 2001). Another splice variant of IRAK-1 lacking exon 11, IRAK-1c, associates with MyD88, Tollip and TRAF6 but acts as a negative regulator by its inability to be phosphorylated by IRAK-4 and subsequently autophosphorylated (Rao *et al.* 2005). Further suggested negative regulators of the IL-1

signaling pathway are Smad6, PP2C β -1 and PP2C ϵ . Binding of Smad6 to Pellino1 prevents its involvement in the signaling (Choi *et al.* 2006). TAK1-mediated signaling can be inhibited *via* TAK1 dephosphorylation by PP2C β -1 isoform of protein phosphatase PP2C (Hanada *et al.* 2001). Another member of the PP2C family, PP2C ϵ , might be involved in the attenuation of TAK1 function as well (Li *et al.* 2003).

IL-1 and atherosclerosis

Coronary arterial disease is a chronic disease that results from the formation of an atheromatous plaque in the artery (so called atherosclerosis - "hardening of the artery") and subsequent reduction of arterial lumen diameter. Death can occur through myocardial infarction, cerebrovascular events and renal failure. The classical risk factors for atherosclerosis include elevated LDL cholesterol level, smoking, obesity, diabetes or sedentary lifestyle. However, in the course of time it has become evident that atherosclerosis may also have autoimmune components and that many cytokines are likely to participate in the pathogenesis of this disorder especially in people lacking classical risk factors. Current data show clearly that atherosclerosis is associated with activation of the inflammatory processes and with systemic increase of proinflammatory molecules such as IL-1, IL-6, TNF or C-reactive protein and thus it turns from a disease caused by a simple accumulation of lipids into a complex disorder influenced by the inflammatory response of the arterial wall (reviewed in (Libby 2002)). The proatherogenic effect of IL-1 is attributed to its ability to modulate a number of key events involved in the complex inflammatory process of atherogenesis such as the vessel wall inflammation, leukocyte chemotaxis and adhesion or plaque rupture (Bochner *et al.* 1991, Libby *et al.* 1995, Von Der Thusen *et al.* 2003, Waehre *et al.* 2004).

The involvement of interleukin-1 during atherogenesis could be exemplified by its ability to promote atheromatous plaque instability due to upregulation of matrix metalloproteinases at the site of plaque formation. Increased levels of proinflammatory cytokines IL-1 and TNF as well as the chemokine IL-8 are found within the atheromatous plaque (Tipping and Hancock 1993, Apostolopoulos *et al.* 1996) together with matrix metalloproteinases that are synthesized by smooth muscle cells, endothelial cells and macrophages (Galis *et al.* 1995, Rajavashisth *et al.* 1999b). These zinc-dependent endopeptidases have the capacity to degrade

various components of extracellular matrix (reviewed in (Newby 2005)) and indeed, they are capable to induce collagen breakdown in atheromatous plaques (Shah *et al.* 1995). The matrix metalloproteinases have been shown to be upregulated by IL-1 and other proinflammatory cytokines in smooth vascular cells, endothelial cells and macrophages (Hanemaaijer *et al.* 1993, Galis *et al.* 1994, Rajavashisth *et al.* 1999a,b, Herman *et al.* 2001) and inversely, their induction in smooth muscle cells can be inhibited by IL-1Ra (Lee *et al.* 1995). Increased expression of matrix metalloproteinases induced by proinflammatory cytokines such as IL-1 might therefore promote destabilization of atheromatous plaques and may lead to thrombosis (Galis *et al.* 1995).

Further proatherogenic potential of IL-1 is attributed to the ability of this cytokine to modulate cell adhesion and subsequent monocyte infiltration into the subendothelial space. According to Kirii *et al.* (2003), the absence of IL-1 β decreases the severity of atherosclerosis in apoE deficient mice, possibly through decreased expressions of vascular cell adhesion molecule-1 (VCAM-1) and monocyte chemoattractant protein-1 (MCP-1) in the aorta. These adhesion-promoting molecules are likely to attract monocytes to the arterial intima. Following adhesion, monocytes enter the intima, differentiate into macrophages and then change into foam cells, that is one of the key steps in atherogenesis.

Moreover, IL-1 has been shown to influence the metabolism of plasma levels of cholesterol and serum amyloid A (SAA), both of which are frequently associated with atherogenesis. Following the atherogenic diet, mice lacking IL-1 α showed a reduced SAA level and a lower atherosclerotic lesion area when compared to wild type mice, despite a higher total cholesterol level (Kamari *et al.* 2007). Interestingly, the anti-atherogenic effect of IL-1 α knock-out was stronger than the result of IL-1 β deficiency, whose atheroprotective effect was described previously (Kirii *et al.* 2003). The early lesion formation was enhanced specifically by IL-1 α having origin in bone marrow cells, most probably macrophages. On the contrary, this bone marrow-derived IL-1 α had no effect on cholesterol levels (Kamari *et al.* 2007). The involvement of IL-1 in cholesterol metabolism is supported by results showing elevated levels of plasma cholesterol in mice lacking the IL-1 receptor antagonist (Isoda and Ohsuzu 2006).

Excessive angiogenesis in consequence of VEGF induction by IL-1 may be important in atherosclerosis as well. IL-1 α stimulates secretion of

VEGF in a dose-dependent manner and promotes angiogenesis in mice injected subcutaneously with IL-1 α (Salven *et al.* 2002). Similarly, VEGF can be induced by IL-1 β as well (Voronov *et al.* 2003) presumably due to the upregulation of hypoxia inducible factor-1 α (HIF-1 α), a major transcription factor for VEGF (Thornton *et al.* 2000), but IL-1 β -mediated proangiogenic activity independent on VEGF induction was observed in monocytes treated with osteopontin too (Naldini *et al.* 2006). Since osteopontin is abundantly expressed in the atheromatous lesions (Giachelli *et al.* 1993), its interaction with the proinflammatory cytokine IL-1 β could possibly contribute to the growth of atherosclerotic lesions and destabilization of the atheromatous plaques.

Up to the present, there have been reported numerous links between angiogenesis and inflammation that consist in vascular permeability increase, activation of the endothelium, cell migration and differentiation and of course influence of a range of cytokines and regulatory factors. Joint action of angiogenesis and inflammation has been reported in a number of diseases including osteoarthritis (reviewed in (Bonnet and Walsh 2005), rheumatoid arthritis (Paleolog 2002) or lung diseases (Walsh and Pearson 2001); however, the direct link between formation of new vessels and development or clinical outcome of atherosclerosis has not been established until now (Khurana *et al.* 2005).

According to recent genetic studies, certain polymorphisms in IL-1 genes are known to influence the plasma levels of C-reactive protein (Berger *et al.* 2002, Latkovskis *et al.* 2004). Elevated levels of C-reactive protein, a classical acute phase protein, indicate an increased risk of atherothrombotic and cardiovascular events (Danesh *et al.* 2000, 2004). Studies in Italian, Chinese and Korean populations confirmed association of some of the interleukin-1 gene cluster polymorphisms with ischemic stroke (Lee *et al.* 2004, Iacoviello *et al.* 2005, Lai *et al.* 2006).

Other recent works have shown that certain bacterial species such as *Chlamydia pneumoniae*, *Helicobacter pylori* and *Porphyromonas gingivalis* aggravate atherosclerotic lesion development in animal models and that infectious agents including those that cause periodontal disease may also play an important role (Epstein *et al.* 1999, Chi *et al.* 2004, Hoge and Amar 2007). Bacterial infection triggers release of IL-1 by two different mechanisms: antigenic bacterial products such as lipopolysaccharide (LPS) stimulate IL-1 release from cells *via* Toll-like receptors and alternatively, recognition

of bacteria *via* pattern recognition receptors such as CD-14 enhances IL-1 release from macrophages. Certain IL-1 gene polymorphisms can influence the process of coronary artery disease in patients with *Chlamydia pneumoniae* infection and can lead to an increased risk of myocardial infarction (Momiyama *et al.* 2001). An epidemiological association between periodontitis and cardiovascular diseases has been reported in multiple studies as well (Kornman *et al.* 1999, Kornman and Duff 2001).

The key importance of IL-1 as a proatherogenic factor is further supported by other studies that point out the role of IL-1 receptor antagonist in the process of atherogenesis. A study by Merhi-Soussi *et al.* (2005) demonstrates that the IL-1/IL-1Ra ratio plays a critical role in the pathogenesis of vascular inflammation and atherosclerosis. Indeed, mice homozygous for the null mutation of IL-1Ra develop lethal arterial inflammation (Nicklin *et al.* 2000). It has been also shown that frequency of allele 2 of the IL-1RN gene coding for IL-1Ra is increased in patients with single vessel coronary artery disease and IL-1RN allele distribution is different in single and multiple coronary arterial diseases (Francis *et al.* 1999). Furthermore, the IL-1 receptor antagonist was demonstrated to inhibit neointima formation after coronary artery injury (Isoda *et al.* 2003, Morton *et al.* 2005) and to be involved in metabolism of cholesterol (Isoda and Ohsuzu 2006). IL-1Ra^{-/-} knock-out mice are unable to synthesize mRNA of cholesterol 7 α -hydroxylase, an enzyme that is involved in cholesterol conversion into bile acid, and display increased plasma cholesterol levels compared to mice with normal IL-1Ra expression (Isoda and Ohsuzu 2006). Thus, deficiency in the IL-1 receptor antagonist obviously has a proatherogenic potential and these observations point again toward the significance of IL-1 as an important agent in atherogenesis.

When it comes to obesity that is a classical risk factor for atherosclerosis, it has been shown that besides many other activities affecting nearly all organ systems, the perivascular white adipose tissue (pWAT) acts as an important source of secreted cytokines, chemokines and various metabolically active molecules such as TNF- α , TGF- β , leptin, adiponectin, MCP-1, IL-8, IL-6, IL-1 or IL-1Ra. Since the distance of pWAT from the vascular wall has been estimated to be less than 100 μ m, the factors produced by perivascular adipocytes could interact with the blood vessels and contribute to progression of atherosclerosis by facilitating infiltration

of leukocytes and monocytes, promoting angiogenesis and enhancing smooth muscle cell proliferation (Henrichot *et al.* 2005, Thalmann and Meier 2007).

Interestingly, serum levels of IL-1Ra are over 7-fold elevated in obese patients (Meier *et al.* 2002) and adipose tissue acts as an important source of IL-1Ra (Juge-Aubry *et al.* 2003). Although it seems that the overexpressed IL-1Ra may act against inflammation and should therefore represent a benefit, it seems that due to its metabolic activities, the increased production of the antagonistic protein is likely to promote resistance to leptin, weight gain and insulin resistance (Meier *et al.* 2002, Somm *et al.* 2005).

IL-1 and myocardial infarction

Apart from the initiation and progression of atherosclerosis, inflammation plays a role also in activation of the coagulation process that contributes to the development of myocardial infarction. This cardiac event usually occurs when a blood clot formed in the artery after the atheromatous plaque rupture blocks the heart artery and the heart muscle starves of oxygen. Patients with myocardial infarction show significantly elevated levels of proinflammatory cytokines including IL-1, IL-6 or TNF (Miyao *et al.* 1993, Guillen *et al.* 1995) and also of the IL-1 receptor antagonist (Patti *et al.* 2004).

It has been shown that interleukin-1 β enhances expression of tissue factor and induces procoagulant activity (Schwager and Jungi 1994). According to Iacoviello *et al.* (2005), certain polymorphisms in the IL-1 β gene can be associated with incidence of myocardial infarction at young age, probably due to the inflammation-activated blood coagulation. However, Momiyama *et al.* (2005) states more precisely the IL-1 β gene polymorphism to be associated with myocardial infarction only in patients with *Chlamydia pneumoniae* seropositivity.

Prolonged ischemia or oxygen shortage in consequence of an insufficient blood supply may result in damage and necrosis of myocardial cells. Such an event is likely to be followed by sterile inflammation triggered by a variety of molecules released from dying cells. A recent report by Chen *et al.* (2007) shows that the IL-1R-MyD88 pathway is involved in the sterile inflammatory response that could contribute to the development of many diseases including cardiovascular disorders. This inflammatory response is mediated by neutrophils

recruited to the site of injury and requires an intact IL-1R signaling on none-bone-marrow-derived cells. In this pathway, IL-1 α appears to be the key cytokine mediator. In IL-1R $^{-/-}$ and MyD88 $^{-/-}$ knock-out mice, the sterile inflammatory response mediated by neutrophils is markedly reduced while monocyte recruitment to the site of injury remains relatively intact. Therefore, blocking the IL-1 α action in order to reduce the extent of tissue damage might not impair the host defense responses mediated by monocytes (Chen *et al.* 2007).

During the postinfarct period, many functional and morphological changes can occur within the heart. Adverse remodeling with left-ventricular dilatation represents a postinfarct complication that involves a number of functional changes and may lead to heart failure. One of the therapeutical approaches to the treatment of the infarcted heart that was used successfully in experimental animals is the transplantation of skeletal myoblasts. However, the survival of the grafted cells might be reduced by the acute inflammatory response mediated by IL-1 β (Suzuki *et al.* 2004). Recently, Murtuza *et al.* (2004) showed that transplantation of skeletal myoblast secreting IL-1Ra results in the improvement of systolic function, attenuation of adverse remodeling and reduction in myocyte hypertrophy. Moreover, in mice lacking the IL-1 receptor type I, adverse remodeling of the infarcted heart is markedly decreased due to a reduced collagen deposition in the absence of IL-1 signaling (Bujak *et al.* 2008). Targeting IL-1 as a molecule modulating the pathological heart remodeling could therefore have an important clinical relevance (Murtuza *et al.* 2004).

Though, interleukin-1 might be involved also in cardiac repair mechanism. In human cardiac cells, IL-1 α along with other inflammatory mediators upregulates the production of plasminogen activator inhibitor 1 (PAI-1) (Macfelda *et al.* 2002). Previous studies showed that treatment with PAI-1 prevents the cardiac rupture after myocardial infarction in mice (Heymans *et al.* 1999). Thus, IL-1 α might contribute to the wound healing and cardiac repair after the stroke.

IL-1 and myocarditis

Myocarditis is a disease that may have an autoimmune background and is accompanied by the infiltration of inflammatory cells in the myocardium and subsequent inflammation of the heart muscle. In most cases it is caused by infection with various viruses

(enterovirus, coxsackie virus, cytomegalovirus, poliovirus, influenza virus and others) and apparently, IL-1 is involved in this process (Lane *et al.* 1992, Shioi *et al.* 1996). As a model of myocarditis, mice infected with coxsackie virus B3 (CB3) have been established. The major autoantigen in this model is the cardiac myosine (Fairweather *et al.* 2001).

Several studies reported increased levels of IL-1 α and IL-1 β during coxsackie virus-induced myocarditis (Matsumori *et al.* 1994, 1999, Okuno *et al.* 2000). IL-1 treatment enhances the CB3-induced myocarditis in genetically resistant mice (Lane *et al.* 1992) while gene therapy based on plasmids expressing IL-1Ra is effective for treatment of viral myocarditis in tested mice (Nakano *et al.* 2001, Lim *et al.* 2002). Mikami *et al.* (1996) showed that IL-1 together with other proinflammatory mediators such as TNF- α originated from inflammatory cells infiltrating the myocardial lesions and as a consequence of that, inducible NO synthase mRNA and protein were induced specifically after coxsackievirus B3 virus infection in mice. IL-1-mediated activation of iNOS causes excessive production of nitric oxide and subsequent cytotoxicity and negative inotropic effect on myocardial tissue (Finkel *et al.* 1992, Pinsky *et al.* 1995, Nakano *et al.* 2001). Indeed, treatment with low-dose iNOS inhibitor results in decreased myocardial injury in mice (Mikami *et al.* 1997).

It has been also shown that interleukin-1 β can bind to the IL-1 receptors on the surface of dendritic cells and stimulate them to produce cytokines including IL-12, required for activation of autoreactive CD4⁺ cells that are responsible for the development of autoimmune myocarditis. In IL-1R^{-/-} knock-out mice immunized with a self-peptide, activation of autoreactive CD4⁺ cells was impaired. These data confirmed the key role of interleukin-1 in pathogenesis of autoimmune myocarditis (Eriksson *et al.* 2003). Fairweather *et al.* (2003) investigated the role of IL-12 on the development of CB3-induced myocarditis. In their experiments they showed that serum levels of IL-1 β and severity of inflammation are significantly reduced in coxsackie virus-infected IL-12^{-/-} knock-out mice which directly leads to the conclusion that IL-12 modulates the production of IL-1 β . The development of chronic myocarditis that follows the initial phase of acute myocarditis can be suppressed by IL-12-induced IFN- γ in the murine myocarditis model. IFN- γ contributes to reducing the expression of IL-1 β in the heart and

decreasing the overall severity of the disease (Fairweather *et al.* 2004). During chronic myocarditis, the incidence of cardiac fibrosis and pericarditis is increased in IFN- γ ^{-/-} knock-out mice and the level of IL-1 α and IL-1 β in hearts of these mice is elevated (Fairweather *et al.* 2004, Grun *et al.* 2005).

Treatment with digoxin increases the intracardiac production of IL-1 and decreases the survival of mice infected with encephalomyocarditis virus (Matsumori *et al.* 1999). The latter result should be taken into consideration since this drug has been used for treatment of viral myocarditis.

IL-1 and dilated cardiomyopathy

Dilated cardiomyopathy is a non-curable disease of the heart muscle during which the heart becomes enlarged and is unable to pump the blood strongly and efficiently. This usually leads to heart failure. In the majority of cases the cause of dilated cardiomyopathy is unknown, however the factors that contribute to the development of this disease are viral infection (coxsackie B viruses), pregnancy, excessive alcohol intake, genetic disorders or autoimmunity.

Patients with dilated cardiomyopathy show elevated levels of IL-1 β (Francis *et al.* 1998, Vanderheyden *et al.* 2005) that is produced by endothelial cells and by myocytes themselves (Francis *et al.* 1998). On the contrary, the IL-1Ra gene expression is significantly lower in patients with dilated cardiomyopathy as compared to controls (Westphal *et al.* 2008). Therefore, this imbalance between the proinflammatory mediator IL-1 β and its antagonistic molecule IL-1Ra could underlie the pathogenesis and progression of dilated cardiomyopathy. Besides the potential enhancement of the inflammation in the heart tissue, the action of IL-1 and IL-1-induced cytokines on heart muscle cells can induce cardiodepressant effects (Gulick *et al.* 1989, Cain *et al.* 1999, Matsumori *et al.* 1999) that can be attenuated by beta-adrenergic blockers (Prabhu *et al.* 2000, Ichihara *et al.* 2006).

Conclusions

Both IL-1 α and IL-1 β are cytokines with pleiotropic effects in vertebrate organism and the cytokine network, that they make part of, regulates many essential processes. Current data support the notion that both IL-1 isoforms together with their negative regulator IL-1Ra play an essential role in development of various

Table 1. Mechanisms of involvement of the IL-1 cytokines in cardiovascular diseases.

Biological effect	Molecule involved	Corresponding mechanism	Disease/disorder	Citation
<i>proadhesive activity</i>	IL-1 β	stimulation of adhesion-promoting molecules (VCAM-1)	atherosclerosis	Kirii <i>et al.</i> 2003
<i>atheromatous plaque destabilization</i>	IL-1 α , IL-1 β	upregulation of matrix metalloproteinases	atherosclerosis	Rajavashisth <i>et al.</i> 1999a,b, Galis <i>et al.</i> 1995, Shah <i>et al.</i> 1995
<i>modulation of cholesterol plasma level</i>	IL-1 α , IL-1 β	SAA induction	atherosclerosis	Kamari <i>et al.</i> 2007, Merhi-Soussi <i>et al.</i> 2005
<i>stimulation of angiogenesis</i>	IL-1 α , IL-1 β	VEGF induction	atherosclerosis	Salven <i>et al.</i> 2002, Voronov <i>et al.</i> 2003
<i>vessel wall inflammation</i>	IL-1 α , IL-1 β	induction of inflammatory pathways through IL-1R I	atherosclerosis	Nicklin <i>et al.</i> 2000
<i>induction of procoagulant activity</i>	IL-1 β	stimulation of tissue factor expression	myocardial infarction	Schwager and Jungi 1994
<i>tissue damage, disease aggravation</i>	IL-1 α	mediation of sterile inflammation	myocardial infarction	Chen <i>et al.</i> 2007
<i>enhancement of adverse remodeling</i>	IL-1 β	modulation of collagen deposition	myocardial infarction	Bujak <i>et al.</i> 2000, Murtuza <i>et al.</i> 2004
<i>myocardial dysfunction, disease aggravation</i>	IL-1 α , IL-1 β	iNOS induction	myocarditis	Lim <i>et al.</i> 2002, Mikami <i>et al.</i> 1996, Nakano <i>et al.</i> 2001
<i>development of the disease</i>	IL-1 β	IL-12-mediated activation of autoreactive CD4 ⁺ cells	autoimmune myocarditis	Eriksson <i>et al.</i> 2003

cardiovascular diseases including atherosclerosis, myocardial infarction, myocarditis, dilated cardiomyopathy, infective endocarditis or chagasic cardiomyopathy. IL-1 gene variations that are associated with overexpression of inflammatory mediators correlate also with increased risk of cardiovascular events.

Actions of the proteins belonging to the IL-1 family on circulatory system are complex and imbalance between the agonists and IL-1Ra might have an important impact on the cardiovascular function. So far, it was demonstrated that these molecules may be involved in regulation of inflammation, angiogenesis, coagulation processes or cholesterol metabolism, they mediate

chemotaxis and facilitate adhesion (Table 1). Furthermore, IL-1 along with other proinflammatory cytokines as well as proteins induced by these are present in the myocardium of infants with congenital cardiac defects such as tetralogy of Fallot or ventricular septal defects (Qing *et al.* 2003). Taking into account the current wide application of cytokine antagonists like IL-1Ra/anakinra in treatment of serious autoimmune disorders, these therapeutic antagonists should be studied also from the point of view of the influence of dysbalanced cytokine levels on the cardiovascular system. An example of a successful application of IL-1Ra production in local tissues that yielded promising results

might be the transplantation of myoblasts secreting IL-1Ra that had led to improvement of the postinfarct myocardial function (Murtuza *et al.* 2004), another might be the IL-1Ra gene therapy treatment of viral myocarditis (Nakano *et al.* 2001, Lim *et al.* 2002).

Taken together, there is a considerable evidence of implication of IL-1 α , IL-1 β and IL-1Ra in the cardiovascular system and its various pathologies. However, future research on this field will be needed in order to elucidate the complex action of IL-1 on cardiovascular system in its whole extent.

Conflict of Interest

There is no conflict of interest.

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Abbreviations

IL, interleukin; IL-1R, interleukin-1 receptor; IL-1Ra, interleukin-1 receptor antagonist; NF- κ B, nuclear factor-kappa B; AP-1, activator protein-1; MAP, mitogen-activated protein; iNOS - inducible nitric oxide synthase; VEGF, vascular endothelial growth factor; IL-1RAcP,

interleukin-1 receptor accessory protein; CSN, COP9 signalosome; CDK2, cyclin-dependent kinase 2; Rb, retinoblastoma gene product; MyD88, myeloid differentiation primary response gene (88); Tollip, Toll-interacting protein; IRAK, interleukin-1 receptor-associated kinase; TRAF, tumor necrosis factor receptor-associated factor; TIFA, TRAF-interacting protein with a forkhead-associated domain; TAK, transforming growth factor-beta-activated kinase; TAB, transforming growth factor-beta-activated kinase-1-binding protein; I κ B, inhibitor of kappa B; MEKK, mitogen-activated protein kinase kinase kinase; MKK, mitogen-activated protein kinase kinase; JNK, c-Jun N-terminal kinase; TNF, tumor necrosis factor; FGF, fibroblast growth factor; TLR, Toll-like receptor; TIR, Toll/interleukin-1 receptor; IL-1 α NTP, N-terminal peptide of interleukin-1alpha; PCAF, p300/CREB-binding protein-associated factor; HAX-1, HS-1 associated protein X-1; PP2C β -1, protein phosphatase 2C beta-1; LDL, low-density lipoprotein; VCAM-1, vascular cell adhesion molecule-1; MCP-1, monocyte chemoattractant protein-1; SAA, serum amyloid A; HIF-1 α , hypoxia inducible factor-1alpha; LPS, lipopolysaccharide; pWAT, perivascular white adipose tissue; TGF- β , transforming growth factor beta; PAI-1, plasminogen activator inhibitor 1; IFN- γ , interferon gamma; CB3, coxsackie virus B3.

References

- AGRESTI A, BIANCHI ME: HMGB proteins and gene expression. *Curr Opin Genet Dev* **13**: 170-178, 2003.
- APOSTOLOPOULOS J, DAVENPORT P, TIPPING PG: Interleukin-8 production by macrophages from atheromatous plaques. *Arterioscler Thromb Vasc Biol* **16**: 1007-1012, 1996.
- BANDA NK, GUTHRIDGE C, SHEPPARD D, CAIRNS KS, MUGGLI M, BECH-OTSCHIR D, DUBIEL W, AREND WP: Intracellular IL-1 receptor antagonist type 1 inhibits IL-1-induced cytokine production in keratinocytes through binding to the third component of the COP9 signalosome. *J Immunol* **174**: 3608-3616, 2005.
- BEISSERT S, HOSOI J, STRATIGOS A, BRISSETTE J, GRABBE S, SCHWARZ T, GRANSTEIN RD: Differential regulation of epidermal cell tumor-antigen presentation by IL-1alpha and IL-1beta. *J Invest Dermatol* **111**: 609-615, 1998.
- BERGER P, McCONNELL JP, NUNN M, KORNMAN KS, SORRELL J, STEPHENSON K, DUFF GW: C-reactive protein levels are influenced by common IL-1 gene variations. *Cytokine* **17**: 171-174, 2002.
- BOCHNER BS, LUSCINSKAS FW, GIMBRONE MA JR, NEWMAN W, STERBINSKY SA, DERSE-ANTHONY CP, KLUNK D, SCHLEIMER RP: Adhesion of human basophils, eosinophils, and neutrophils to interleukin 1-activated human vascular endothelial cells: contributions of endothelial cell adhesion molecules. *J Exp Med* **173**: 1553-1557, 1991.
- BONNET CS, WALSH DA: Osteoarthritis, angiogenesis and inflammation. *Rheumatology (Oxford)* **44**: 7-16, 2005.
- BUJAK M, DOBACZEWSKI M, CHATILA K, MENDOZA LH, LI N, REDDY A, FRANGOGIANNIS NG: Interleukin-1 receptor type I signaling critically regulates infarct healing and cardiac remodeling. *Am J Pathol* **173**: 57-67, 2008.

- BURNS K, CLATWORTHY J, MARTIN L, MARTINON F, PLUMPTON C, MASCHERA B, LEWIS A, RAY K, TSCHOPP J, VOLPE F: Tollip, a new component of the IL-1RI pathway, links IRAK to the IL-1 receptor. *Nat Cell Biol* **2**: 346-351, 2000.
- BURÝŠKOVÁ M, POSPÍŠEK M, GROTHEY A, SIMMET T, BURÝŠEK L: Intracellular interleukin-1 α functionally interacts with histone acetyltransferase complexes. *J Biol Chem* **279**: 4017-4026, 2004.
- CAIN BS, MELDRUM DR, DINARELLO CA, MENG X, JOO KS, BANERJEE A, HARKEN AH: Tumor necrosis factor- α and interleukin-1 β synergistically depress human myocardial function. *Crit Care Med* **27**: 1309-1318, 1999.
- CAO Z, XIONG J, TAKEUCHI M, KURAMA T, GOEDDEL DV: TRAF6 is a signal transducer for interleukin-1. *Nature* **383**: 443-446, 1996.
- CARRIERE V, ROUSSEL L, ORTEGA N, LACORRE DA, AMERICH L, AGUILAR L, BOUCHE G, GIRARD JP: IL-33, the IL-1-like cytokine ligand for ST2 receptor, is a chromatin-associated nuclear factor in vivo. *Proc Natl Acad Sci USA* **104**: 282-287, 2007.
- CHEN CJ, KONO H, GOLENBOCK D, REED G, AKIRA S, ROCK KL: Identification of a key pathway required for the sterile inflammatory response triggered by dying cells. *Nat Med* **13**: 851-856, 2007.
- CHEUNG PC, NEBREDA AR, COHEN P: TAB3, a new binding partner of the protein kinase TAK1. *Biochem J* **378**: 27-34, 2004.
- CHI H, MESSAS E, LEVINE RA, GRAVES DT, AMAR S: Interleukin-1 receptor signaling mediates atherosclerosis associated with bacterial exposure and/or a high-fat diet in a murine apolipoprotein E heterozygote model: pharmacotherapeutic implications. *Circulation* **110**: 1678-1685, 2004.
- CHOI KC, LEE YS, LIM S, CHOI HK, LEE CH, LEE EK, HONG S, KIM IH, KIM SJ, PARK SH: Smad6 negatively regulates interleukin 1-receptor-Toll-like receptor signaling through direct interaction with the adaptor Pellino1. *Nat Immunol* **7**: 1057-1065, 2006.
- CROSTON GE, CAO Z, GOEDDEL DV: NF- κ B activation by interleukin-1 (IL-1) requires an IL-1 receptor-associated protein kinase activity. *J Biol Chem* **270**: 16514-16517, 1995.
- DANESH J, WHEELER JG, HIRSCHFIELD GM, EDA S, EIRIKSDOTTIR G, RUMLEY A, LOWE GD, PEPYS MB, GUDNASON V: C-reactive protein and other circulating markers of inflammation in the prediction of coronary heart disease. *N Engl J Med* **350**: 1387-1397, 2004.
- DANESH J, WHINCUP P, WALKER M, LENNON L, THOMSON A, APPLEBY P, GALLIMORE JR, PEPYS MB: Low grade inflammation and coronary heart disease: prospective study and updated meta-analyses. *BMJ* **321**: 199-204, 2000.
- DENG L, WANG C, SPENCER E, YANG L, BRAUN A, YOU J, SLAUGHTER C, PICKART C, CHEN ZJ: Activation of the I κ B kinase complex by TRAF6 requires a dimeric ubiquitin-conjugating enzyme complex and a unique polyubiquitin chain. *Cell* **103**: 351-361, 2000.
- DEWBERRY RM, KING AR, CROSSMAN DC, FRANCIS SE: Interleukin-1 receptor antagonist (IL-1ra) modulates endothelial cell proliferation. *FEBS Lett* **582**: 886-890, 2008.
- DINARELLO CA: Biologic basis for interleukin-1 in disease. *Blood* **87**: 2095-2147, 1996.
- EA CK, SUN L, INOUE J, CHEN ZJ: TIFA activates I κ B kinase (IKK) by promoting oligomerization and ubiquitination of TRAF6. *Proc Natl Acad Sci USA* **101**: 15318-15323, 2004.
- EPSTEIN SE, ZHOU YF, ZHU J: Infection and atherosclerosis: emerging mechanistic paradigms. *Circulation* **100**: e20-28, 1999.
- ERIKSSON U, KURRER MO, SONDEREGGER I, IEZZI G, TAFURI A, HUNZIKER L, SUZUKI S, BACHMAIER K, BINGISSER RM, PENNINGER JM, KOPF M: Activation of dendritic cells through the interleukin 1 receptor 1 is critical for the induction of autoimmune myocarditis. *J Exp Med* **197**: 323-331, 2003.
- FAIRWEATHER D, FRISANCHO-KISS S, YUSUNG SA, BARRETT MA, DAVIS SE, GATEWOOD SJ, NJOKU DB, ROSE NR: Interferon- γ protects against chronic viral myocarditis by reducing mast cell degranulation, fibrosis, and the profibrotic cytokines transforming growth factor- β 1, interleukin-1 β , and interleukin-4 in the heart. *Am J Pathol* **165**: 1883-1894, 2004.
- FAIRWEATHER D, KAYA Z, SHELLAM GR, LAWSON CM, ROSE NR: From infection to autoimmunity. *J Autoimmun* **16**: 175-186, 2001.

- FAIRWEATHER D, YUSUNG S, FRISANCHO S, BARRETT M, GATEWOOD S, STEELE R, ROSE NR: IL-12 receptor beta 1 and Toll-like receptor 4 increase IL-1 beta- and IL-18-associated myocarditis and coxsackievirus replication. *J Immunol* **170**: 4731-4737, 2003.
- FERRARI D, CHIOZZI P, FALZONI S, HANAU S, DI VIRGILIO F: Purinergic modulation of interleukin-1 beta release from microglial cells stimulated with bacterial endotoxin. *J Exp Med* **185**: 579-582, 1997.
- FICHERA LE, ALBAREDA MC, LAUCELLA SA, POSTAN M: Intracellular growth of *Trypanosoma cruzi* in cardiac myocytes is inhibited by cytokine-induced nitric oxide release. *Infect Immun* **72**: 359-363, 2004.
- FINKEL MS, ODDIS CV, JACOB TD, WATKINS SC, HATTLER BG, SIMMONS RL: Negative inotropic effects of cytokines on the heart mediated by nitric oxide. *Science* **257**: 387-389, 1992.
- FLOREZ O, ZAFRA G, MORILLO C, MARTIN J, GONZALEZ CI: Interleukin-1 gene cluster polymorphism in chagas disease in a Colombian case-control study. *Hum Immunol* **67**: 741-748, 2006.
- FRANCIS SE, CAMP NJ, DEWBERRY RM, GUNN J, SYRRIS P, CARTER ND, JEFFERY S, KASKI JC, CUMBERLAND DC, DUFF GW, CROSSMAN DC: Interleukin-1 receptor antagonist gene polymorphism and coronary artery disease. *Circulation* **99**: 861-866, 1999.
- FRANCIS SE, HOLDEN H, HOLT CM, DUFF GW: Interleukin-1 in myocardium and coronary arteries of patients with dilated cardiomyopathy. *J Mol Cell Cardiol* **30**: 215-223, 1998.
- GADINA M, JEFFERIES CA: IL-33: a sheep in wolf's clothing? *Sci STKE* **2007**: pe31, 2007.
- GALIS ZS, MUSZYNSKI M, SUKHOVA GK, SIMON-MORRISSEY E, LIBBY P: Enhanced expression of vascular matrix metalloproteinases induced in vitro by cytokines and in regions of human atherosclerotic lesions. *Ann NY Acad Sci* **748**: 501-507, 1995.
- GALIS ZS, MUSZYNSKI M, SUKHOVA GK, SIMON-MORRISSEY E, UNEMORI EN, LARK MW, AMENTO E, LIBBY P: Cytokine-stimulated human vascular smooth muscle cells synthesize a complement of enzymes required for extracellular matrix digestion. *Circ Res* **75**: 181-189, 1994.
- GERY I, WAKSMAN BH: Potentiation of the T-lymphocyte response to mitogens. II. The cellular source of potentiating mediator(s). *J Exp Med* **136**: 143-155, 1972.
- GIACHELLI CM, BAE N, ALMEIDA M, DENHARDT DT, ALPERS CE, SCHWARTZ SM: Osteopontin is elevated during neointima formation in rat arteries and is a novel component of human atherosclerotic plaques. *J Clin Invest* **92**: 1686-1696, 1993.
- GREENFEDER SA, NUNES P, KWEE L, LABOW M, CHIZZONITE RA, JU G: Molecular cloning and characterization of a second subunit of the interleukin 1 receptor complex. *J Biol Chem* **270**: 13757-13765, 1995.
- GRUN K, MARKOVA B, BOHMER FD, BERNDT A, KOSMEHL H, LEIPNER C: Elevated expression of PDGF-C in coxsackievirus B3-induced chronic myocarditis. *Eur Heart J* **26**: 728-739, 2005.
- GUILLEN I, BLANES M, GOMEZ-LECHON MJ, CASTELL JV: Cytokine signaling during myocardial infarction: sequential appearance of IL-1 beta and IL-6. *Am J Physiol* **269**: R229-R235, 1995.
- GULICK T, CHUNG MK, PIEPER SJ, LANGE LG, SCHREINER GF: Interleukin 1 and tumor necrosis factor inhibit cardiac myocyte beta-adrenergic responsiveness. *Proc Natl Acad Sci USA* **86**: 6753-6757, 1989.
- HACHAM M, ARGOV S, WHITE RM, SEGAL S, APTE RN: Different patterns of interleukin-1alpha and interleukin-1beta expression in organs of normal young and old mice. *Eur Cytokine Netw* **13**: 55-65, 2002.
- HANADA M, NINOMIYA-TSUJI J, KOMAKI K, OHNISHI M, KATSURA K, KANAMARU R, MATSUMOTO K, TAMURA S: Regulation of the TAK1 signaling pathway by protein phosphatase 2C. *J Biol Chem* **276**: 5753-5759, 2001.
- HANEMAAIJER R, KOOLWIJK P, LE CLERCQ L, DE VREE WJ, VAN HINSBERGH VW: Regulation of matrix metalloproteinase expression in human vein and microvascular endothelial cells. Effects of tumour necrosis factor alpha, interleukin 1 and phorbol ester. *Biochem J* **296**: 803-809, 1993.
- HENRICHOT E, JUGE-AUBRY CE, PERNIN A, PACHE JC, VELEBIT V, DAYER JM, MEDA P, CHIZZOLINI C, MEIER CA: Production of chemokines by perivascular adipose tissue: a role in the pathogenesis of atherosclerosis? *Arterioscler Thromb Vasc Biol* **25**: 2594-2599, 2005.

- HERMAN MP, SUKHOVA GK, LIBBY P, GERDES N, TANG N, HORTON DB, KILBRIDE M, BREITBART RE, CHUN M, SCHONBECK U: Expression of neutrophil collagenase (matrix metalloproteinase-8) in human atheroma: a novel collagenolytic pathway suggested by transcriptional profiling. *Circulation* **104**: 1899-1904, 2001.
- HEYMANS S, LUTTUN A, NUYENS D, THEILMEIER G, CREEMERS E, MOONS L, DYSPERSIN GD, CLEUTJENS JP, SHIPLEY M, ANGELLILO A, LEVI M, NUBE O, BAKER A, KESHET E, LUPU F, HERBERT JM, SMITS JF, SHAPIRO SD, BAES M, BORGERS M, COLLEN D, DAEMEN MJ, CARMELIET P: Inhibition of plasminogen activators or matrix metalloproteinases prevents cardiac rupture but impairs therapeutic angiogenesis and causes cardiac failure. *Nat Med* **5**: 1135-1142, 1999.
- HOGUE M, AMAR S: Role of interleukin-1 in bacterial atherogenesis. *Timely Top Med Cardiovasc Dis* **11**: E5, 2007.
- HORIO T, NISHIKIMI T, YOSHIHARA F, NAGAYA N, MATSUO H, TAKISHITA S, KANGAWA K: Production and secretion of adrenomedullin in cultured rat cardiac myocytes and nonmyocytes: stimulation by interleukin-1beta and tumor necrosis factor-alpha. *Endocrinology* **139**: 4576-4580, 1998.
- IACOVIELLO L, DI CASTELNUOVO A, GATTONE M, PEZZINI A, ASSANELLI D, LORENZET R, DEL ZOTTO E, COLOMBO M, NAPOLEONE E, AMORE C, D'ORAZIO A, PADOVANI A, DE GAETANO G, GIANNUZZI P, DONATI MB: Polymorphisms of the interleukin-1beta gene affect the risk of myocardial infarction and ischemic stroke at young age and the response of mononuclear cells to stimulation *in vitro*. *Arterioscler Thromb Vasc Biol* **25**: 222-227, 2005.
- ICHIHARA S, YAMADA Y, ICHIHARA G, KANAZAWA H, HASHIMOTO K, KATO Y, MATSUSHITA A, OIKAWA S, YOKOTA M, IWASE M: Attenuation of oxidative stress and cardiac dysfunction by bisoprolol in an animal model of dilated cardiomyopathy. *Biochem Biophys Res Commun* **350**: 105-113, 2006.
- ISODA K, KAMEZAWA Y, TADA N, SATO M, OHSUZU F: Myocardial hypertrophy in transgenic mice overexpressing human interleukin 1alpha. *J Card Fail* **7**: 355-364, 2001.
- ISODA K, OHSUZU F: The effect of interleukin-1 receptor antagonist on arteries and cholesterol metabolism. *J Atheroscler Thromb* **13**: 21-30, 2006.
- ISODA K, SHIIGAI M, ISHIGAMI N, MATSUKI T, HORAI R, NISHIKAWA K, KUSUHARA M, NISHIDA Y, IWAKURA Y, OHSUZU F: Deficiency of interleukin-1 receptor antagonist promotes neointimal formation after injury. *Circulation* **108**: 516-518, 2003.
- JANSSENS S, BEYAERT R: Functional diversity and regulation of different interleukin-1 receptor-associated kinase (IRAK) family members. *Mol Cell* **11**: 293-302, 2003.
- JENSEN LE, WHITEHEAD AS: IRAK1b, a novel alternative splice variant of interleukin-1 receptor-associated kinase (IRAK), mediates interleukin-1 signaling and has prolonged stability. *J Biol Chem* **276**: 29037-29044, 2001.
- JIANG Z, JOHNSON HJ, NIE H, QIN J, BIRD TA, LI X: Pellino 1 is required for interleukin-1 (IL-1)-mediated signaling through its interaction with the IL-1 receptor-associated kinase 4 (IRAK4)-IRAK-tumor necrosis factor receptor-associated factor 6 (TRAF6) complex. *J Biol Chem* **278**: 10952-10956, 2003.
- JIANG Z, NINOMIYA-TSUJI J, QIAN Y, MATSUMOTO K, LI X: Interleukin-1 (IL-1) receptor-associated kinase-dependent IL-1-induced signaling complexes phosphorylate TAK1 and TAB2 at the plasma membrane and activate TAK1 in the cytosol. *Mol Cell Biol* **22**: 7158-7167, 2002.
- JUGE-AUBRY CE, SOMM E, GIUSTI V, PERNIN A, CHICHEPORTICHE R, VERDUMO C, ROHNER-JEANRENAUD F, BURGER D, DAYER JM, MEIER CA: Adipose tissue is a major source of interleukin-1 receptor antagonist: upregulation in obesity and inflammation. *Diabetes* **52**: 1104-1110, 2003.
- KAKKAR R, LEE RT: The IL-33/ST2 pathway: therapeutic target and novel biomarker. *Nat Rev Drug Discov* **7**: 827-840, 2008.
- KAMARI Y, WERMAN-VENKERT R, SHAISH A, WERMAN A, HARARI A, GONEN A, VORONOV E, GROSSKOPF I, SHARABI Y, GROSSMAN E, IWAKURA Y, DINARELLO CA, APTE RN, HARATS D: Differential role and tissue specificity of interleukin-1alpha gene expression in atherogenesis and lipid metabolism. *Atherosclerosis* **195**: 31-38, 2007.
- KAVITA U, MIZEL SB: Differential sensitivity of interleukin-1 alpha and -beta precursor proteins to cleavage by calpain, a calcium-dependent protease. *J Biol Chem* **270**: 27758-27765, 1995.
- KAWAGUCHI Y: IL-1 alpha gene expression and protein production by fibroblasts from patients with systemic sclerosis. *Clin Exp Immunol* **97**: 445-450, 1994.

- KAWAGUCHI Y, NISHIMAGI E, TOCHIMOTO A, KAWAMOTO M, KATSUMATA Y, SOEJIMA M, KANNO T, KAMATANI N, HARA M: Intracellular IL-1 α -binding proteins contribute to biological functions of endogenous IL-1 α in systemic sclerosis fibroblasts. *Proc Natl Acad Sci USA* **103**: 14501-14506, 2006.
- KHURANA R, SIMONS M, MARTIN JF, ZACHARY IC: Role of angiogenesis in cardiovascular disease: a critical appraisal. *Circulation* **112**: 1813-1824, 2005.
- KIRII H, NIWA T, YAMADA Y, WADA H, SAITO K, IWAKURA Y, ASANO M, MORIWAKI H, SEISHIMA M: Lack of interleukin-1 β decreases the severity of atherosclerosis in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* **23**: 656-660, 2003.
- KISHIMOTO K, MATSUMOTO K, NINOMIYA-TSUJI J: TAK1 mitogen-activated protein kinase kinase is activated by autophosphorylation within its activation loop. *J Biol Chem* **275**: 7359-7364, 2000.
- KOBAYASHI K, HERNANDEZ LD, GALAN JE, JANEWAY CA, JR., MEDZHITOV R, FLAVELL RA: IRAK-M is a negative regulator of Toll-like receptor signaling. *Cell* **110**: 191-202, 2002.
- KOLLEWE C, MACKENSEN AC, NEUMANN D, KNOP J, CAO P, LI S, WESCHE H, MARTIN MU: Sequential autophosphorylation steps in the interleukin-1 receptor-associated kinase-1 regulate its availability as an adapter in interleukin-1 signaling. *J Biol Chem* **279**: 5227-5236, 2004.
- KORNMAN KS: Interleukin 1 genetics, inflammatory mechanisms, and nutrigenetic opportunities to modulate diseases of aging. *Am J Clin Nutr* **83**: 475S-483S, 2006.
- KORNMAN KS, DUFF GW: Candidate genes as potential links between periodontal and cardiovascular diseases. *Ann Periodontol* **6**: 48-57, 2001.
- KORNMAN KS, MARTHA PM, DUFF GW: Genetic variations and inflammation: a practical nutrigenomics opportunity. *Nutrition* **20**: 44-49, 2004.
- KORNMAN KS, PANKOW J, OFFENBACHER S, BECK J, Di GIOVINE F, DUFF GW: Interleukin-1 genotypes and the association between periodontitis and cardiovascular disease. *J Periodontal Res* **34**: 353-357, 1999.
- LAI J, ZHOU D, XIA S, SHANG Y, ZHU J, PAN J, HUA B, ZHU Y, CUI L: Association of interleukin-1 gene cluster polymorphisms with ischemic stroke in a Chinese population. *Neurol India* **54**: 366-369, 2006.
- LANE JR, NEUMANN DA, LAFOND-WALKER A, HERSKOWITZ A, ROSE NR: Interleukin 1 or tumor necrosis factor can promote Cocksackie B3-induced myocarditis in resistant B10.A mice. *J Exp Med* **175**: 1123-1129, 1992.
- LATKOVSKIS G, LICIS N, KALNINS U: C-reactive protein levels and common polymorphisms of the interleukin-1 gene cluster and interleukin-6 gene in patients with coronary heart disease. *Eur J Immunogenet* **31**: 207-213, 2004.
- LEE BC, AHN SY, DOO HK, YIM SV, LEE HJ, JIN SY, HONG SJ, LEE SH, KIM SD, SEO JC, LEEM KH, CHUNG JH: Susceptibility for ischemic stroke in Korean population is associated with polymorphisms of the interleukin-1 receptor antagonist and tumor necrosis factor- α genes, but not the interleukin-1 β gene. *Neurosci Lett* **357**: 33-36, 2004.
- LEE E, GRODZINSKY AJ, LIBBY P, CLINTON SK, LARK MW, LEE RT: Human vascular smooth muscle cell-monocyte interactions and metalloproteinase secretion in culture. *Arterioscler Thromb Vasc Biol* **15**: 2284-2289, 1995.
- LEE FS, HAGLER J, CHEN ZJ, MANIATIS T: Activation of the I κ B α kinase complex by MEKK1, a kinase of the JNK pathway. *Cell* **88**: 213-222, 1997.
- LI MG, KATSURA K, NOMIYAMA H, KOMAKI K, NINOMIYA-TSUJI J, MATSUMOTO K, KOBAYASHI T, TAMURA S: Regulation of the interleukin-1-induced signaling pathways by a novel member of the protein phosphatase 2C family (PP2Cepsilon). *J Biol Chem* **278**: 12013-12021, 2003.
- LI S, STRELOW A, FONTANA EJ, WESCHE H: IRAK-4: a novel member of the IRAK family with the properties of an IRAK-kinase. *Proc Natl Acad Sci U S A* **99**: 5567-5572, 2002.
- LIBBY P: Inflammation in atherosclerosis. *Nature* **420**: 868-874, 2002.
- LIBBY P, SUKHOVA G, LEE RT, GALIS ZS: Cytokines regulate vascular functions related to stability of the atherosclerotic plaque. *J Cardiovasc Pharmacol* **25** (Suppl 2): S9-S12, 1995.
- LIM BK, CHOE SC, SHIN JO, HO SH, KIM JM, YU SS, KIM S, JEON ES: Local expression of interleukin-1 receptor antagonist by plasmid DNA improves mortality and decreases myocardial inflammation in experimental coxsackieviral myocarditis. *Circulation* **105**: 1278-1281, 2002.

- LOPEZ NJ, JARA L, VALENZUELA CY: Association of interleukin-1 polymorphisms with periodontal disease. *J Periodontol* **76**: 234-243, 2005.
- LYE E, MIRTSOS C, SUZUKI N, SUZUKI S, YEH WC: The role of interleukin 1 receptor-associated kinase-4 (IRAK-4) kinase activity in IRAK-4-mediated signaling. *J Biol Chem* **279**: 40653-40658, 2004.
- MACFELDA K, WEISS TW, KAUN C, BREUSS JM, ZORN G, OBERNDORFER U, VOEGELE-KADLETZ M, HUBER-BECKMANN R, ULLRICH R, BINDER BR, LOSERT UM, MAURER G, PACHER R, HUBER K, WOJTA J: Plasminogen activator inhibitor 1 expression is regulated by the inflammatory mediators interleukin-1alpha, tumor necrosis factor-alpha, transforming growth factor-beta and oncostatin M in human cardiac myocytes. *J Mol Cell Cardiol* **34**: 1681-1691, 2002.
- MAIER JA, STATUTO M, RAGNOTTI G: Endogenous interleukin 1 alpha must be transported to the nucleus to exert its activity in human endothelial cells. *Mol Cell Biol* **14**: 1845-1851, 1994.
- MANDINOVA A, SOLDI R, GRAZIANI I, BAGALA C, BELLUM S, LANDRISCINA M, TARANTINI F, PRUDOVSKY I, MACIAG T: S100A13 mediates the copper-dependent stress-induced release of IL-1alpha from both human U937 and murine NIH 3T3 cells. *J Cell Sci* **116**: 2687-2696, 2003.
- MATSUMORI A, IGATA H, ONO K, IWASAKI A, MIYAMOTO T, NISHIO R, SASAYAMA S: High doses of digitalis increase the myocardial production of proinflammatory cytokines and worsen myocardial injury in viral myocarditis: a possible mechanism of digitalis toxicity. *Jpn Circ J* **63**: 934-940, 1999.
- MATSUMORI A, YAMADA T, SUZUKI H, MATOBA Y, SASAYAMA S: Increased circulating cytokines in patients with myocarditis and cardiomyopathy. *Br Heart J* **72**: 561-566, 1994.
- McMAHON GA, GARFINKEL S, PRUDOVSKY I, HU X, MACIAG T: Intracellular precursor interleukin (IL)-1alpha, but not mature IL-1alpha, is able to regulate human endothelial cell migration in vitro. *J Biol Chem* **272**: 28202-28205, 1997.
- MEIER CA, BOBBIONI E, GABAY C, ASSIMACOPOULOS-JEANNET F, GOLAY A, DAYER JM: IL-1 receptor antagonist serum levels are increased in human obesity: a possible link to the resistance to leptin? *J Clin Endocrinol Metab* **87**: 1184-1188, 2002.
- MERHI-SOUSSI F, KWAK BR, MAGNE D, CHADJICHRISTOS C, BERTI M, PELLI G, JAMES RW, MACH F, GABAY C: Interleukin-1 plays a major role in vascular inflammation and atherosclerosis in male apolipoprotein E-knockout mice. *Cardiovasc Res* **66**: 583-593, 2005.
- METYAS SK, HOFFMAN HM: Anakinra prevents symptoms of familial cold autoinflammatory syndrome and Raynaud's disease. *J Rheumatol* **33**: 2085-2087, 2006.
- MIKAMI S, KAWASHIMA S, KANAZAWA K, HIRATA K, HOTTA H, HAYASHI Y, ITOH H, YOKOYAMA M: Low-dose N omega-nitro-L-arginine methyl ester treatment improves survival rate and decreases myocardial injury in a murine model of viral myocarditis induced by coxsackievirus B3. *Circ Res* **81**: 504-511, 1997.
- MIKAMI S, KAWASHIMA S, KANAZAWA K, HIRATA K, KATAYAMA Y, HOTTA H, HAYASHI Y, ITO H, YOKOYAMA M: Expression of nitric oxide synthase in a murine model of viral myocarditis induced by coxsackievirus B3. *Biochem Biophys Res Commun* **220**: 983-989, 1996.
- MIYAO Y, YASUE H, OGAWA H, MISUMI I, MASUDA T, SAKAMOTO T, MORITA E: Elevated plasma interleukin-6 levels in patients with acute myocardial infarction. *Am Heart J* **126**: 1299-1304, 1993.
- MOMIYAMA Y, HIRANO R, TANIGUCHI H, NAKAMURA H, OHSUZU F: Effects of interleukin-1 gene polymorphisms on the development of coronary artery disease associated with Chlamydia pneumoniae infection. *J Am Coll Cardiol* **38**: 712-717, 2001.
- MOMIYAMA Y, OHMORI R, OHSUZU F: Association between IL-1beta gene polymorphism and myocardial infarction. *Arterioscler Thromb Vasc Biol* **25**: e36, 2005.
- MORTON AC, ARNOLD ND, GUNN J, VARCOE R, FRANCIS SE, DOWER SK, CROSSMAN DC: Interleukin-1 receptor antagonist alters the response to vessel wall injury in a porcine coronary artery model. *Cardiovasc Res* **68**: 493-501, 2005.
- MOSLEY B, URDAL DL, PRICKETT KS, LARSEN A, COSMAN D, CONLON PJ, GILLIS S, DOWER SK: The interleukin-1 receptor binds the human interleukin-1 alpha precursor but not the interleukin-1 beta precursor. *J Biol Chem* **262**: 2941-2944, 1987.

- MURTUZA B, SUZUKI K, BOU-GHARIOS G, BEAUCHAMP JR, SMOLENSKI RT, PARTRIDGE TA, YACCOUB MH: Transplantation of skeletal myoblasts secreting an IL-1 inhibitor modulates adverse remodeling in infarcted murine myocardium. *Proc Natl Acad Sci USA* **101**: 4216-4221, 2004.
- MUZIO M, NI J, FENG P, DIXIT VM: IRAK (Pelle) family member IRAK-2 and MyD88 as proximal mediators of IL-1 signaling. *Science* **278**: 1612-1615, 1997.
- NAKAE S, ASANO M, HORAI R, IWAKURA Y: Interleukin-1 beta, but not interleukin-1 alpha, is required for T-cell-dependent antibody production. *Immunology* **104**: 402-409, 2001.
- NAKANO A, MATSUMORI A, KAWAMOTO S, TAHARA H, YAMATO E, SASAYAMA S, MIYAZAKI JI: Cytokine gene therapy for myocarditis by in vivo electroporation. *Hum Gene Ther* **12**: 1289-1297, 2001.
- NALDINI A, LEALI D, PUCCI A, MORENA E, CARRARO F, NICO B, RIBATTI D, PRESTA M: Cutting edge: IL-1beta mediates the proangiogenic activity of osteopontin-activated human monocytes. *J Immunol* **177**: 4267-4270, 2006.
- NEWBY AC: Dual role of matrix metalloproteinases (matrixins) in intimal thickening and atherosclerotic plaque rupture. *Physiol Rev* **85**: 1-31, 2005.
- NICKLIN MJ, HUGHES DE, BARTON JL, URE JM, DUFF GW: Arterial inflammation in mice lacking the interleukin 1 receptor antagonist gene. *J Exp Med* **191**: 303-312, 2000.
- NICOLL JA, MRAK RE, GRAHAM DI, STEWART J, WILCOCK G, MACGOWAN S, ESIRI MM, MURRAY LS, DEWAR D, LOVE S, MOSS T, GRIFFIN WS: Association of interleukin-1 gene polymorphisms with Alzheimer's disease. *Ann Neurol* **47**: 365-368, 2000.
- NINOMIYA-TSUJI J, KISHIMOTO K, HIYAMA A, INOUE J, CAO Z, MATSUMOTO K: The kinase TAK1 can activate the NIK-I kappaB as well as the MAP kinase cascade in the IL-1 signalling pathway. *Nature* **398**: 252-256, 1999.
- OKUNO M, NAKAGAWA M, SHIMADA M, SAITO M, HISHINUMA S, YAMAUCHI-TAKIHARA K: Expressional patterns of cytokines in a murine model of acute myocarditis: early expression of cardiotrophin-1. *Lab Invest* **80**: 433-440, 2000.
- PALEOLOG EM: Angiogenesis in rheumatoid arthritis. *Arthritis Res* **4** (Suppl 3): S81-S90, 2002.
- PATTI G, D'AMBROSIO A, MEGA S, GIORGI G, ZARDI EM, ZARDI DM, DICUONZO G, DOBRINA A, Di SCIASCIO G: Early interleukin-1 receptor antagonist elevation in patients with acute myocardial infarction. *J Am Coll Cardiol* **43**: 35-38, 2004.
- PELEGRIN P, SURPRENANT A: Pannexin-1 mediates large pore formation and interleukin-1beta release by the ATP-gated P2X7 receptor. *EMBO J* **25**: 5071-5082, 2006.
- PINSKY DJ, CAI B, YANG X, RODRIGUEZ C, SCIACCA RR, CANNON PJ: The lethal effects of cytokine-induced nitric oxide on cardiac myocytes are blocked by nitric oxide synthase antagonism or transforming growth factor beta. *J Clin Invest* **95**: 677-685, 1995.
- POLLOCK AS, TURCK J, LOVETT DH: The prodomain of interleukin 1alpha interacts with elements of the RNA processing apparatus and induces apoptosis in malignant cells. *FASEB J* **17**: 203-213, 2003.
- PRABHU SD, CHANDRASEKAR B, MURRAY DR, FREEMAN GL: beta-adrenergic blockade in developing heart failure: effects on myocardial inflammatory cytokines, nitric oxide, and remodeling. *Circulation* **101**: 2103-2109, 2000.
- QING M, SCHUMACHER K, HEISE R, WOLTJE M, VAZQUEZ-JIMENEZ JF, RICHTER T, ARRANDA-CARRERO M, HESS J, VON BERNUTH G, SEGHAJE MC: Intramyocardial synthesis of pro- and anti-inflammatory cytokines in infants with congenital cardiac defects. *J Am Coll Cardiol* **41**: 2266-2274, 2003.
- RAINERO I, BO M, FERRERO M, VALFRE W, VAULA G, PINESSI L: Association between the interleukin-1alpha gene and Alzheimer's disease: a meta-analysis. *Neurobiol Aging* **25**: 1293-1298, 2004.
- RAJAVASHISTH TB, LIAO JK, GALIS ZS, TRIPATHI S, LAUFS U, TRIPATHI J, CHAI NN, XU XP, JOVINGE S, SHAH PK, LIBBY P: Inflammatory cytokines and oxidized low density lipoproteins increase endothelial cell expression of membrane type 1-matrix metalloproteinase. *J Biol Chem* **274**: 11924-11929, 1999a.
- RAJAVASHISTH TB, XU XP, JOVINGE S, MEISEL S, XU XO, CHAI NN, FISHBEIN MC, KAUL S, CERCEK B, SHARIFI B, SHAH PK: Membrane type 1 matrix metalloproteinase expression in human atherosclerotic plaques: evidence for activation by proinflammatory mediators. *Circulation* **99**: 3103-3109, 1999b.

- RAO N, NGUYEN S, NGO K, FUNG-LEUNG WP: A novel splice variant of interleukin-1 receptor (IL-1R)-associated kinase 1 plays a negative regulatory role in Toll/IL-1R-induced inflammatory signaling. *Mol Cell Biol* **25**: 6521-6532, 2005.
- RUBARTELLI A, COZZOLINO F, TALIO M, SITIA R: A novel secretory pathway for interleukin-1 beta, a protein lacking a signal sequence. *EMBO J* **9**: 1503-1510, 1990.
- SALVEN P, HATTORI K, HEISSIG B, RAFII S: Interleukin-1alpha promotes angiogenesis *in vivo* via VEGFR-2 pathway by inducing inflammatory cell VEGF synthesis and secretion. *FASEB J* **16**: 1471-1473, 2002.
- SHAH PK, FALK E, BADIMON JJ, FERNANDEZ-ORTIZ A, MAILHAC A, VILLAREAL-LEVY G, FALLON JT, REGNSTROM J, FUSTER V: Human monocyte-derived macrophages induce collagen breakdown in fibrous caps of atherosclerotic plaques. Potential role of matrix-degrading metalloproteinases and implications for plaque rupture. *Circulation* **92**: 1565-1569, 1995.
- SHIOI T, MATSUMORI A, SASAYAMA S: Persistent expression of cytokine in the chronic stage of viral myocarditis in mice. *Circulation* **94**: 2930-2937, 1996.
- SCHWAGER I, JUNGI TW: Effect of human recombinant cytokines on the induction of macrophage procoagulant activity. *Blood* **83**: 152-160, 1994.
- SINGH K, BALLIGAND JL, FISCHER TA, SMITH TW, KELLY RA: Regulation of cytokine-inducible nitric oxide synthase in cardiac myocytes and microvascular endothelial cells. Role of extracellular signal-regulated kinases 1 and 2 (ERK1/ERK2) and STAT1 alpha. *J Biol Chem* **271**: 1111-1117, 1996.
- SO A, DE SMEDT T, REVAZ S, TSCHOPP J: A pilot study of IL-1 inhibition by anakinra in acute gout. *Arthritis Res Ther* **9**: R28, 2007.
- SOMM E, HENRICHOT E, PERNIN A, JUGE-AUBRY CE, MUZZIN P, DAYER JM, NICKLIN MJ, MEIER CA: Decreased fat mass in interleukin-1 receptor antagonist-deficient mice: impact on adipogenesis, food intake, and energy expenditure. *Diabetes* **54**: 3503-3509, 2005.
- SONG X, VORONOV E, DVORKIN T, FIMA E, CAGNANO E, BENHARROCH D, SHENDLER Y, BJORKDAHL O, SEGAL S, DINARELLO CA, APTE RN: Differential effects of IL-1 alpha and IL-1 beta on tumorigenicity patterns and invasiveness. *J Immunol* **171**: 6448-6456, 2003.
- STEVENSON FT, BURSTEN SL, FANTON C, LOCKSLEY RM, LOVETT DH: The 31-kDa precursor of interleukin 1 alpha is myristoylated on specific lysines within the 16-kDa N-terminal propeptide. *Proc Natl Acad Sci USA* **90**: 7245-7249, 1993.
- SUTTON C, BRERETON C, KEOGH B, MILLS KH, LAVELLE EC: A crucial role for interleukin (IL)-1 in the induction of IL-17-producing T cells that mediate autoimmune encephalomyelitis. *J Exp Med* **203**: 1685-1691, 2006.
- SUZUKI K, MURTUZA B, BEAUCHAMP JR, BRAND NJ, BARTON PJ, VARELA-CARVER A, FUKUSHIMA S, COPPEN SR, PARTRIDGE TA, YACCOUB MH: Role of interleukin-1beta in acute inflammation and graft death after cell transplantation to the heart. *Circulation* **110** (11 Suppl 1): II219-II224, 2004.
- TAKAESU G, NINOMIYA-TSUJI J, KISHIDA S, LI X, STARK GR, MATSUMOTO K: Interleukin-1 (IL-1) receptor-associated kinase leads to activation of TAK1 by inducing TAB2 translocation in the IL-1 signaling pathway. *Mol Cell Biol* **21**: 2475-2484, 2001.
- TAKATSUNA H, KATO H, GOHDA J, AKIYAMA T, MORIYA A, OKAMOTO Y, YAMAGATA Y, OTSUKA M, UMEZAWA K, SEMBA K, INOUE J: Identification of TIFA as an adapter protein that links tumor necrosis factor receptor-associated factor 6 (TRAF6) to interleukin-1 (IL-1) receptor-associated kinase-1 (IRAK-1) in IL-1 receptor signaling. *J Biol Chem* **278**: 12144-12150, 2003.
- TANAKA T, KANAI H, SEKIGUCHI K, AIHARA Y, YOKOYAMA T, ARAI M, KANDA T, NAGAI R, KURABAYASHI M: Induction of VEGF gene transcription by IL-1 beta is mediated through stress-activated MAP kinases and Sp1 sites in cardiac myocytes. *J Mol Cell Cardiol* **32**: 1955-1967, 2000.
- TELLEZ N, MONTOLIO M, BIARNES M, CASTANO E, SOLER J, MONTANYA E: Adenoviral overexpression of interleukin-1 receptor antagonist protein increases beta-cell replication in rat pancreatic islets. *Gene Ther* **12**: 120-128, 2005.
- THAIK CM, CALDERONE A, TAKAHASHI N, COLUCCI WS: Interleukin-1 beta modulates the growth and phenotype of neonatal rat cardiac myocytes. *J Clin Invest* **96**: 1093-1099, 1995.

- THALMANN S, MEIER CA: Local adipose tissue depots as cardiovascular risk factors. *Cardiovasc Res* **75**: 690-701, 2007.
- THORNTON RD, LANE P, BORGHAEI RC, PEASE EA, CARO J, MOCHAN E: Interleukin 1 induces hypoxia-inducible factor 1 in human gingival and synovial fibroblasts. *Biochem J* **350**: 307-312, 2000.
- TIPPING PG, HANCOCK WW: Production of tumor necrosis factor and interleukin-1 by macrophages from human atheromatous plaques. *Am J Pathol* **142**: 1721-1728, 1993.
- UM JY, JEONG HJ, PARK RK, HONG SH, KIM HM: Aspects of gene polymorphisms in cerebral infarction: inflammatory cytokines. *Cell Mol Life Sci* **62**: 824-833, 2005.
- VANDERHEYDEN M, PAULUS WJ, VOSS M, KNUEFERMANN P, SIVASUBRAMANIAN N, MANN D, BAUMGARTEN G: Myocardial cytokine gene expression is higher in aortic stenosis than in idiopathic dilated cardiomyopathy. *Heart* **91**: 926-931, 2005.
- VON DER THUSEN JH, KUIPER J, VAN BERKEL TJ, BIESSEN EA: Interleukins in atherosclerosis: molecular pathways and therapeutic potential. *Pharmacol Rev* **55**: 133-166, 2003.
- VORONOV E, SHOVAL DS, KRELIN Y, CAGNANO E, BENHARROCH D, IWAKURA Y, DINARELLO CA, APTE RN: IL-1 is required for tumor invasiveness and angiogenesis. *Proc Natl Acad Sci USA* **100**: 2645-2650, 2003.
- WAEHRE T, YNDESTAD A, SMITH C, HAUG T, TUNHEIM SH, GULLESTAD L, FROLAND SS, SEMB AG, AUKRUST P, DAMAS JK: Increased expression of interleukin-1 in coronary artery disease with downregulatory effects of HMG-CoA reductase inhibitors. *Circulation* **109**: 1966-1972, 2004.
- WALSH DA, PEARSON CI: Angiogenesis in the pathogenesis of inflammatory joint and lung diseases. *Arthritis Res* **3**: 147-153, 2001.
- WANG C, DENG L, HONG M, AKKARAJU GR, INOUE J, CHEN ZJ: TAK1 is a ubiquitin-dependent kinase of MKK and IKK. *Nature* **412**: 346-351, 2001.
- WANG H, BLOOM O, ZHANG M, VISHNUBHAKAT JM, OMBRELLINO M, CHE J, FRAZIER A, YANG H, IVANOVA S, BOROVIKOVA L, MANOGUE KR, FAIST E, ABRAHAM E, ANDERSSON J, ANDERSSON U, MOLINA PE, ABUMRAD NN, SAMA A, TRACEY KJ: HMG-1 as a late mediator of endotoxin lethality in mice. *Science* **285**: 248-251, 1999a.
- WANG H, VISHNUBHAKAT JM, BLOOM O, ZHANG M, OMBRELLINO M, SAMA A, TRACEY KJ: Proinflammatory cytokines (tumor necrosis factor and interleukin 1) stimulate release of high mobility group protein-1 by pituicytes. *Surgery* **126**: 389-392, 1999b.
- WERMAN A, WERMAN-VENKERT R, WHITE R, LEE JK, WERMAN B, KRELIN Y, VORONOV E, DINARELLO CA, APTE RN: The precursor form of IL-1 α is an intracrine proinflammatory activator of transcription. *Proc Natl Acad Sci USA* **101**: 2434-2439, 2004.
- WESSENDORF JH, GARFINKEL S, ZHAN X, BROWN S, MACIAG T: Identification of a nuclear localization sequence within the structure of the human interleukin-1 α precursor. *J Biol Chem* **268**: 22100-22104, 1993.
- WESTPHAL E, ROHRBACH S, BUERKE M, BEHR H, DARMER D, SILBER RE, WERDAN K, LOPPNOW H: Altered interleukin-1 receptor antagonist and interleukin-18 mRNA expression in myocardial tissues of patients with dilated cardiomyopathy. *Mol Med* **14**: 55-63, 2008.
- YAMAGUCHI K, SHIRAKABE K, SHIBUYA H, IRIE K, OISHI I, UENO N, TANIGUCHI T, NISHIDA E, MATSUMOTO K: Identification of a member of the MAPKKK family as a potential mediator of TGF- β signal transduction. *Science* **270**: 2008-2011, 1995.
- YAMIN TT, MILLER DK: The interleukin-1 receptor-associated kinase is degraded by proteasomes following its phosphorylation. *J Biol Chem* **272**: 21540-21547, 1997.
- YUCESOY B, PEILA R, WHITE LR, WU KM, JOHNSON VJ, KASHON ML, LUSTER MI, LAUNER LJ: Association of interleukin-1 gene polymorphisms with dementia in a community-based sample: the Honolulu Asia Aging Study. *Neurobiol Aging* **27**: 211-217, 2006.