



Ústav fotoniky  
a elektroniky

# **OPTICAL FIBERS FOR SENSORS**

## **How to employ light for information harvesting?**

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# OUTLINE

- Optical fiber sensors - basic terms
- PCS fibers and ways for increasing the detection sensitivity
- Surface-plasmon fiber-optic sensors
- Gratings in optical fibers
- Fiber-optic tips and tapers

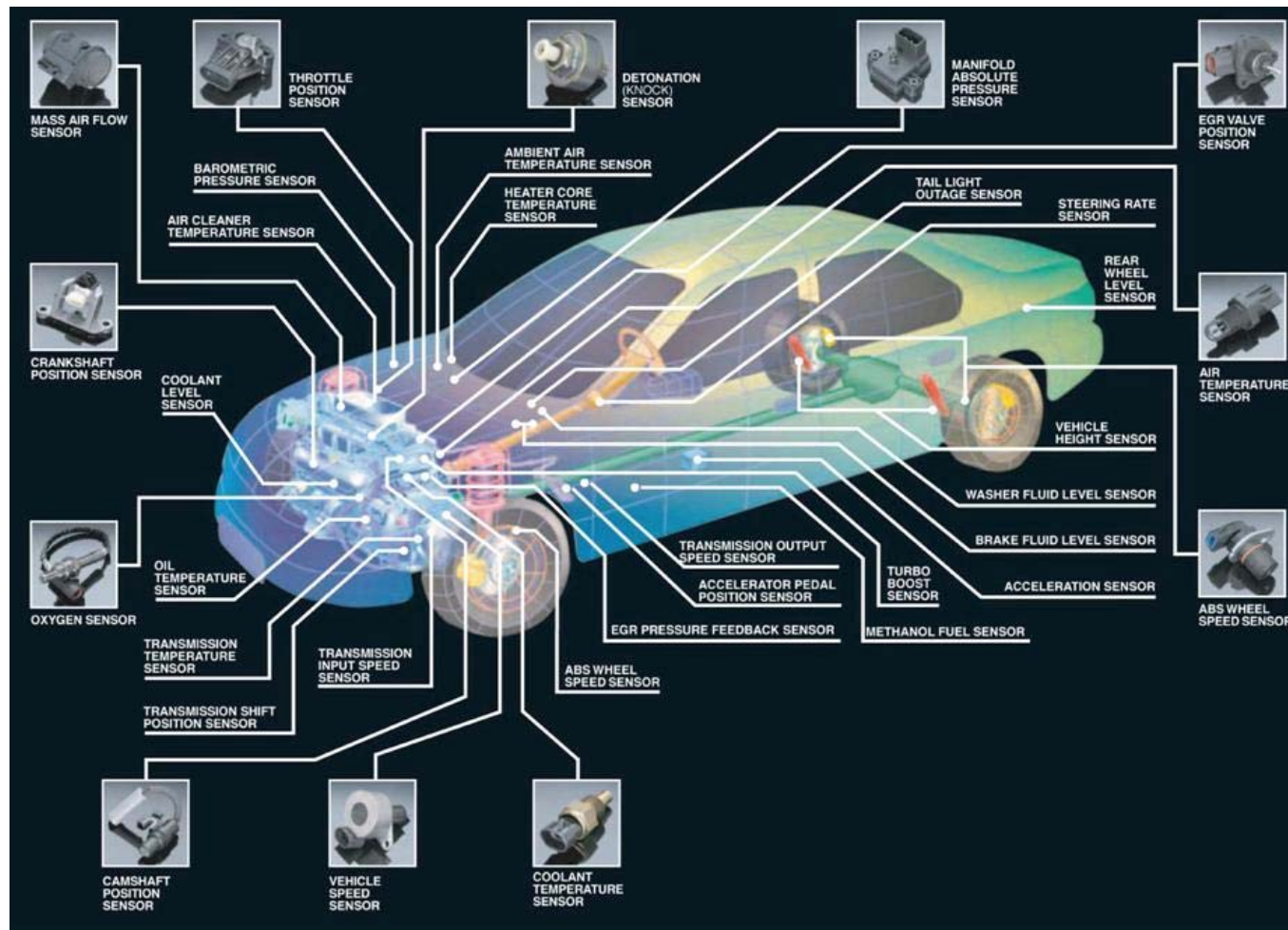
# INTRODUCTION

## **Optical fiber sensitivity to surrounding changes**

- Telecommunications – „in-sensitivity“ to changes in their surrounding (-40 – 50 °C)
- Sensors – controllable sensitive to changes in their surrounding

? Issues for development of special optical fibers sensitive (selectively) to surrounding changes

# SENSORS IN AUTOMOBILE



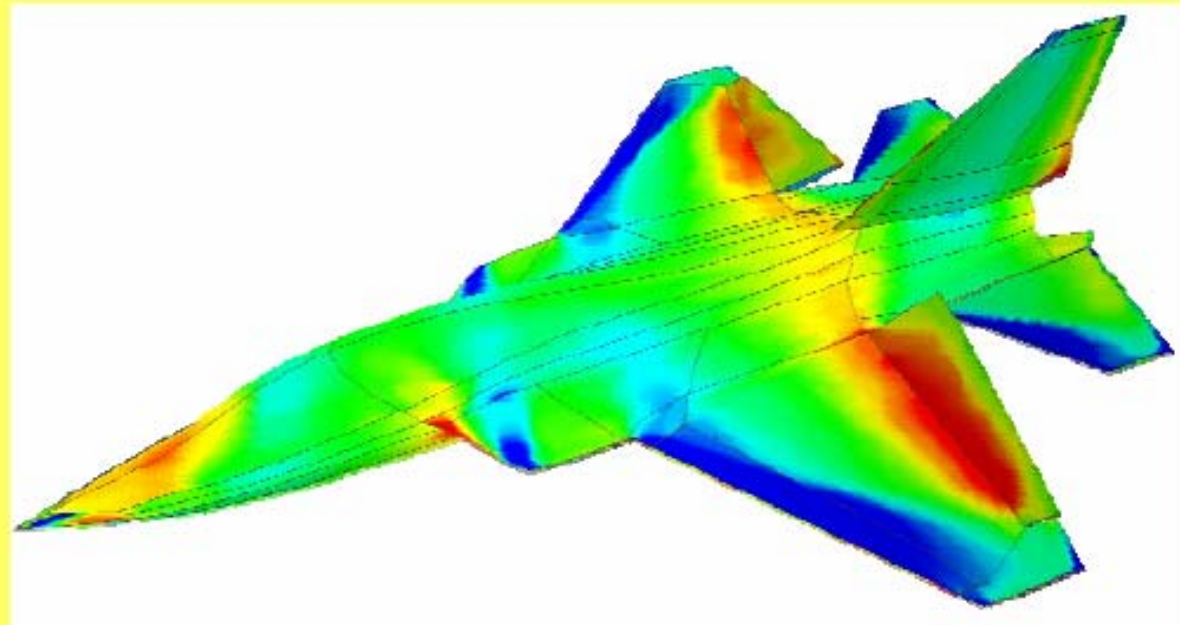
[http://www.accurateautomotiveaz.com/index.php?option=com\\_content&view=article&id=84:com..](http://www.accurateautomotiveaz.com/index.php?option=com_content&view=article&id=84:com..)

Detection of hydrogen leakages from hydrogen-based automobiles

# SENSORS IN AIRPLANES

- \* Pressure-sensitive Paints (PSPs) and
- \* Temperature-Sensitive Paints (TSPs)

Used to "photograph"  
pressure (in fact oxygen)  
or temperature



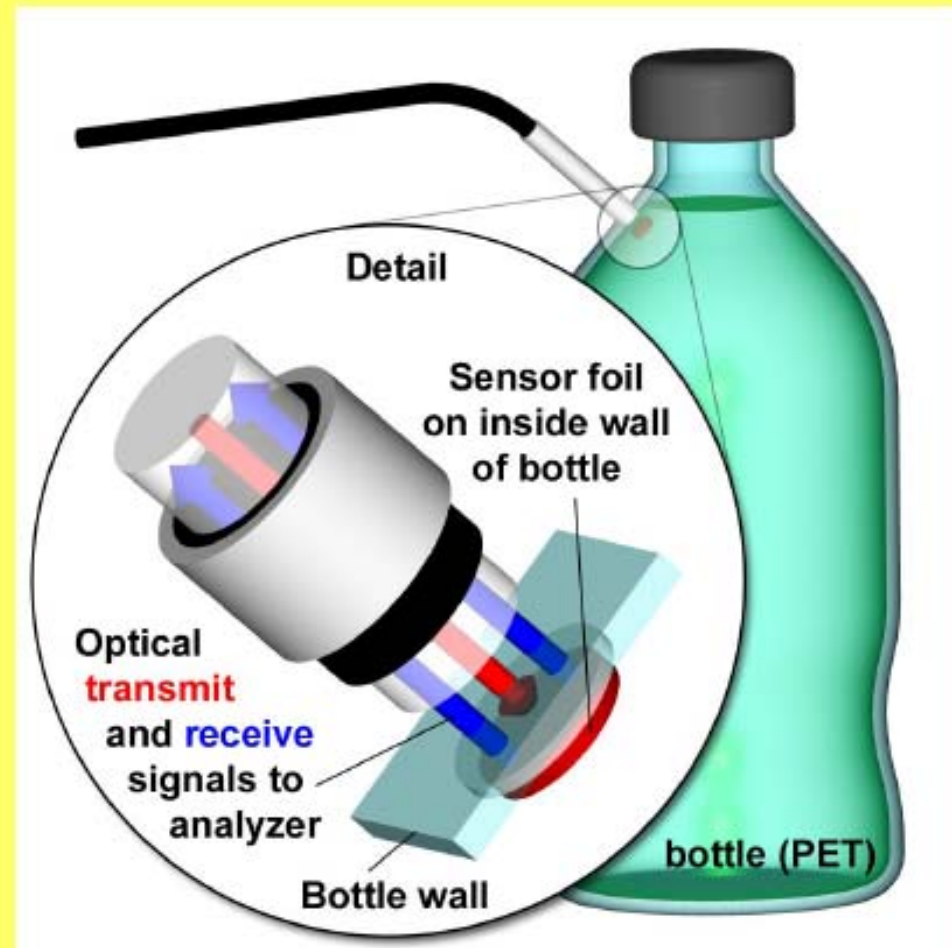
*Book: T. Liu, J. P. Sullivan, Pressure and Temperature Sensitive Paints, 2005, Springer.*

# SENSORS IN FOOD PRODUCTION

**PET is permeable to oxygen and CO<sub>2</sub>**

- \* CO<sub>2</sub> diffuses out
- \* O<sub>2</sub> diffuses in

**Sensor Spots Placed Inside the Bottle**



O.S. Wolfbeis, EUROPTRODE X, 2010, Prague

# SENSORS IN PHARMACY



**Product completeness**

**Product content**

<http://www.keyence.com/products/vision/vision-sensor/iv/applications/application-02.jsp>

# WHAT IS OPTICAL SENSOR?

**Def:**

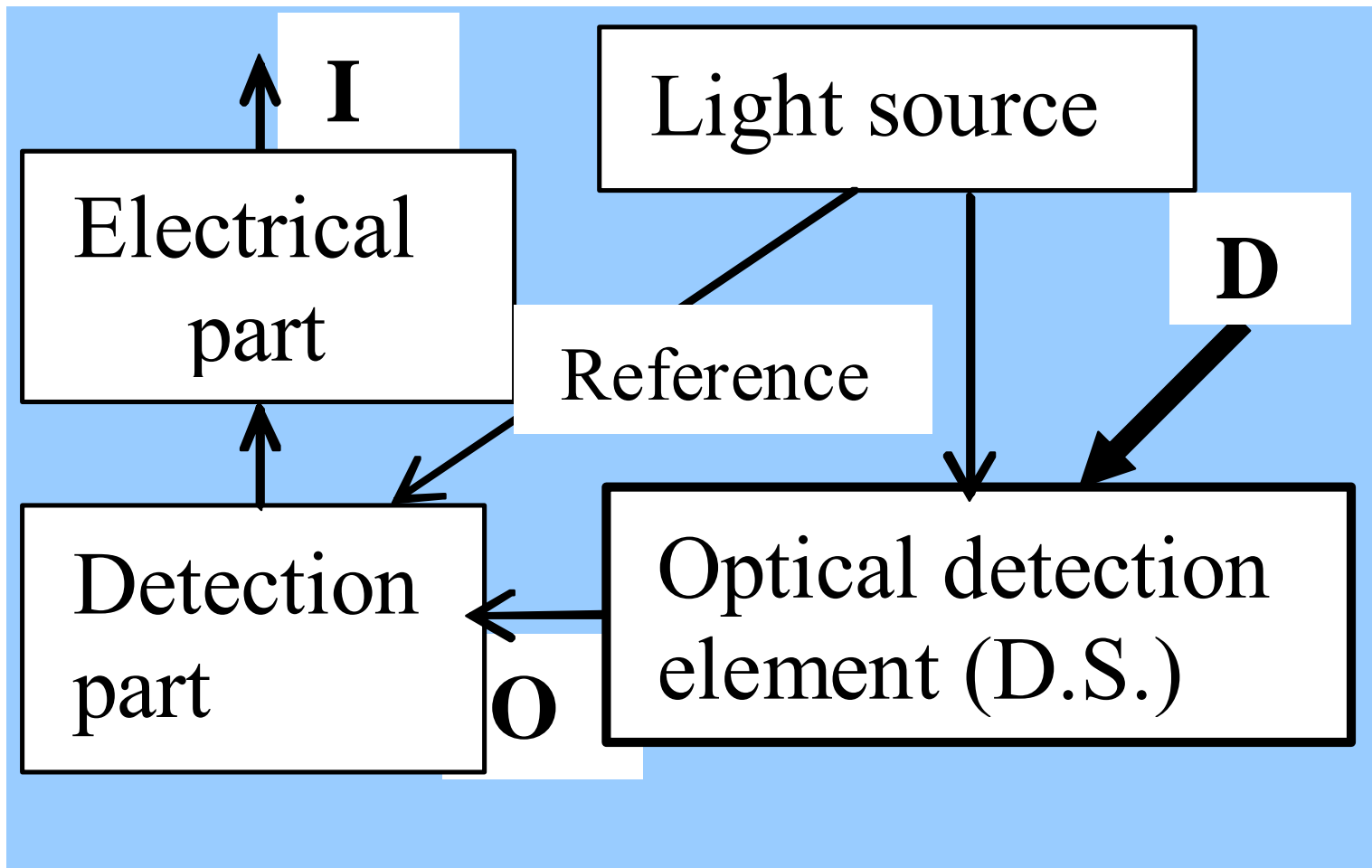
a small device for **continuous and reversible determination of qualitative and quantitative changes**

physical variables (temperature, pressure, etc.), chemical composition in its vicinity by **employing light changes**

(amplitude, phase, frequency, polarisation).



# SET-UP OF OPTICAL SENSOR



D.S. – Detection site

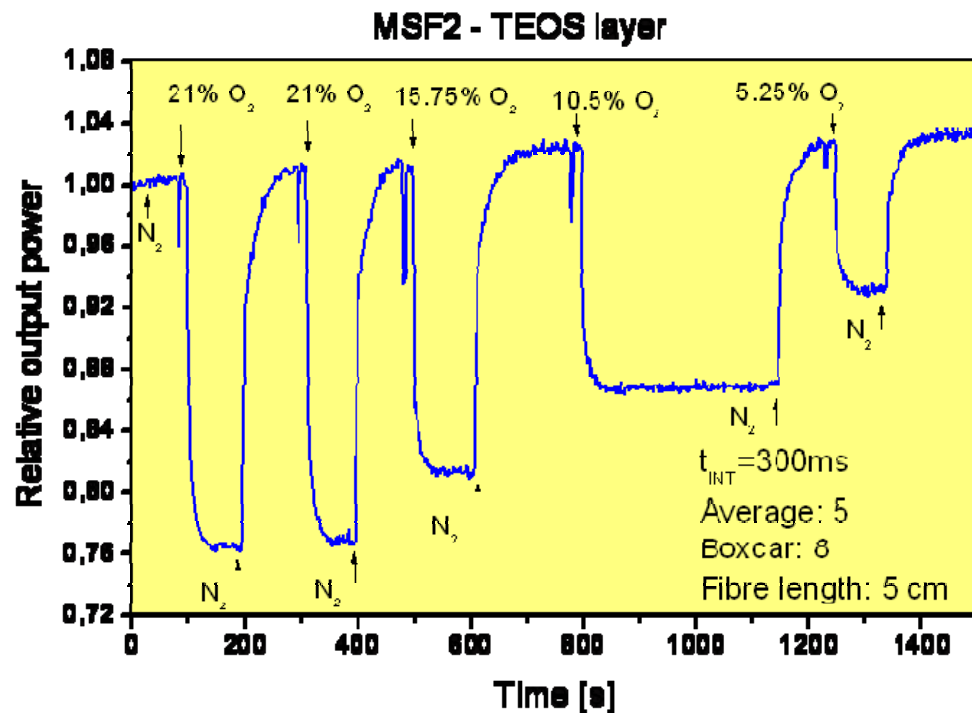
# OPTICAL SENSOR

## **Obligatory parts** of optical sensors

- Light source (halogen lamp, LED, LD)
- Detection part (waveguide, foil, prism, ...)
- Detector (photodiode, photon counter, spectrometer)
- Optical signal treatment

All these parts are integrated in one systems

# TIME-RESPONSE OF SENSOR



Oxygen detection by quenching  
fluorescence of Ru complex

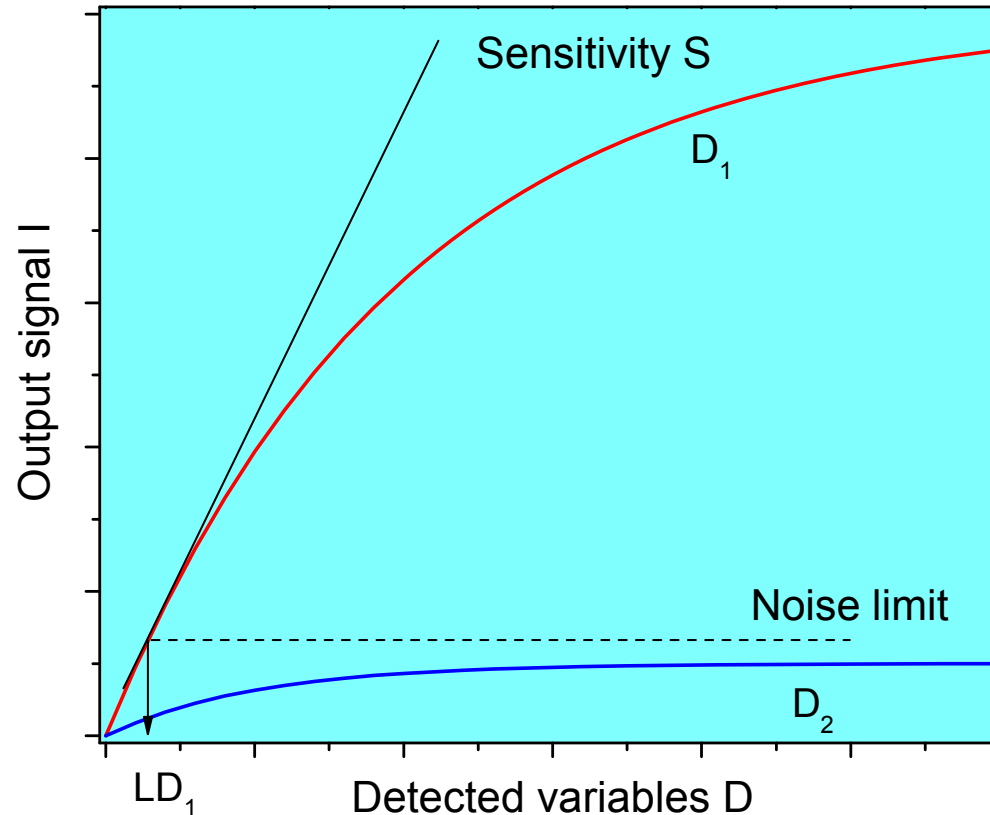
$$I = I_s \left( 1 - \exp\left(\frac{-t}{\tau}\right) \right)$$

$I_s$  saturated value-

$\tau$ - time constant (for  
 $I=0.63 I_s$ )

$\tau$  Range: s - min

# SENSOR SENSITIVITY- CALIBRATION CURVE



Variable  $D_1$  detected in an extent  $LD_1 \rightarrow$

Variable  $D_2$  can't be detected

# SENSITIVITY AND SELECTIVITY

$$S_i = \left( \frac{\partial O}{\partial D_i} \right) \left( \frac{\partial I}{\partial O} \right)$$

## Selectivity

S is high (for the detected variable D)

I < Noise limit or  $S \rightarrow 0$  (the other variables in the detection site)

# OPTICAL CHEMICAL SENSORS

- D: Qualitative and quantitative detection of chemical substances, i.e. their presence and concentrations in detection site
- In detection site light interact with analyte and its characteristics are changed
- O: Light wave characteristics (**amplitude**, frequency, polarisation) employed for detection of D

# WHAT VARIABLES ARE DETECTED?

Analyte comes to the detection site and change

## Refractive index

Gases –  $n \sim 1$

Water -  $n \sim 1,33$ ; Ethanol -  $n \sim 1,37$

Silicone polymers, fluorinated acrylates  $n \sim 1,4$

Silica glass –  $n \sim 1,46$

Toluene -  $n \sim 1,5$

PMMA -  $n \sim 1,49$ , PVC –  $n \sim 1,54-1,56$

Optical glass F2 –  $n \sim 1,51$

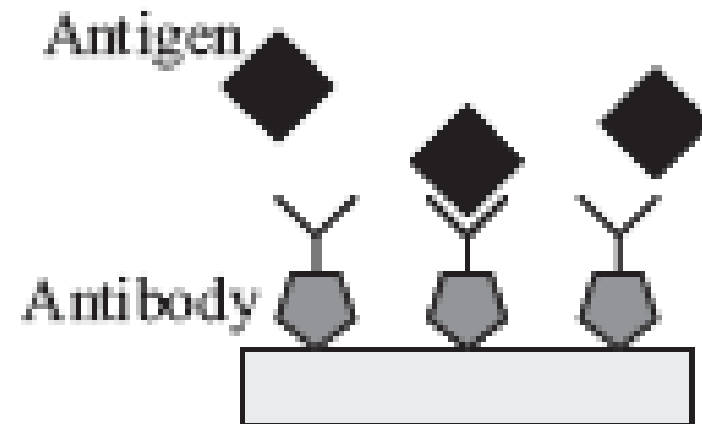
Histidine –  $n=1,7$

# REFRACTIVE-INDEX CHANGES

Refractive-index changes in the detection site are **nonspecific** ⇒ transducer is necessary that accept only detected substance (analyte)

„label free“ immunosensors  
interaction of antibody with  
antigene

Increased density in antibody  
chain →  $n$  increase





# ABSORPTION COEFFICIENT

Detected substance – analyte changes absorption coefficient

NIR spectral bands

(CH~1600-1700 nm, NH ~ 1500 nm, OH ~ 1400 nm  
overtones of fundamental IR frequencies 2900-3600 cm<sup>-1</sup> )

Modified Lambert-Beer law absorbance A

UV spectral bands (250 nm) of aromatic compounds

$$A = \gamma L \sum_{i=1}^N \varepsilon_i c_i$$

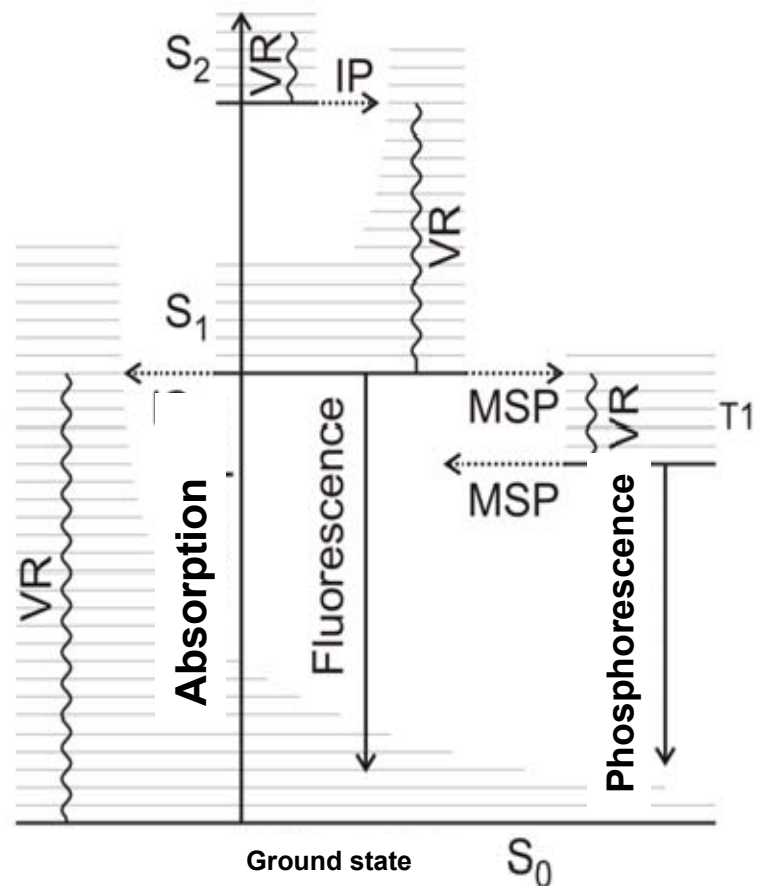
$\gamma < 0.01$  if standard fibers without any change are used

**Direct detection of cells with fiber tips or UV spectra of antibodies**

See e.g.:A. Leung et al. *Sensors and Actuators B* 125 (2007) 688–703

# LUMINESCENCE

Luminescence =  
Fluorescence or  
Phosphorescence



$\overset{IP}{\dashrightarrow}$	Internal transition	$S_i \dashrightarrow T_j$	Nonradiative transition
$\overset{MSP}{\dashrightarrow}$	Intersystem transition	$S_i \dashrightarrow S_j$	Radiative transition
$\overset{VR}{\sim}$	Vibration relaxation		

# LUMINESCENCE

**Fluorescence**: allowed energetical transition without spin change, rapid (lifetime  $\sim \mu\text{s} - \text{ns}$ )

**Phosphorescence**: spin change, not allowed, slow (lifetime  $\sim \text{s} - \text{ms}$ )

Luminescence effects are related to:

Aromatic rings, conjugated double bonds, Ru complexes

# LUMINESCENCE INTENSITY

$$P \approx P_0 \varepsilon(\lambda) Q c L$$

$\varepsilon(\lambda)$  – absorption coefficient  $\sim 10^5$  l/mol/cm Q – quantum efficiency, c – concentration of luminescent analyte (luminofor)

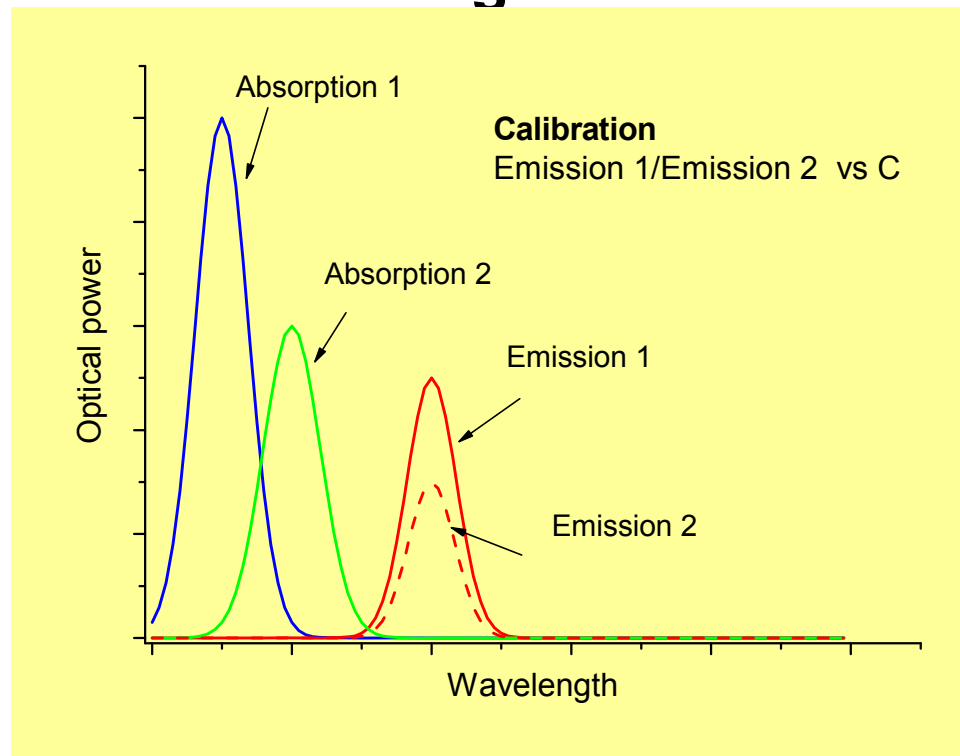
**Excitation way: electroluminescence, photoluminescence, bioluminescence, chemoluminescence**

See e.g.: C.L. Morgan et al. *Clinical Chemistry* 42, No. 2, 1996

# REFERENCE METHODS

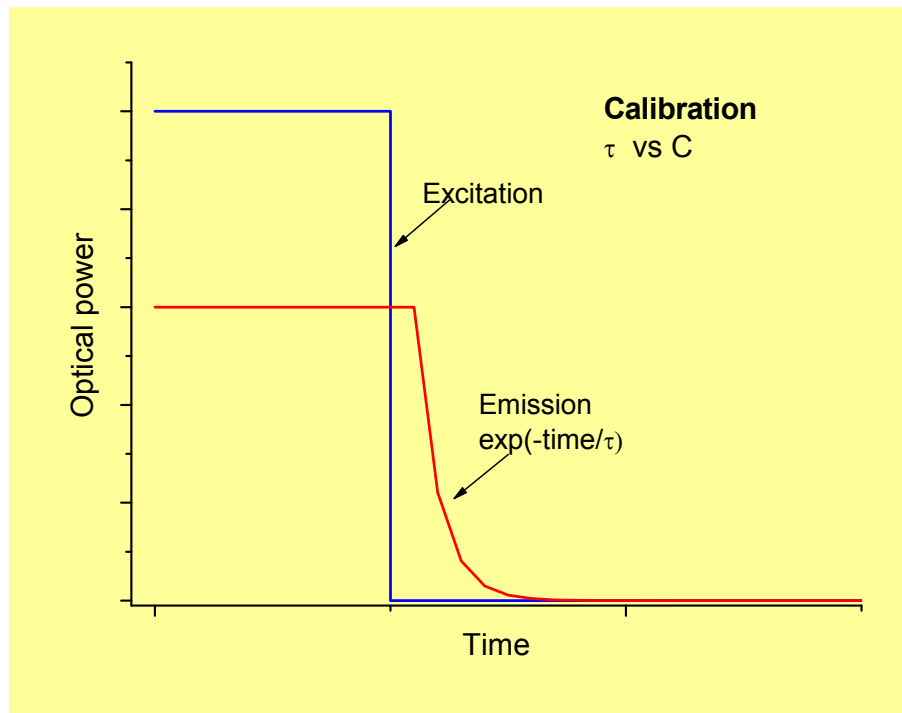
Luminescent intensity  $P$  suffers from random fluctuations → Reference methods:

1. Excitation at two absorption wavelength, detection at one detection wavelength



# LUMINESCENCE LIFETIME

2. Luminescence lifetime  $\tau$  is practically independent of random fluctuations. Some analytes (oxygen, carbon dioxide) induce fluorescence quenching.



**Stern-Volmer equation**

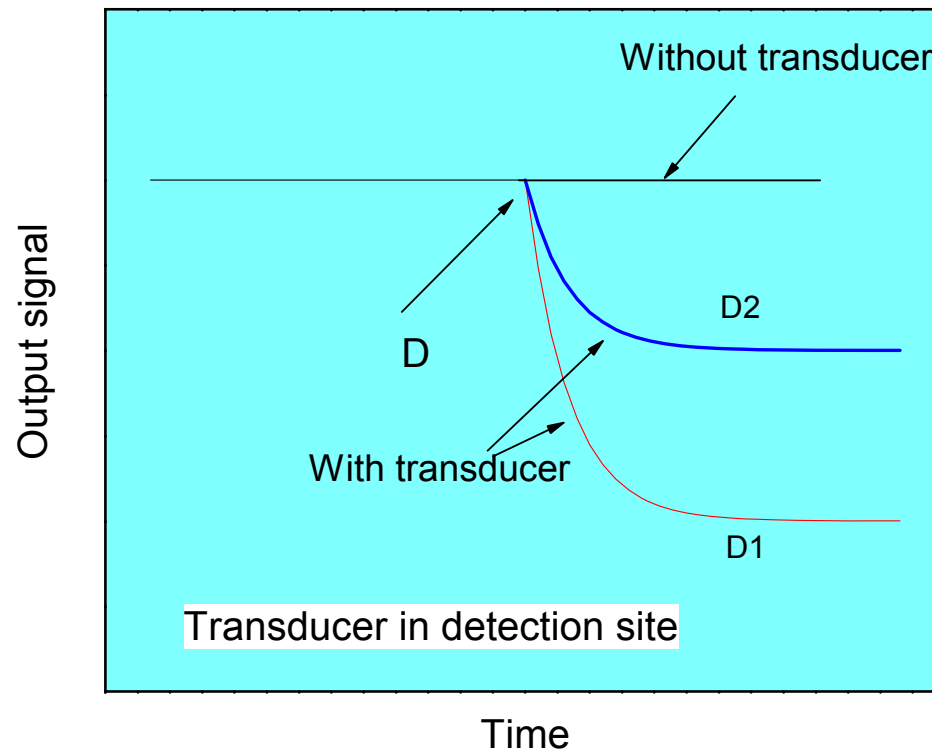
$$\frac{\tau_0}{\tau} = 1 + K_{SV} [Q]$$

**Q j- quencher concentration**

[http://www.jh-inst.cas.cz/~fluorescence/support/Lectures/UFCH\\_fluor04.pps](http://www.jh-inst.cas.cz/~fluorescence/support/Lectures/UFCH_fluor04.pps)

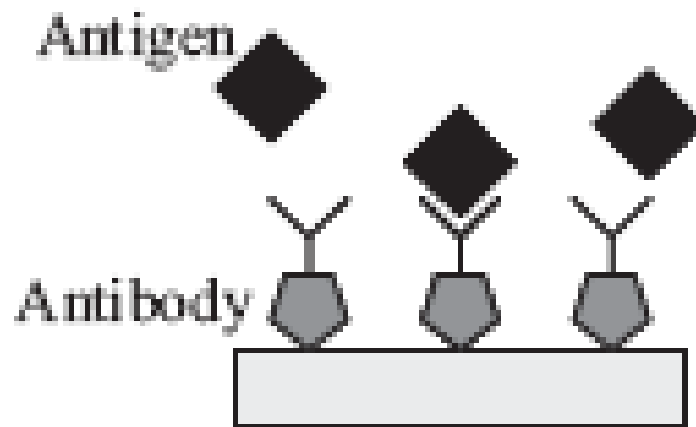
# OPTICAL TRANSDUCERS

If analyte has no optical absorption, luminescence, only it changes refractive index → **transducer** has to be added that changes its optical properties due to interaction with analyte



# REFRACTIVE INDEX - IMMUNOTRANSDUCERS

Based on immunoreaction (affinity reaction) at which antibody (protein) specifically capture analyte (antigen) → refractive-index change



**Gamaglobulins Ig**  
**5 types – G,M,A,E,D**

**Usually IgG**

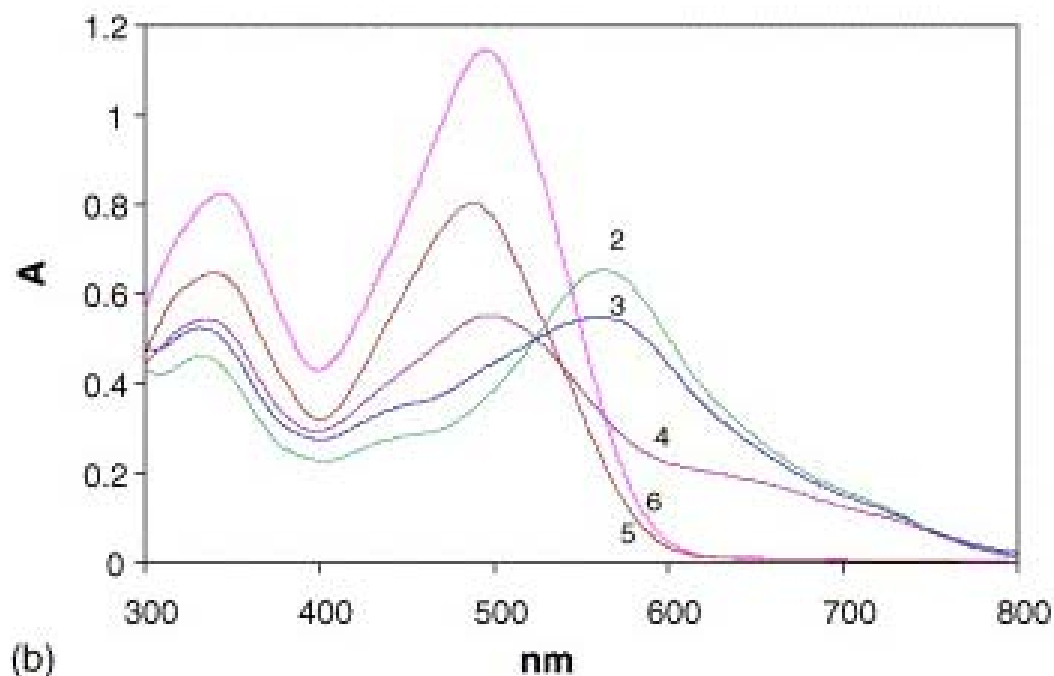
See e.g.: D.W.G. Morrison et al., „Clinical application of micro- and nanoscale biosensors“, Biomedical Nanostructures, Ed. K.E. Gonsalves, C.L. Laurencin, C.R. Halberstadt, L.S. Nair, 433-453, Ch. 17, 2008



# ABSORPTION TRANSDUCERS

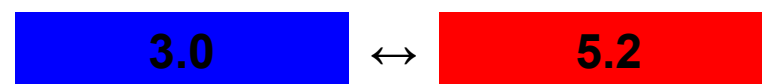
Absorption coefficient

pH indicators:  $\text{BInd} + \text{H}_3\text{O}^+ \rightarrow \text{AInd} + \text{H}_2\text{O}$



*below pH 3.0*

*above pH 5.2*



**Absorption spectra of „Congo red“ for different pH**

P. Hashemi et al., *Sens. Act. B115*, 49-53 (2006)

# ENZYMATIC TRANSDUCERS

Enzymes = biocatalysts

Enzymatic reaction

*Substrate (analyte)  $\xrightarrow{\text{enzym}}$  Product*

Analytes = glucose, pesticides, urine

**Glucose + oxygen  $\xrightarrow{\text{glucoseoxydase}}$  Gluconic acid + hydrogen peroxide**

Product can be detected by luminescence or absorption spectroscopy or other transducer is added (Ru complex for detection of oxygen, pH indicator for detection of gluconic acid)

# DETECTION OF ORGANOPHOSPHOROUS COMPOUNDS

## Enzymatic transducer + pH transducer

Cholinesterase + methyl red

**Acetylcholine + water → cholin + acetic acid**

Reaction takes part at nerve end and enables registration of external changes. Organophosphorous compounds stop the reaction of acetylcholine (inhibition).

Detection of nervous paralytic welfares (Sarin, Soman).

Inhibition of cholinesterase ⇒ no pH change.



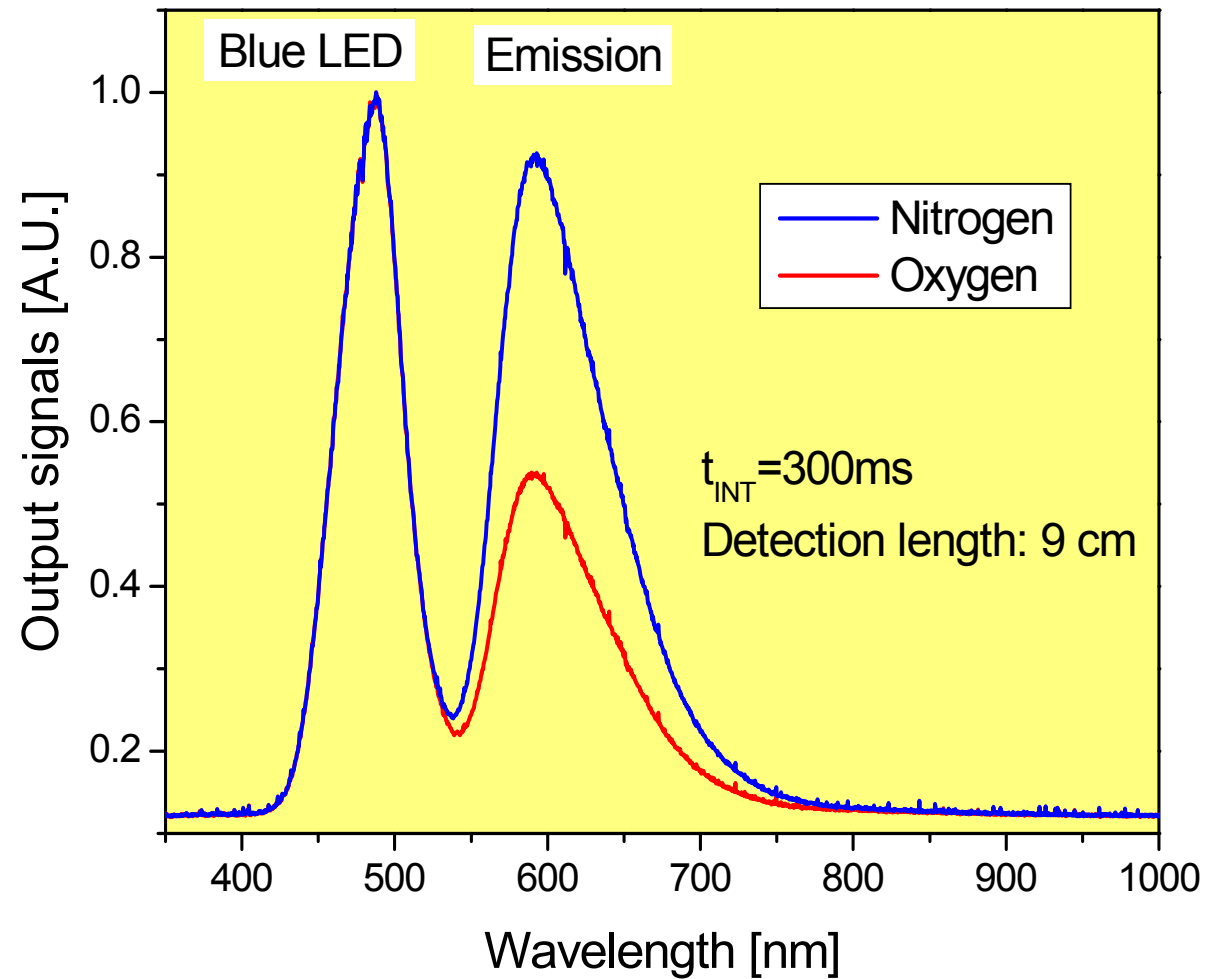
Army detector CHP71

# LUMINESCENT TRANSDUCERS

- Fluorescence marks on IgG, enzymes, DNA chains
- Ru complexes with fluorescence quenched by oxygen
- Fluorescence pH indicators – BCECF, HPTS used in biosensor.

# DETECTION OF OXYGEN

Oxygen  $\leftrightarrow$  Ru(phen)<sub>2</sub>Cl<sub>2</sub>  $\Rightarrow$  fluorescence quenching



# DETECTION MEMBRANES

## Function

1. *Control the refractive index in the detection site*
2. *Increase concentration of analyte in the detection site (partition coefficient  $K_p$  )*

$$K_p = \frac{C_{site}}{C_{external}}$$

## DETECTION MEMBRANES

$K_p \rightarrow \infty$  (for analyte)

$K_p \rightarrow 0$  (other substances in sample)

3. *Immobilize optical transducers in the detection site*

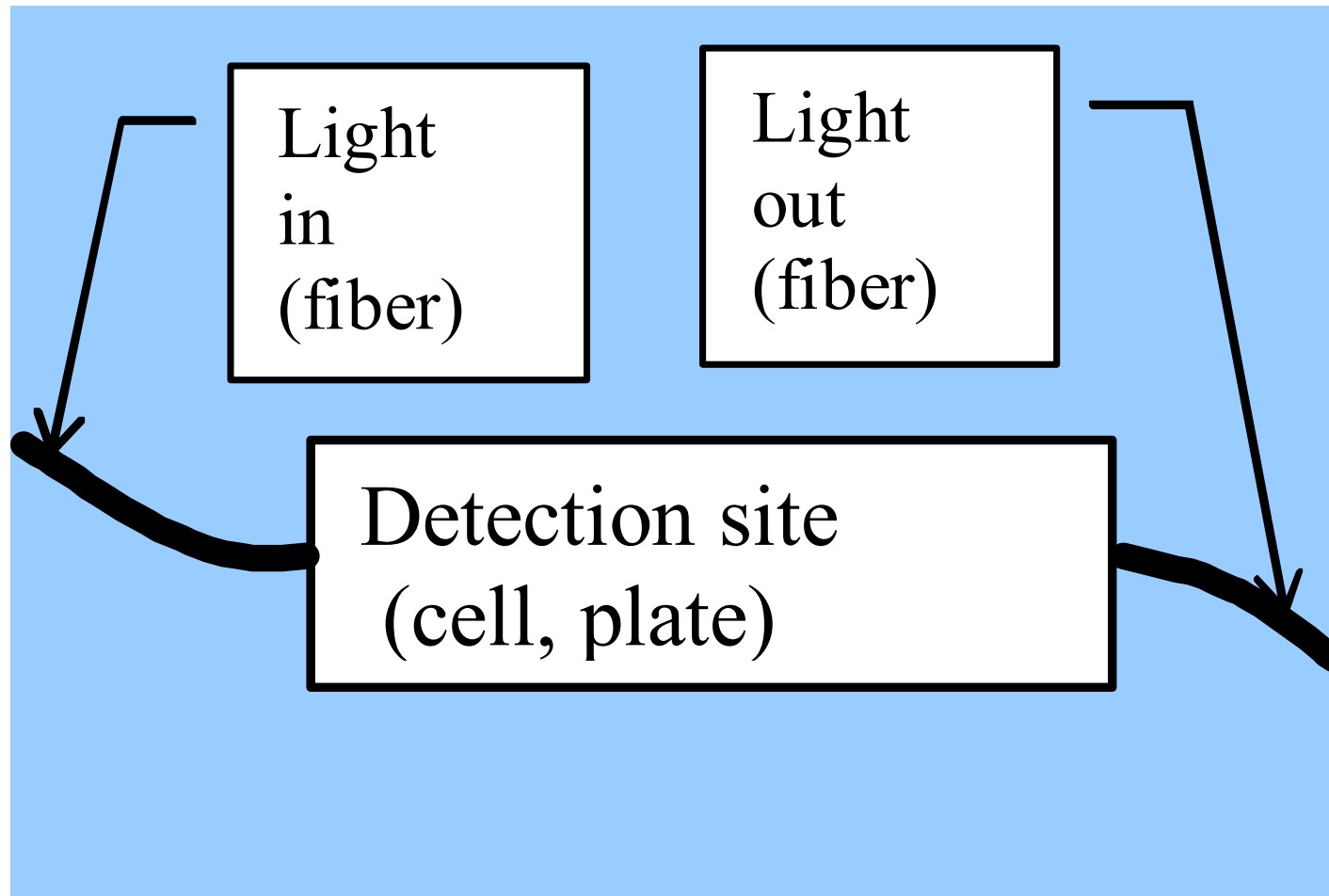
# OPTICAL FIBERS IN

## **FIBER-OPTIC CHEMICAL SENSORS (FOCS)**

- They are used for transmission of light to and from detection site (extrinsic sensors)
- Detection sites can be created in their cladding (intrinsic sensors)
- Detection membranes and optical transducers are applied onto fibers, creating the detection site

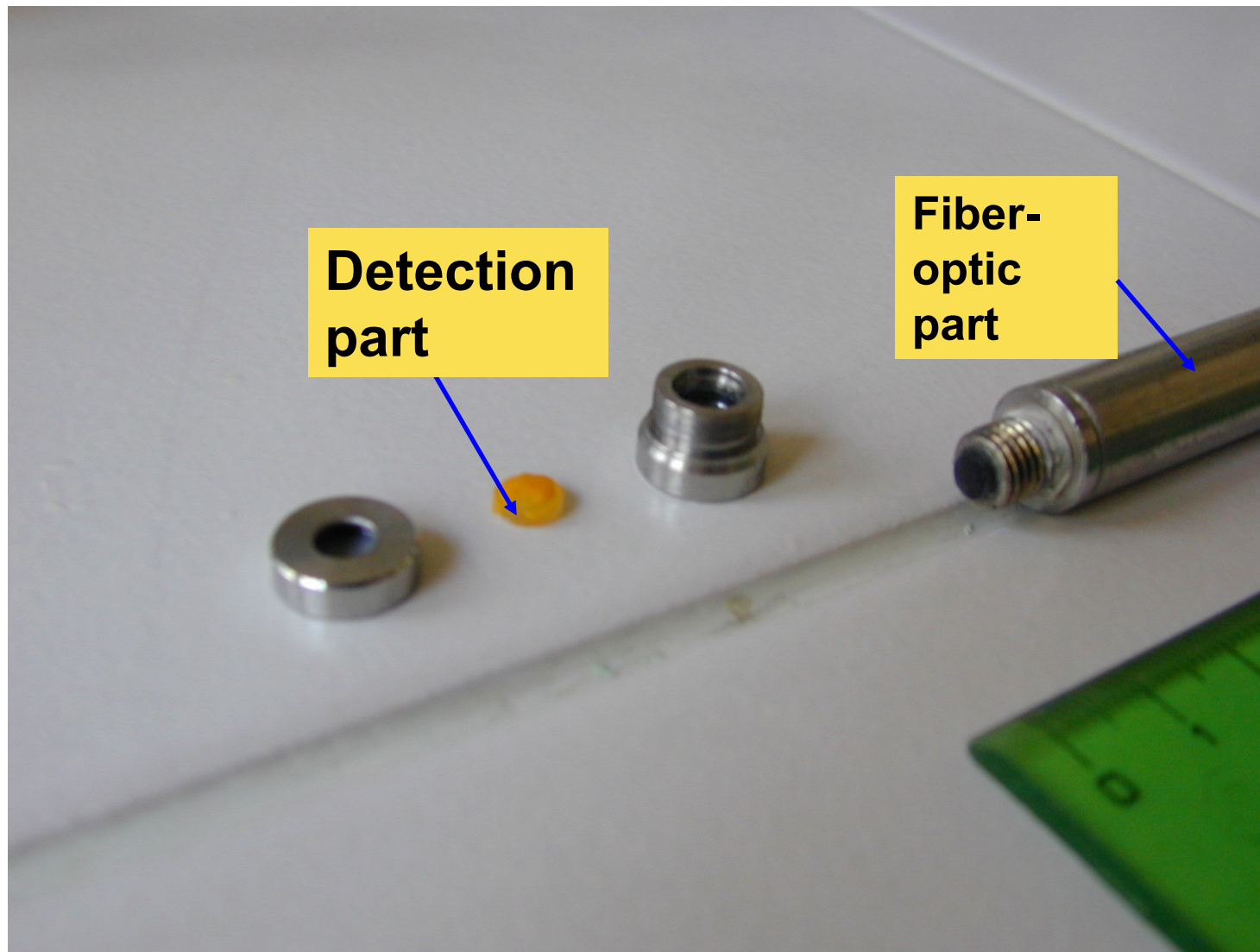


# EXTRINSIC FOCS

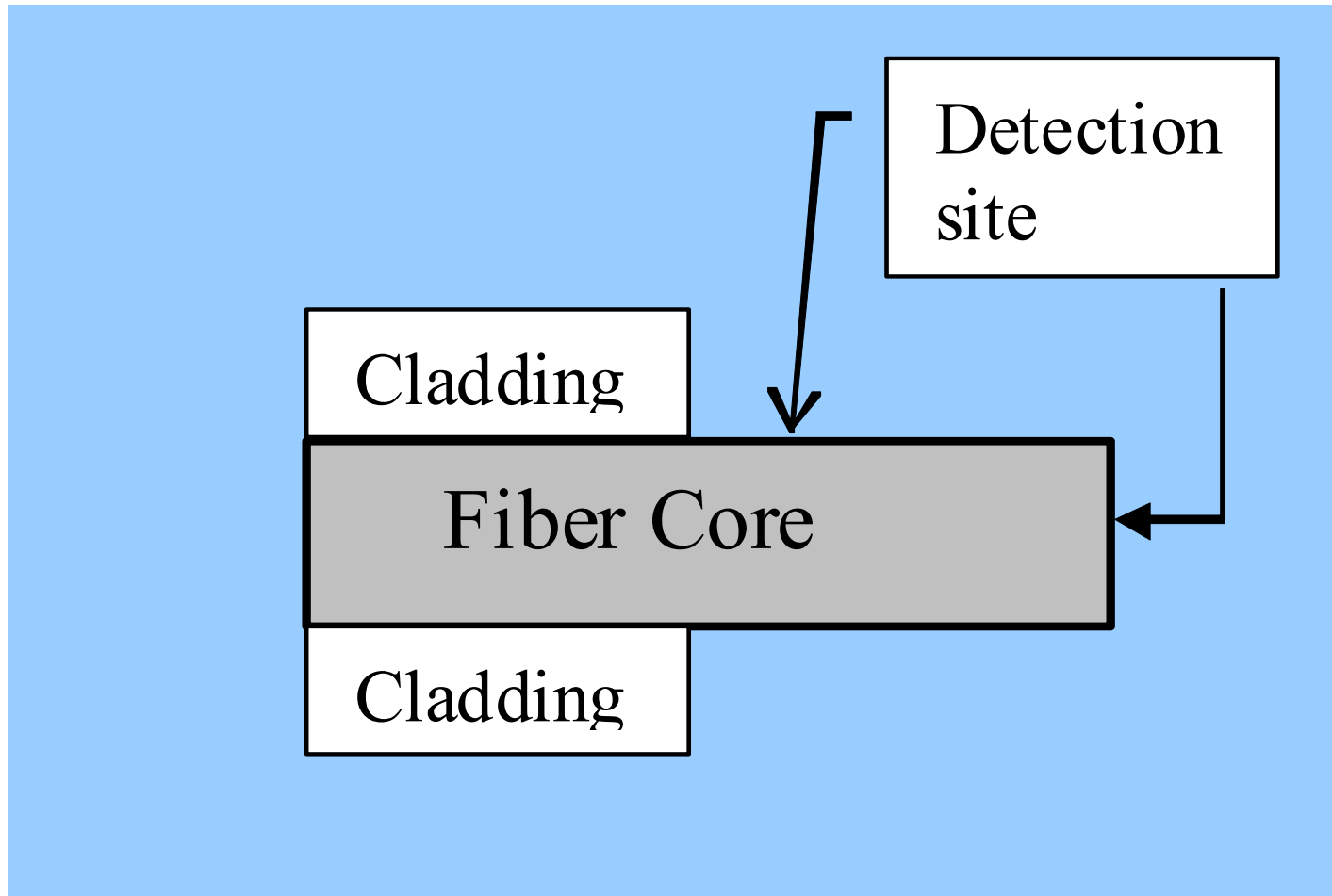


Detection site is outside the fiber (cell, plates)

# IPE - EXTRINSIC FOCUS OF OXYGEN

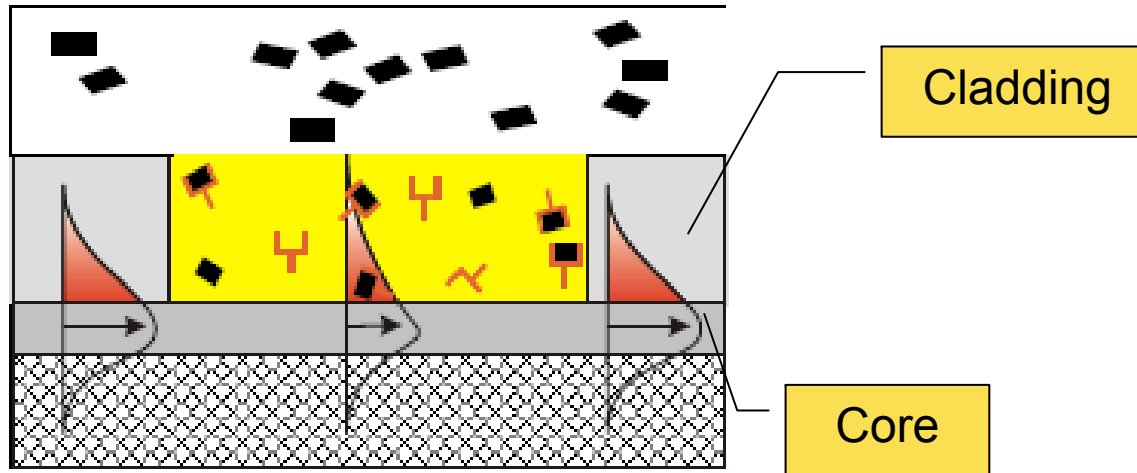


# INTRINSIC FOCS



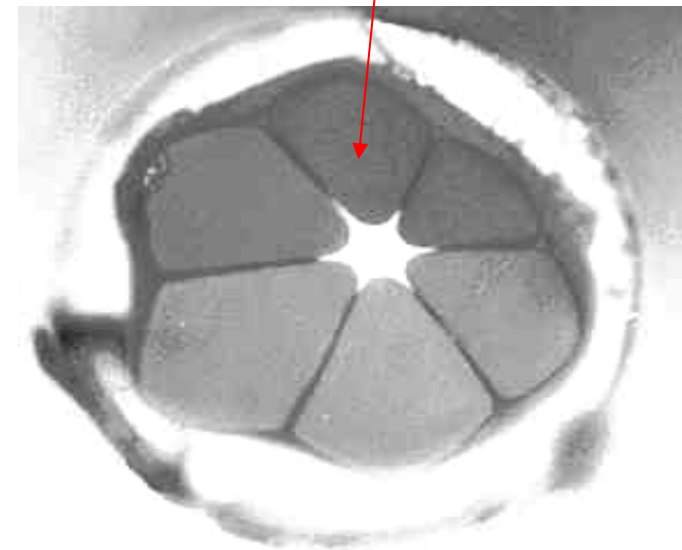
Detection site is in the fiber structure (cladding, core)

# INTRINSIC FOCS



Detected substance  
(gas or solution) in  
direct contact with  
guided modes

**Detection site in cladding  
holes**



80 μm  
↔

## ADVANTAGES OF FOCS

- Small dimensions, long detection lengths
- Distributed and remoted detection

## Disadvantages of FOCS

- Special instrumentation of chemical parts
- Competition with other sensor types (electrical, mechanical)
- Parazitic signals, fawling of detection sites

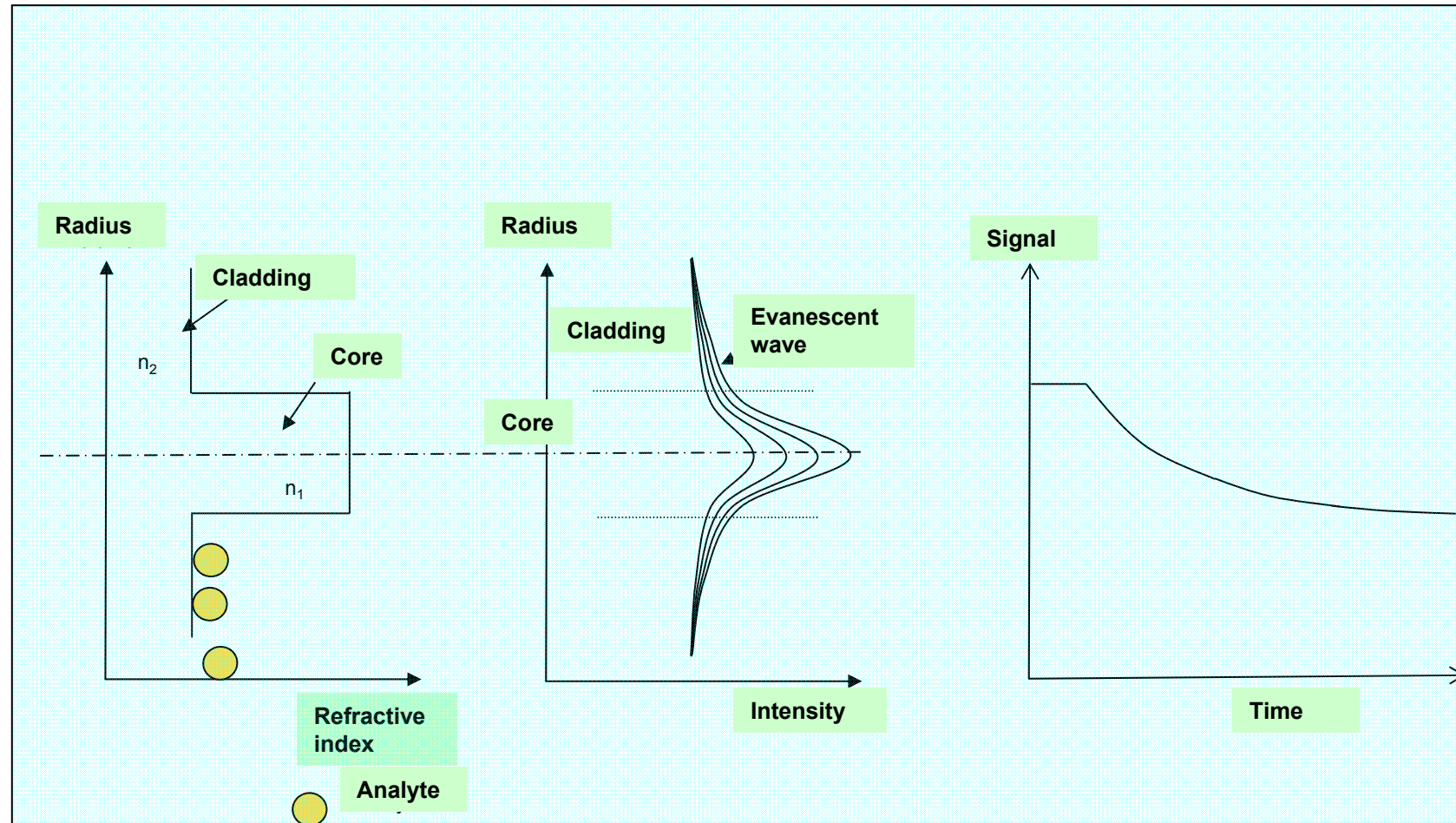
## POTENTIAL APPLICATIONS OF FOCS

- **in remote and dangerous spaces** (petrol tanks, refineries, workshops with flammable and explosive gases, depots of radioactive substances)
- **In medicine** for continuous monitoring of concentrations of gases, pH, in blood or in organs such as stomach or brain.

# DETECTION STRUCTURES OF FOCS

- **Evanescent FOCS**
- **Reflection FOCS**
- **Surface plasmon (SPR) FOCS**
- FOCS based on optical gratings (LPG)
- FOCS based on fiber tips, tapers

# EVANESCENT-WAVE SENSOR PRINCIPLE



Analyte penetrate into the cladding, changes optical properties there → changes in transmission of evanescent wave and signal



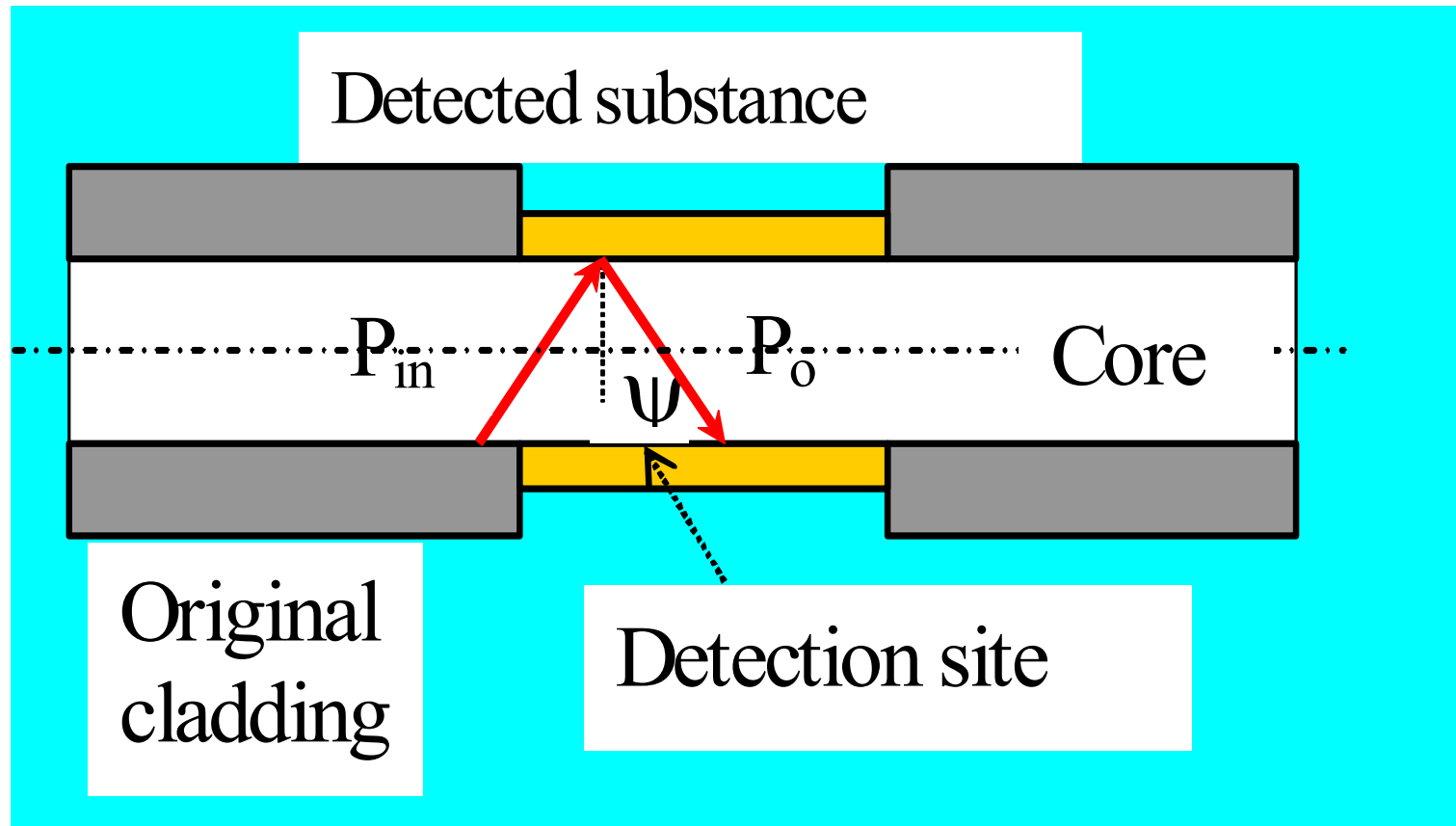
# PRINCIPLE OF EVANESCENT FOCS

**Analyte changes transmission of evanescent wave in the cladding (a decrease of its amplitude) due to:**

- **Refractive index – refraction sensors**
- **Optical absorption - attenuated total reflection sensors**
- **Luminescence – luminescence sensors**

**Consequently amplitude of part of optical mode propagating in the core (evanescent wave is a part of optical mode) changes which is detected as change of the output signal.**

# RAY-OPTIC MODEL OF EVANESCENT-WAVE SENSOR



$$P_o = P_{in} R$$

$R$  – power reflection coefficient  $R \leq 1$

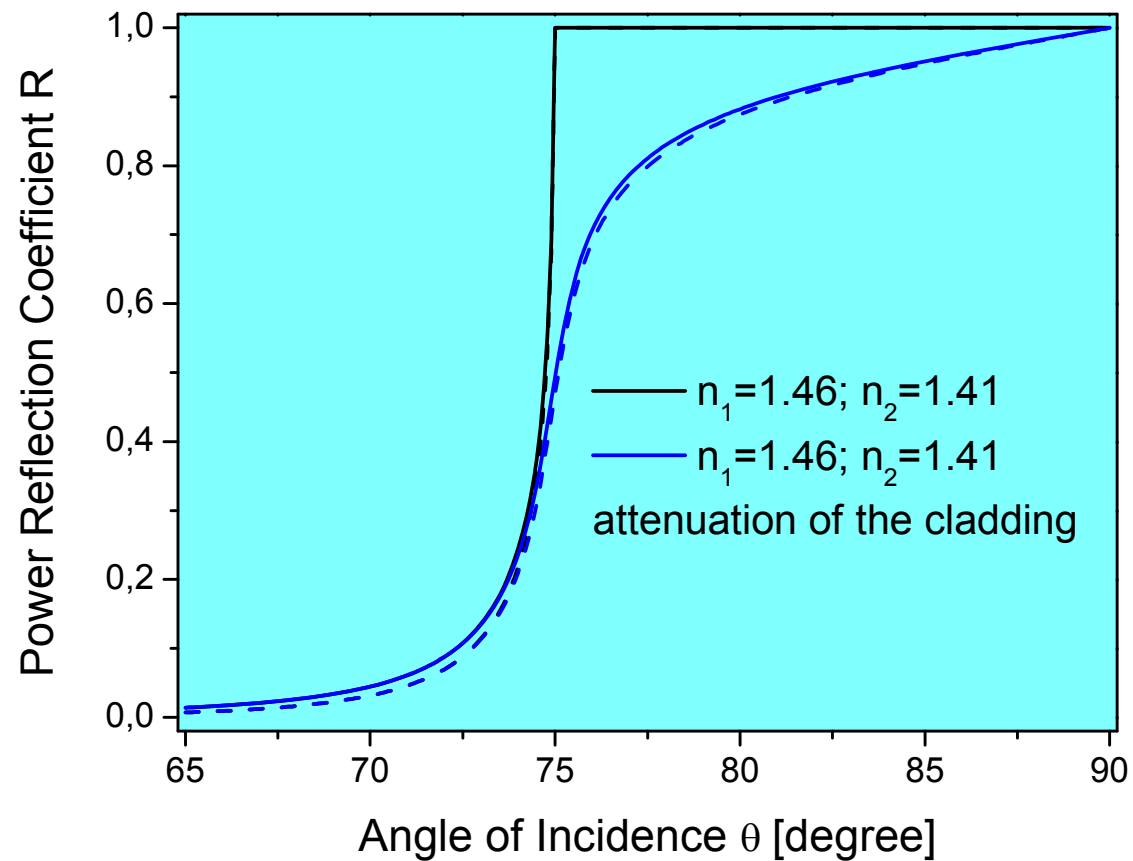
# OPTICAL POWER TRANSMITTED BY RAY

$$P_i = P_{0i} R^{N_i}$$

$N_i \sim L/d$  – number of reflections

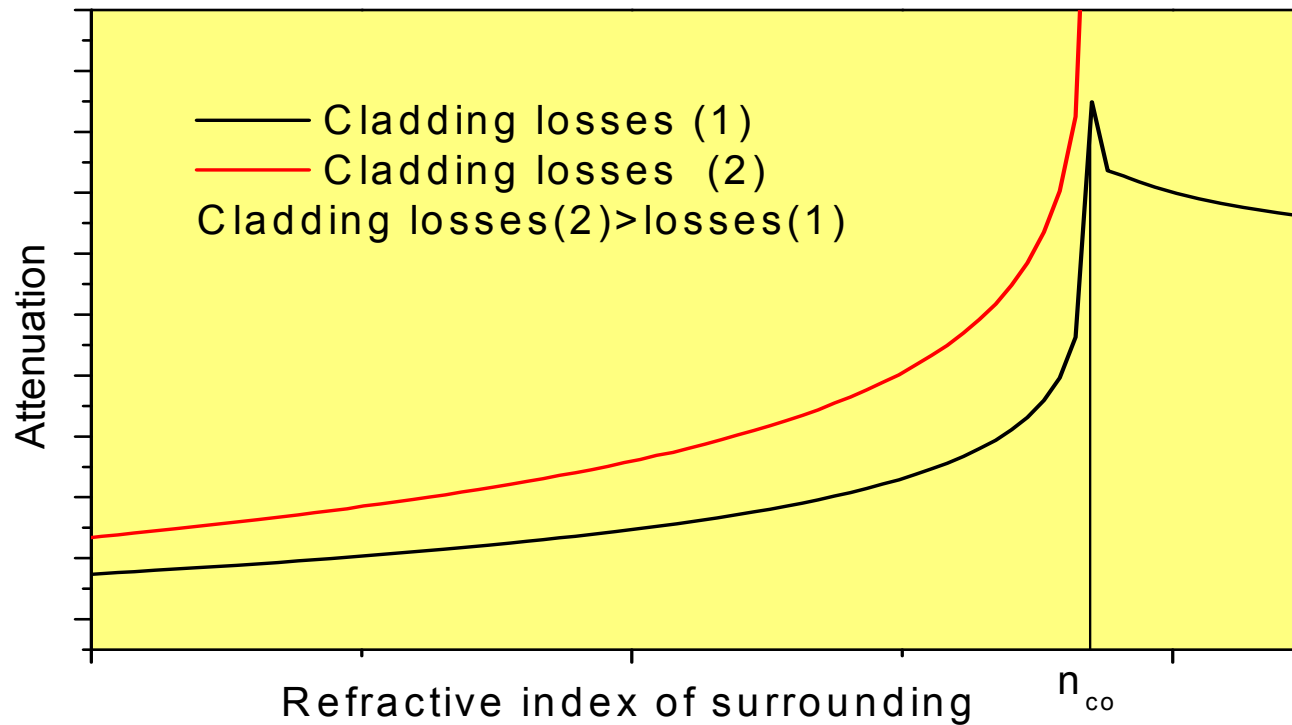
$P_{0i}$  – optical power launched into  $i$ -th ray  
on the input face of the fiber

# REFLECTION COEFFICIENT- REFLECTIVITY R



$\theta=90-\Psi$  – axial angle complementary to reflection angle

# TRANSMITTED POWER ↔ RI AND CLADDING LOSS



Attenuation =  $10 \log(\text{Input power}/\text{Transmitted power})$

Attenuation  $\uparrow \leftrightarrow$  Transmitted power  $\downarrow$

# TRANSMITTED POWER ↔ ATR SENSORS

## Meridional rays – Approximate formula

$$P_i = P_{0i} \exp(-\gamma L) \left( \frac{\theta}{\theta_c} \right)^2$$
$$\gamma \approx \varepsilon_{cl} \frac{\lambda}{\pi d \sqrt{n_{co}^2 - n_{cl.}^2}} \frac{1}{\sqrt{1 - \left( \frac{\theta}{\theta_c} \right)^2}}$$

$\theta = \pi/2 - \Psi$  – complementary angle to the angle of reflection  $\Psi$  (**ATR – Attenuated Total Reflection**)

# LIMITS OF EVANESCENT-WAVE SENSORS

## Sensitivity

~1% of total power transmitted in the cladding of standard optical fibers → low sensitivity to optical changes in cladding.

$$\gamma < 0.01$$

## Detection depth

Intensity of evanescent wave exponentially decreases from the core/cladding boundary

Penetration depth  $d_p$  – place in the cladding where intensity = 1/e intensity on the boundary

$$d_p \sim 0.1 - 1 \mu\text{m}$$

$$d_p = \frac{\lambda}{2\pi \sqrt{n_1^2 \sin^2 \Psi - n_2^2}}$$

# WAYS FOR SENSITIVITY INCREASE

Higher sensitivity  $\leftrightarrow$  Lower value of transmitted power  $P_i$

1. Increase of detection length  $L$
2. Increase of bulk absorption coefficient  $\varepsilon$
3. Decrease of core diameter  $d$
4. Approaching  $n_{cl} \rightarrow n_{co}$
5. Approaching  $\theta \rightarrow \theta_c$

**These approaches are used for multimode fibers**



# EVANESCENT-WAVE SENSORS ON MULTIMODE FIBERS

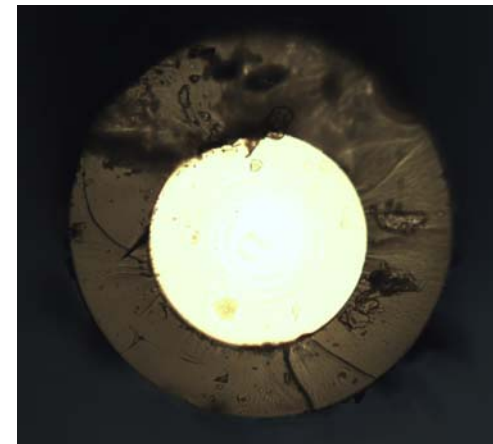
- **Polymer-Clad Silica (PCS) fibers**

Low prices, good availability, robustness, simple modification, losses 0.01 dB/m

Core: silica  $n=1.46$

$d \sim 0.2-1$  mm,

Cladding polydimethylsiloxane (PDMS),  
fluorinated acrylate ( $n=1.41$ )



# EVANESCENT-WAVE SENSORS ON MULTIMODE FIBERS

- **Polymer-Clad Glass (PCG) fibers** with expensive, commercially hardly available, losses  $\sim$  dB/m

core: optical glass ( $n \sim 1.5-1.6$ )

$d \sim 0.4 - 0.6$  mm

cladding: PDMS, UV acrylate

# EVANESCENT-WAVE SENSORS ON MULTIMODE FIBERS

- **Polymeric fibers**

cheap, commercially available, flexible, higher optical losses  $\sim$  dB/m

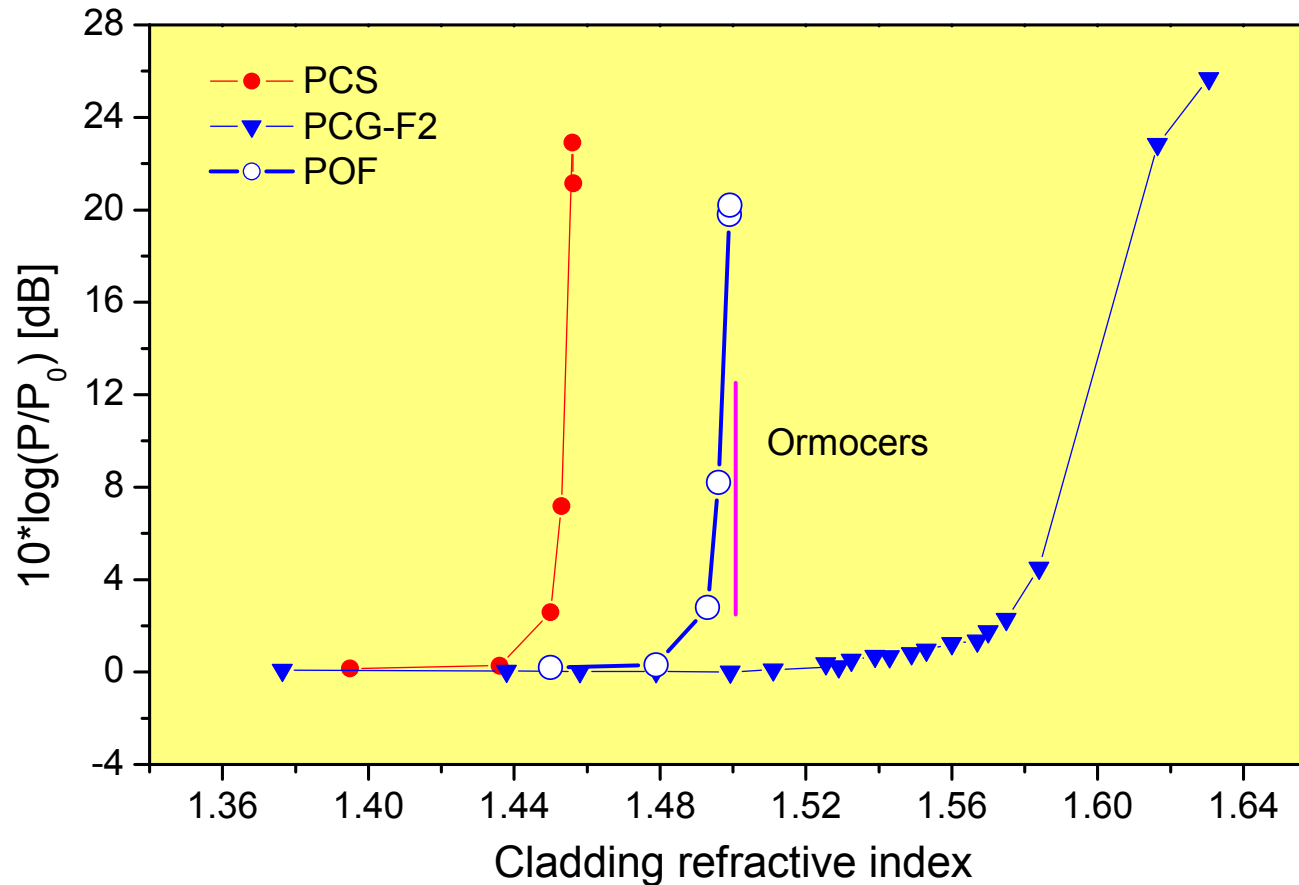
core: polymethylmetacrylate (PMMA)  $n \sim 1.49$   
 $d \sim 1$  mm

cladding: less dense PMMA or other polymers

In evanescent-wave sensors cladding is usually removed on a length of several cm  $\rightarrow$  making thus core/cladding boundary accessible for analytes

# SENSITIVITY INCREASE: $n_{cl} \rightarrow n_{co}$

## Calibration curve

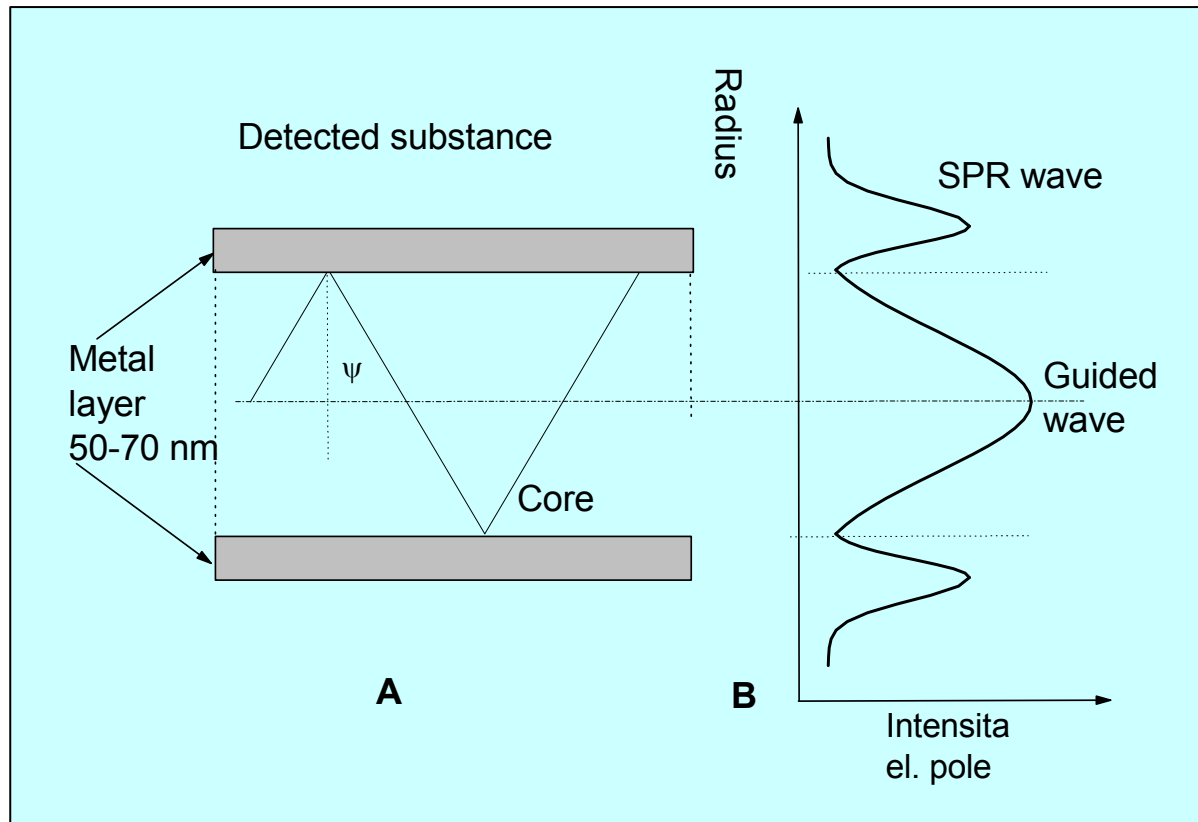


Sensitivity is high only for analyte refractive indices approaching to  $n_{co}$

### SURFACE PLASMON RESONANCE (SPR) SENSORS

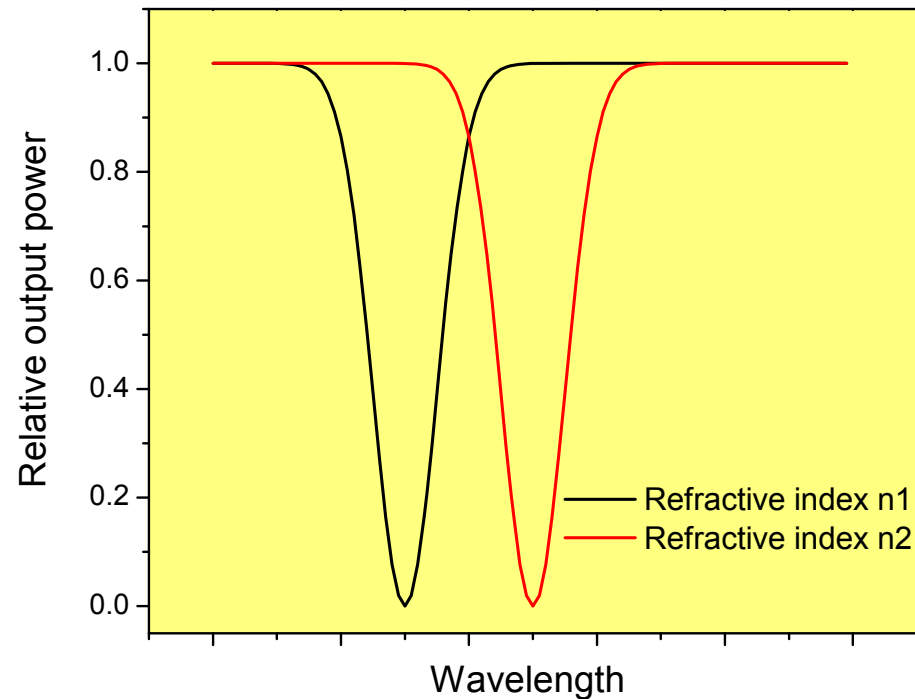
- a thin layer Au or Ag (30-50 nm) applied onto the core/cladding boundary
- evanescent wave excites free electrons in the layer to a higher energy level
- at proper wavelength, propagation constant, refractive index of surrounding energy from the wave is transferred to electrons → output power decreases (surface plasmon resonance - SPR)

# SCHEME OF SPR FOCs



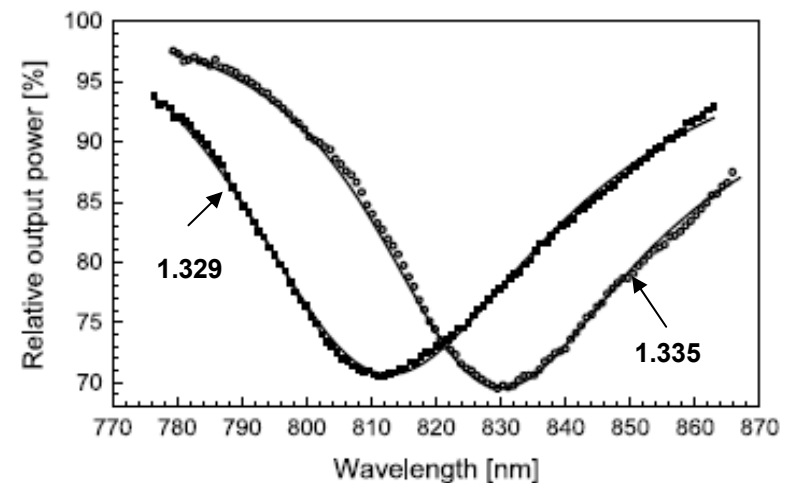
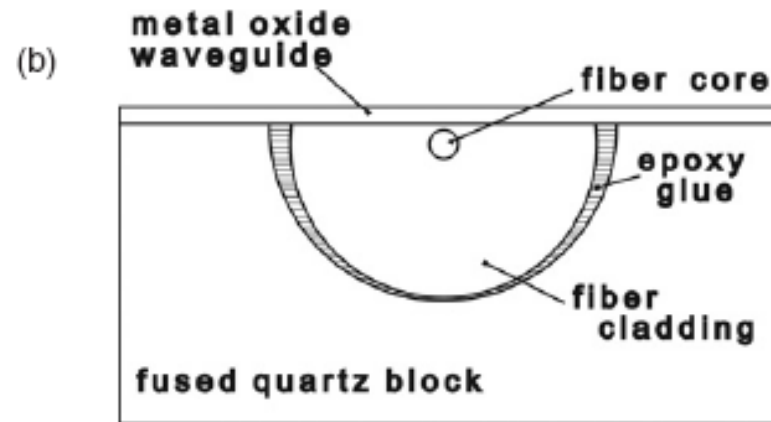
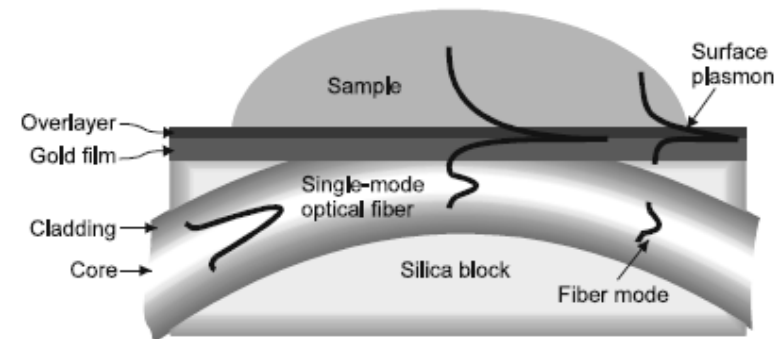
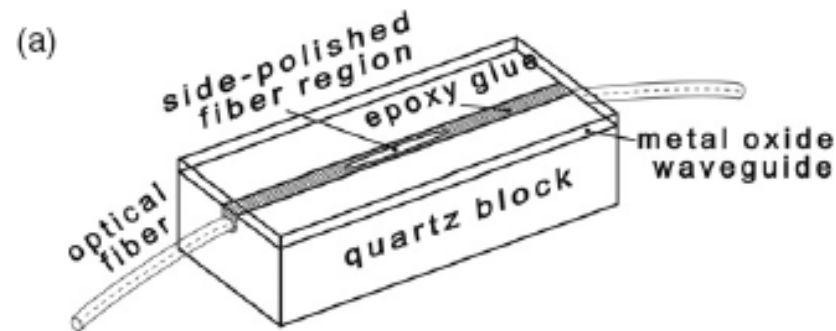
SPR wave penetrate into analyte on the layer boundary (100nm) and can be used for detection of analyte refractive index

# SP RESONANCE



SP resonance= dip observed at the output power spectrum, position depends on wavelength and analyte refractive index. Can be used also in aqueous solutions even with silica fibers.

# IPE SPR FOCSS – SIDE POLISHED FIBER

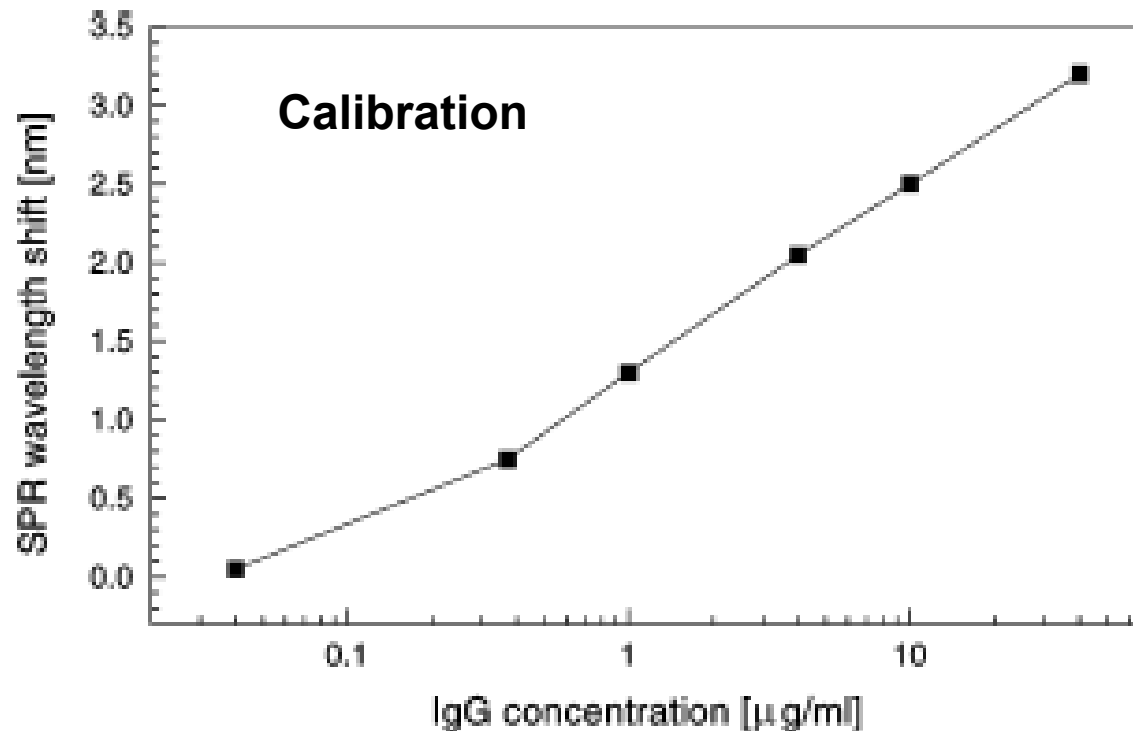


R. Slavik et al., Novel spectral fiber optic sensor based on surface plasmon resonance, *Sens. Actuators B74*, 106-111 (2001)



# SPR FIBER-OPTIC BIOSENSOR

SM fiber side-polished, Au layer 30 nm;  
immunotransducer antigen to IgG fixed on the fiber ,  
detection of IgG in a buffer solution (PBS) a BSA.



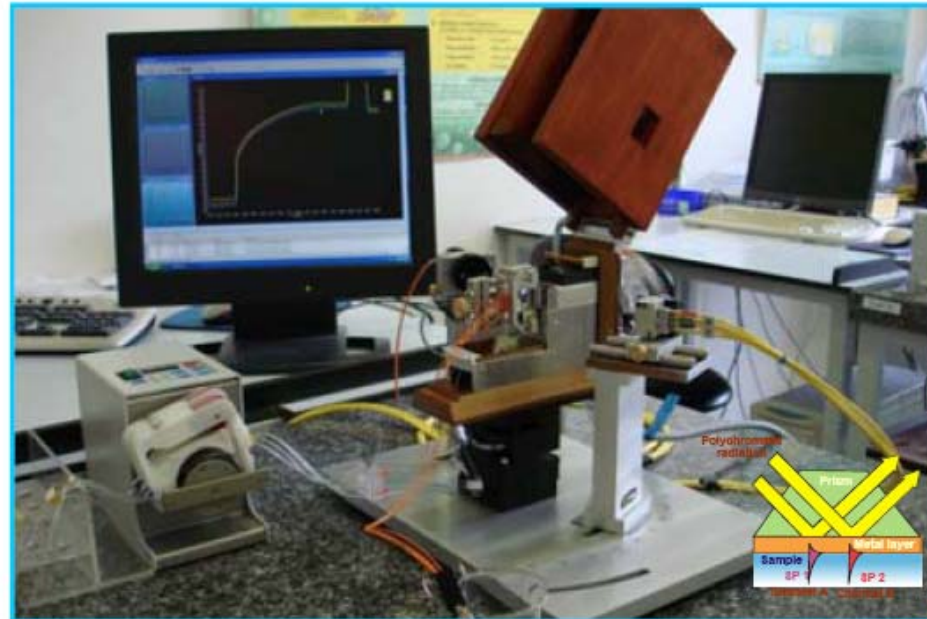
**PBS – Phosphate Buffered Saline (pH=7,4), BSA-Bovine Serum Albumine**

# IPE LABORATORY OF SPR SENSORS

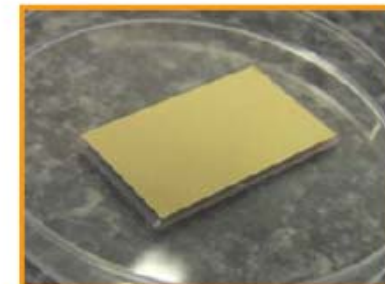
Employs usually optical prism for SP excitation (details [www.ufe.cz](http://www.ufe.cz))

- Spectroscopy of surface plasmons.
- Four sensing channels, (flow chamber volume 0.5  $\mu\text{L}$  per channel)
- Temp. stabilization (stability  $< 0.02^\circ\text{C}$ )

**RI RESOLUTION:**  
 $< 2 \times 10^{-7}$  RIU  
**OPERATING RANGE:**  
1.32-1.45 RIU

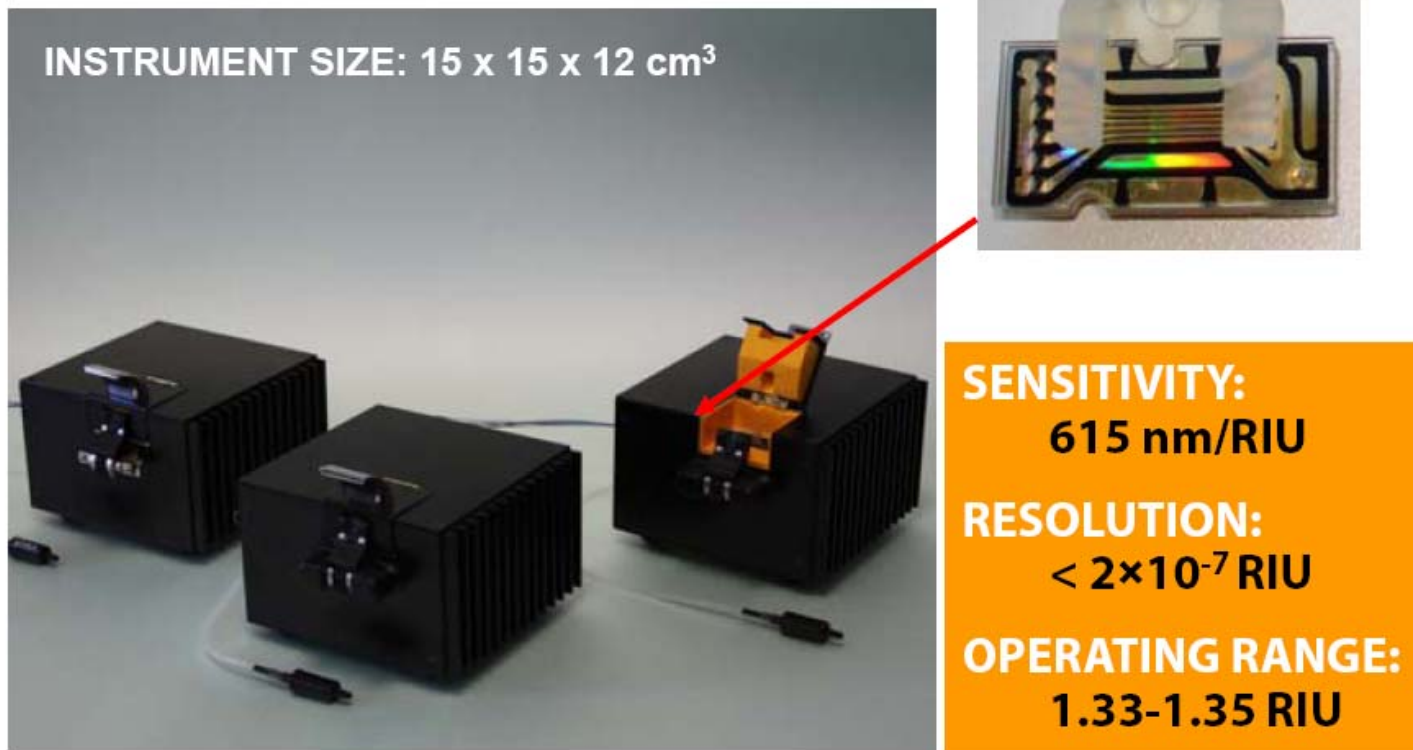


Four-channel SPR sensor and (top) and detail of an SPR chip (right).



# IPE PROTOTYPE OF SPR SENSOR

Based on Au layers excited by optical grating

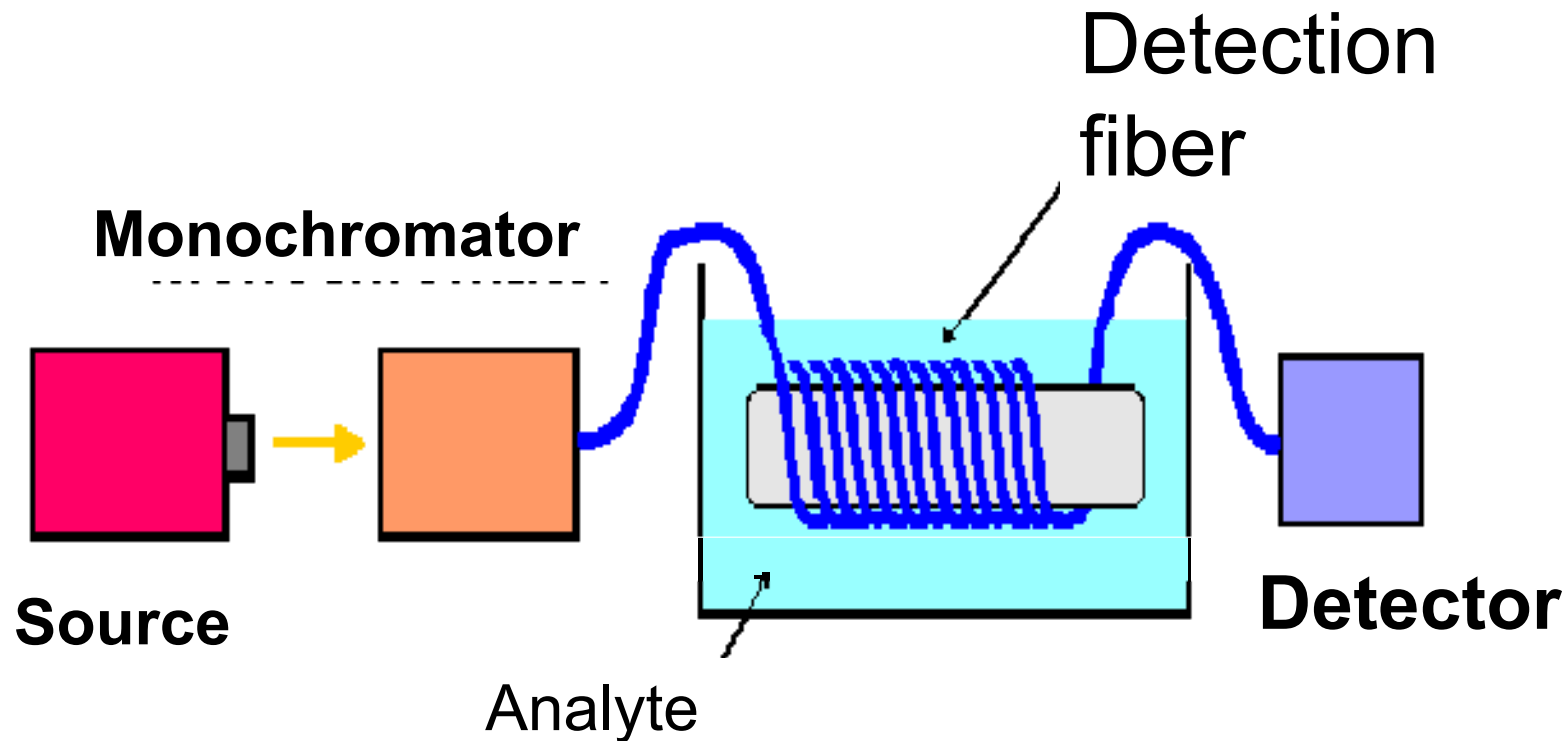


Laboratory prototype of 6-channel SPRCD sensor.

Used in > 5 countries, Hungary, Turkey, USA Federal Drug Agency

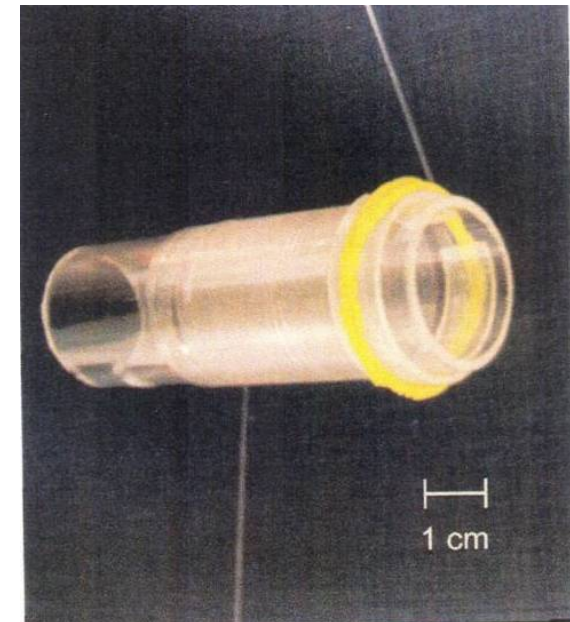
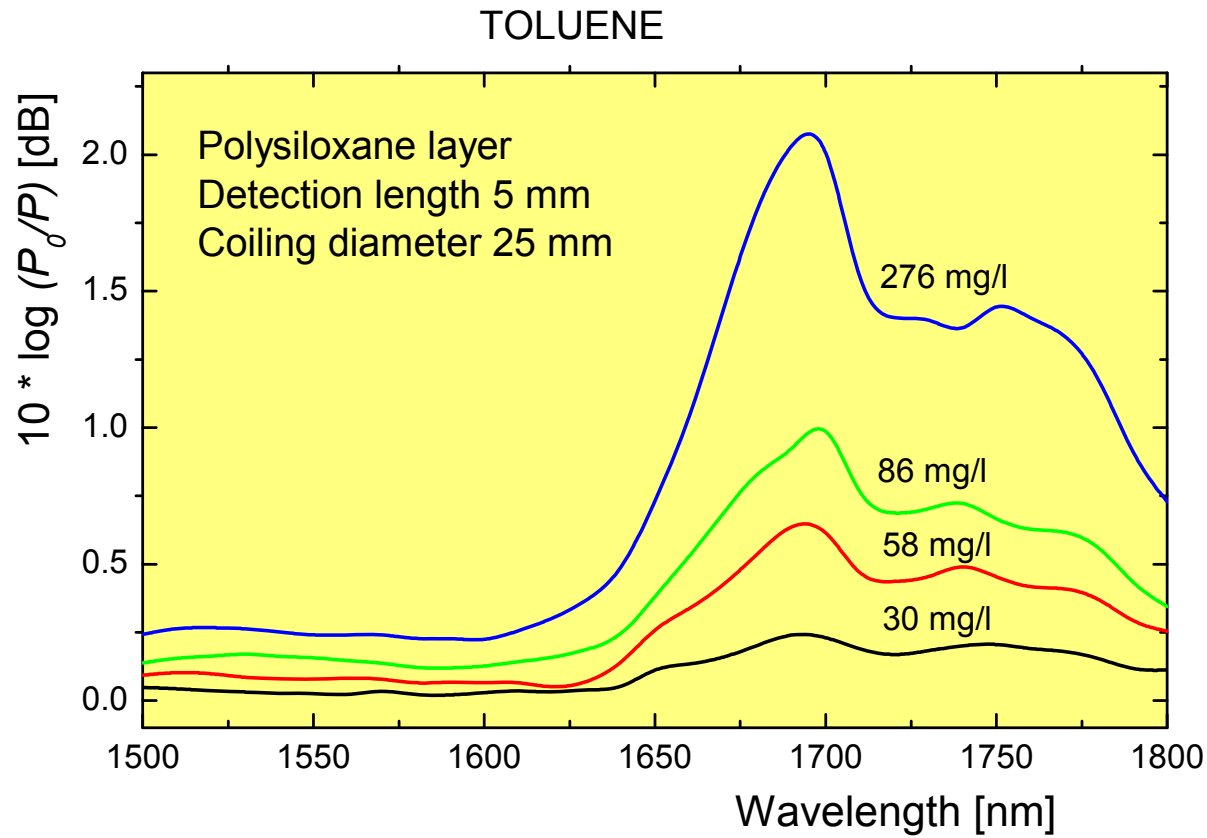
M. Pilarik, M.Vala, I. Tichý, J. Homola, *Biosens. Bioelectr.* 24, 3430–3435 (2009).

# SENSITIVITY INCREASE: $L \uparrow$



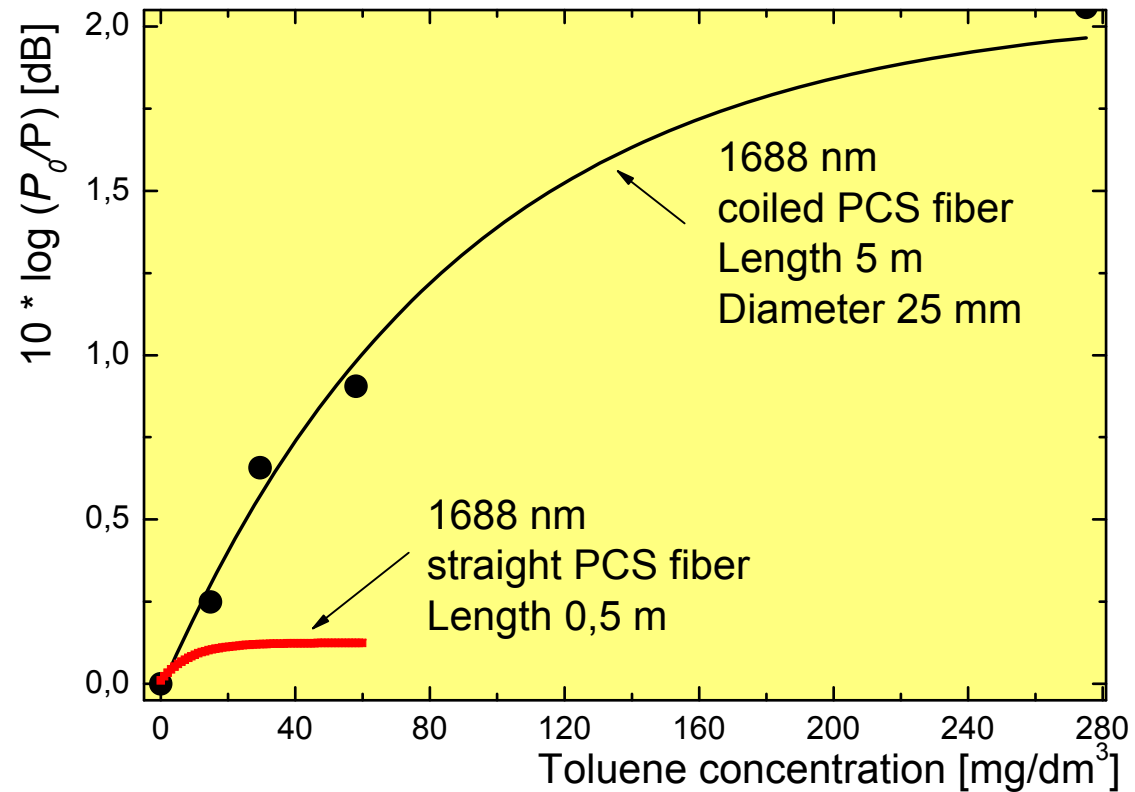
Up to 10 m of PCS fiber (core diameter 0.2 mm) coated with hydrophobic siloxane polymer coiled onto a glass rod (a diameter of 25 mm)

# SPECTRAL DETECTION OF TOLUENE IN WATER



PCS fiber 0.2 mm,  
cladding PDMS

# CALIBRATION CURVES



Effects:  $L$ ,  $n_{cl}$ ,  $\uparrow$  and  $\theta \downarrow$

## SENSITIVITY INCREASE: $d \downarrow$

Single-mode and GI fibers can't be directly used for detection because the core is surrounded by glass (thickness 30-50 $\mu\text{m}$ )

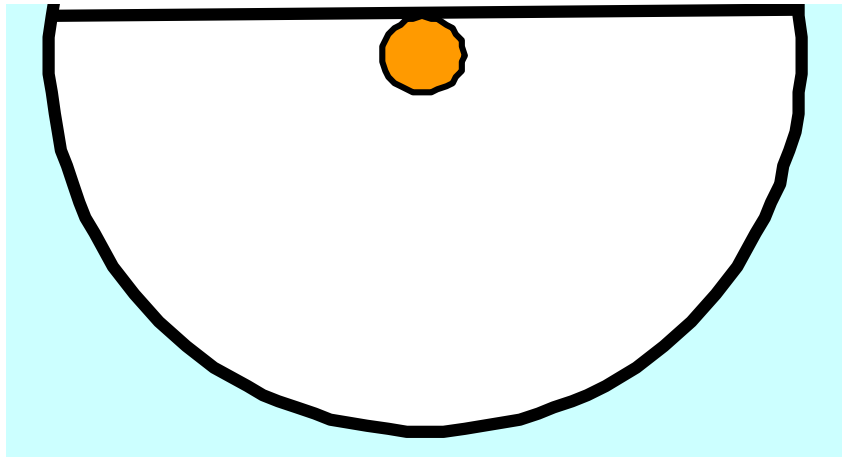


Outer glass tube and cladding have to be removed to open core/cladding boundary for analytes

D-shaped fibers or s fibers developed

# D-FIBERS

## D-fibers (B. Culshaw-UK-1985)



D-fiber prepared from a  
D-polished preform

Or

by polishing SM fibers

Applied for CH<sub>4</sub>, detection, SPR biosensors

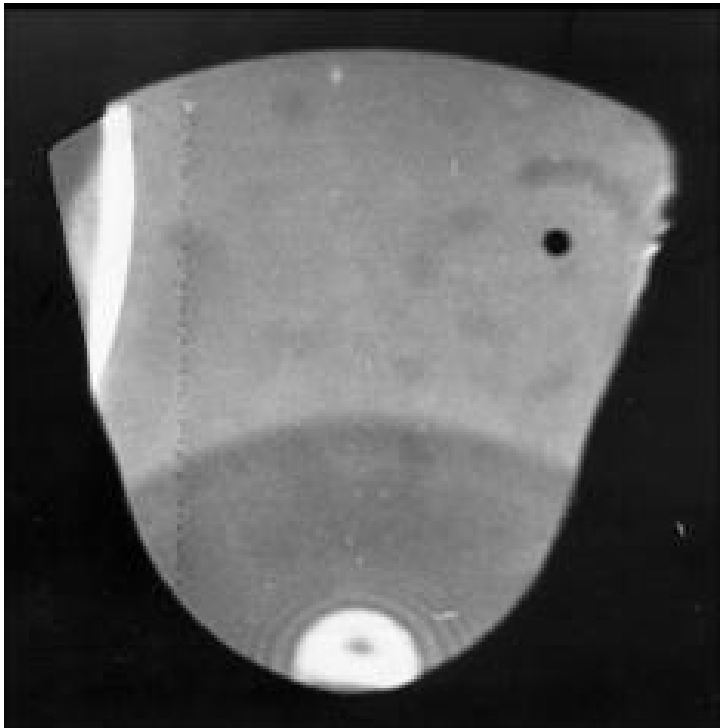
G. Stewart, W. Jin, B. Culshaw, *Sens. Act. B* 38, 42-47, 1997

R. Slavik et al., *Sens. Actuators B* 74, 106-111 (2001)



# IPE SECTORIAL (s) FIBERS

Similar to D fiber with easy access of analyte to the core/surrounding boundary; enables decrease of core diameter  $d_c$  .



S fiber is robust  $\leftrightarrow$  large sectorial part (0.3 mm)

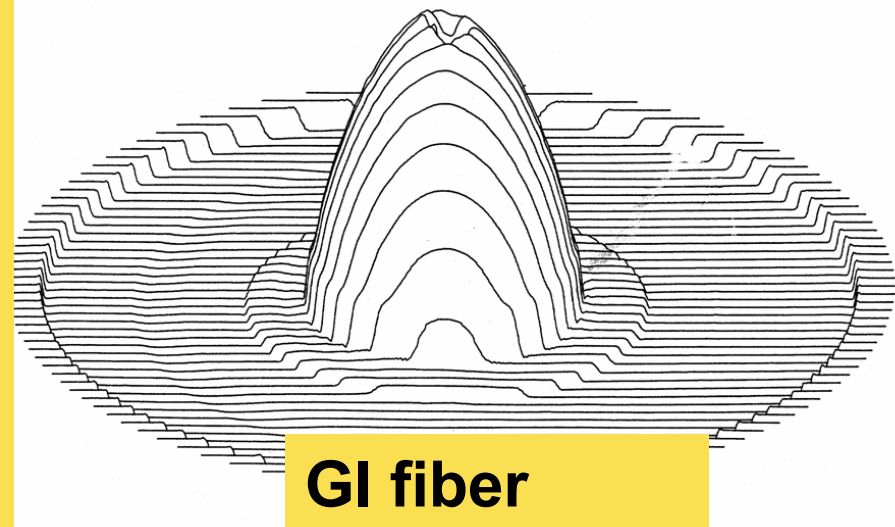
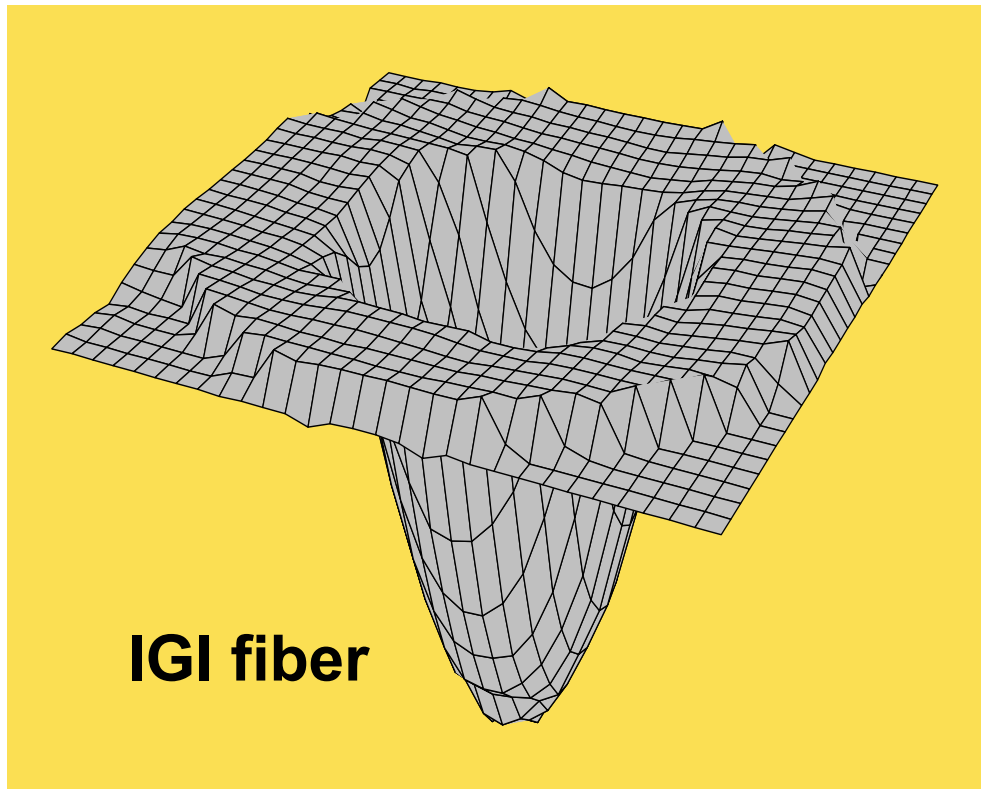
Core diameter -  $30\mu\text{m}$  (2-5x increase of detection sensitivity compared to PCS fiber  $d=0.3\text{ mm}$ )

S fiber tested for pH detection

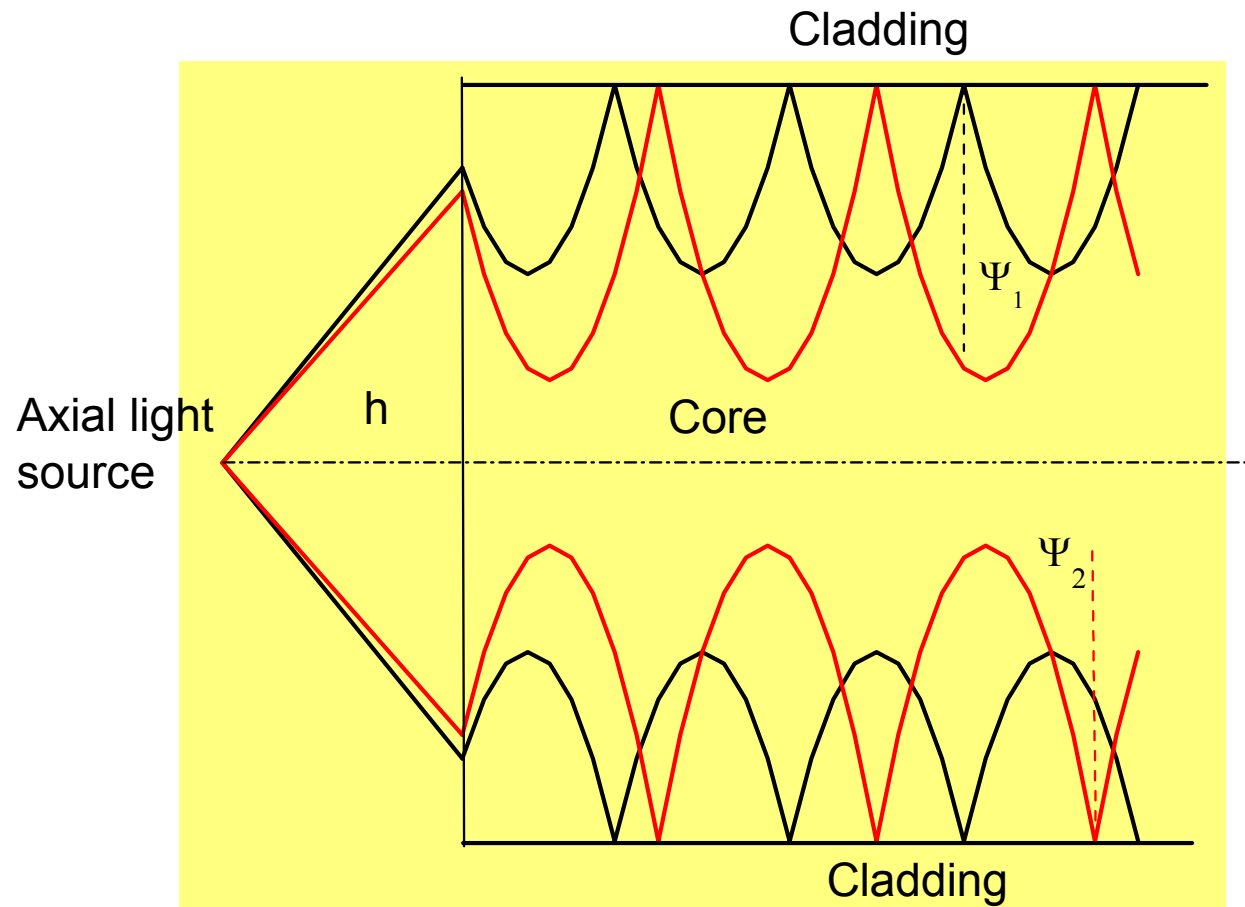
V. Matejec et al., *Sens. Actuators B* 38-39 (1997) 334-338

# CONTROL OF ANGLE $\theta$

Multimode fibers with inverted graded-index (IGI) refractive-index profile; Enable control of  $\theta$  on the core/cladding boundary



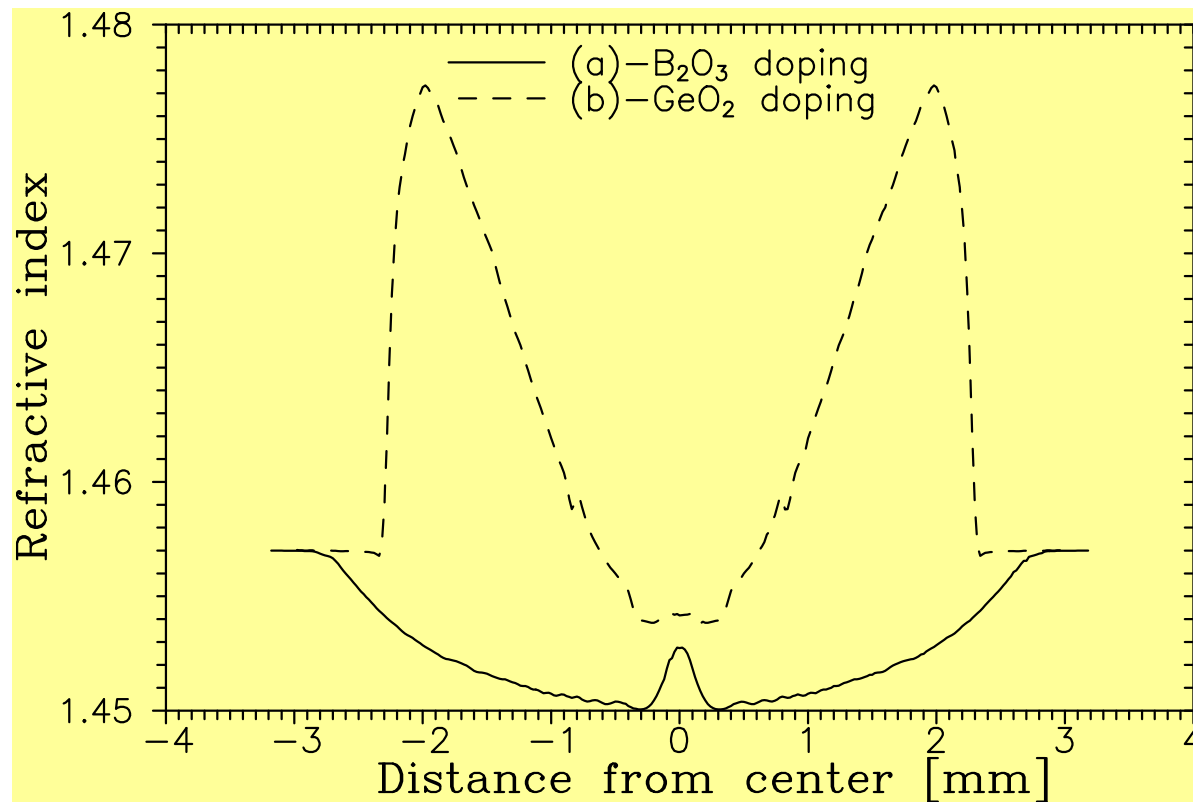
# IGI FIBER RAY TRAJECTORIES



**Nearly the same angle incident angle  $\Psi$  on the core/cladding boundary for different rays  $\rightarrow$  the same sensitivity**

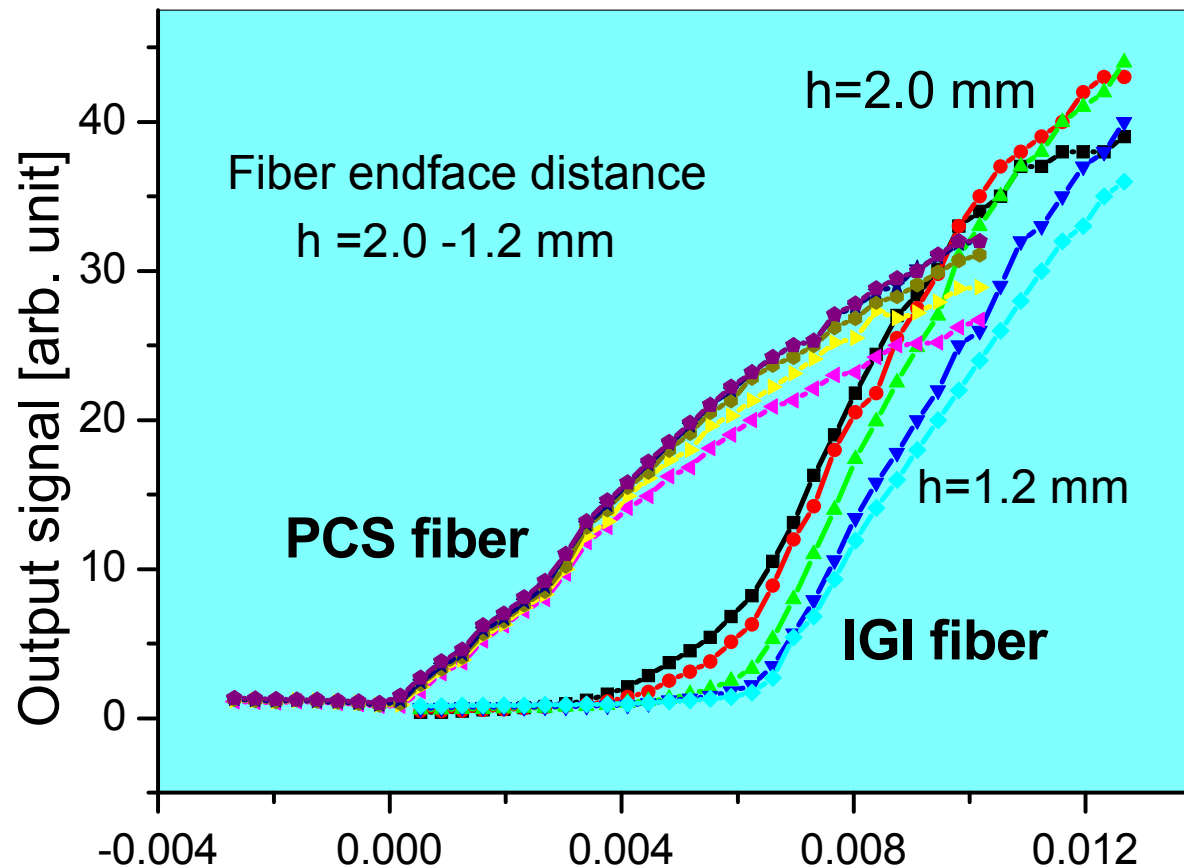
# PREPARATION OF IGI FIBERS

MCVD method, boron oxide or germanium oxide silica doping;  
Sensitivity to refractive index can be controlled by doping in a range 1.45-1.51. Fiber diameter 100-500 $\mu\text{m}$



Minimum RI determines detected refractive index

# IPE IGI FIBER – BORON OXIDE DOPED



Core/cladding refractive-index difference

**Sensitivity to RI changes for  $n=1.45$**

M. Chomat et al., *Sens. Act B* 87, 257-268 (2002)

# USING OF IGI FIBERS

**IGI fibers tested in SPR biosensors, SPR sensors for detection of toluene in water - Collaboration with Ecole Centrale de Lyon**

**IGI fiber for toluene detection using SPR sensor, detection length 1,5 cm, Au layer 50 nm coated with detection membrane;**

**Refractive-index changes of the membrane due to toluene penetration in it detected (limit of detection mg/l)**

**F. Bardine, These de la Universite Claude Monnet, Saint Etienne, 2004**

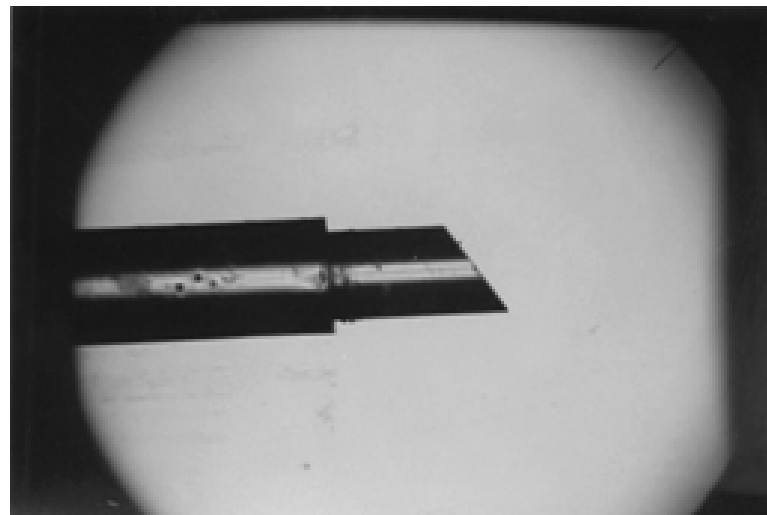
# OTHER IPE APPROACHES FOR SENSITIVITY IMPROVING

## Control of $\theta$

Excitation by an inclined collimated beam

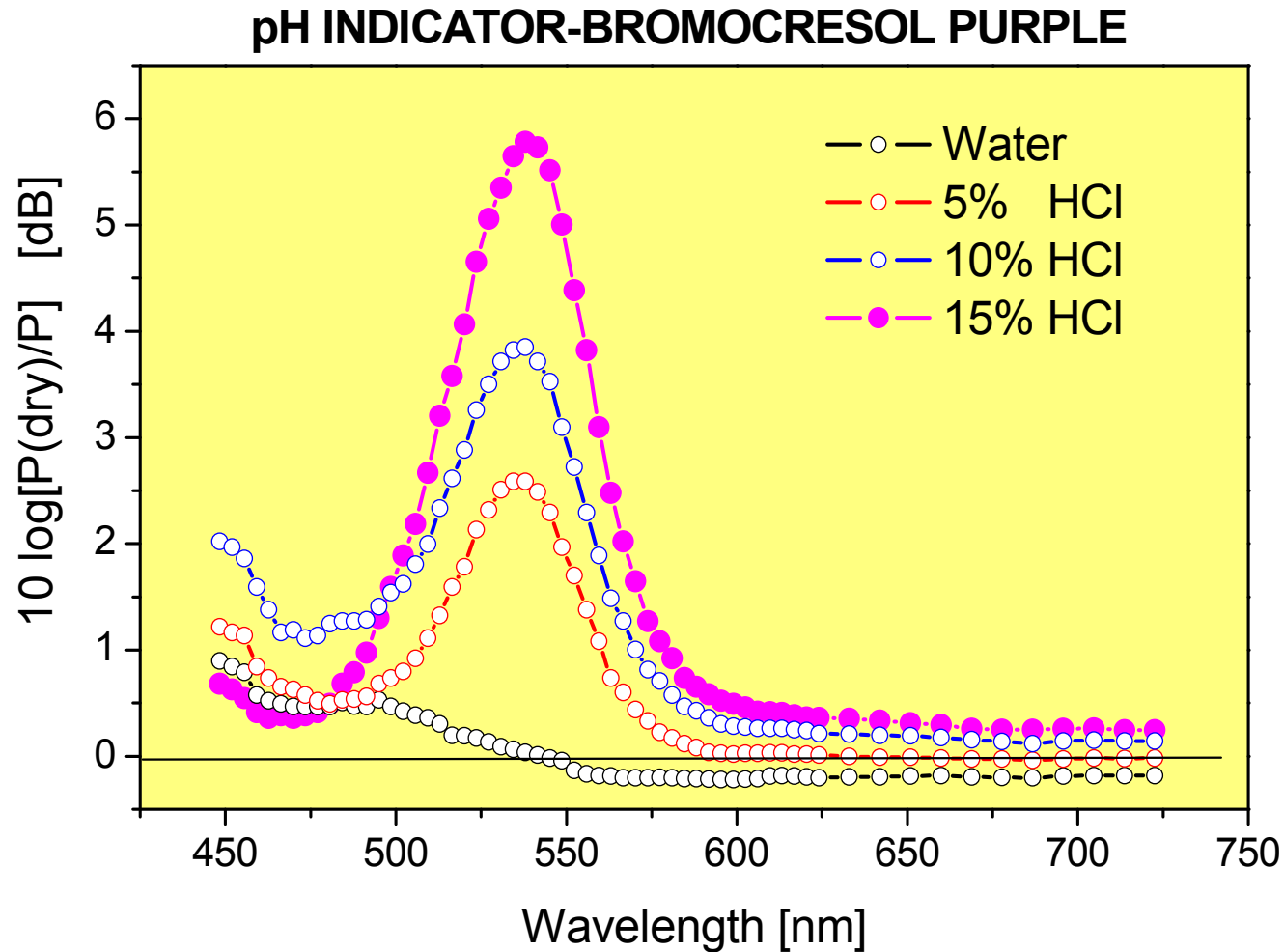
Beveled fibers

Output mode filters



Beveled fiber

# pH MEASUREMENTS WITH PCS FIBERS



pH transducer in a porous layer applied onto PCS fiber



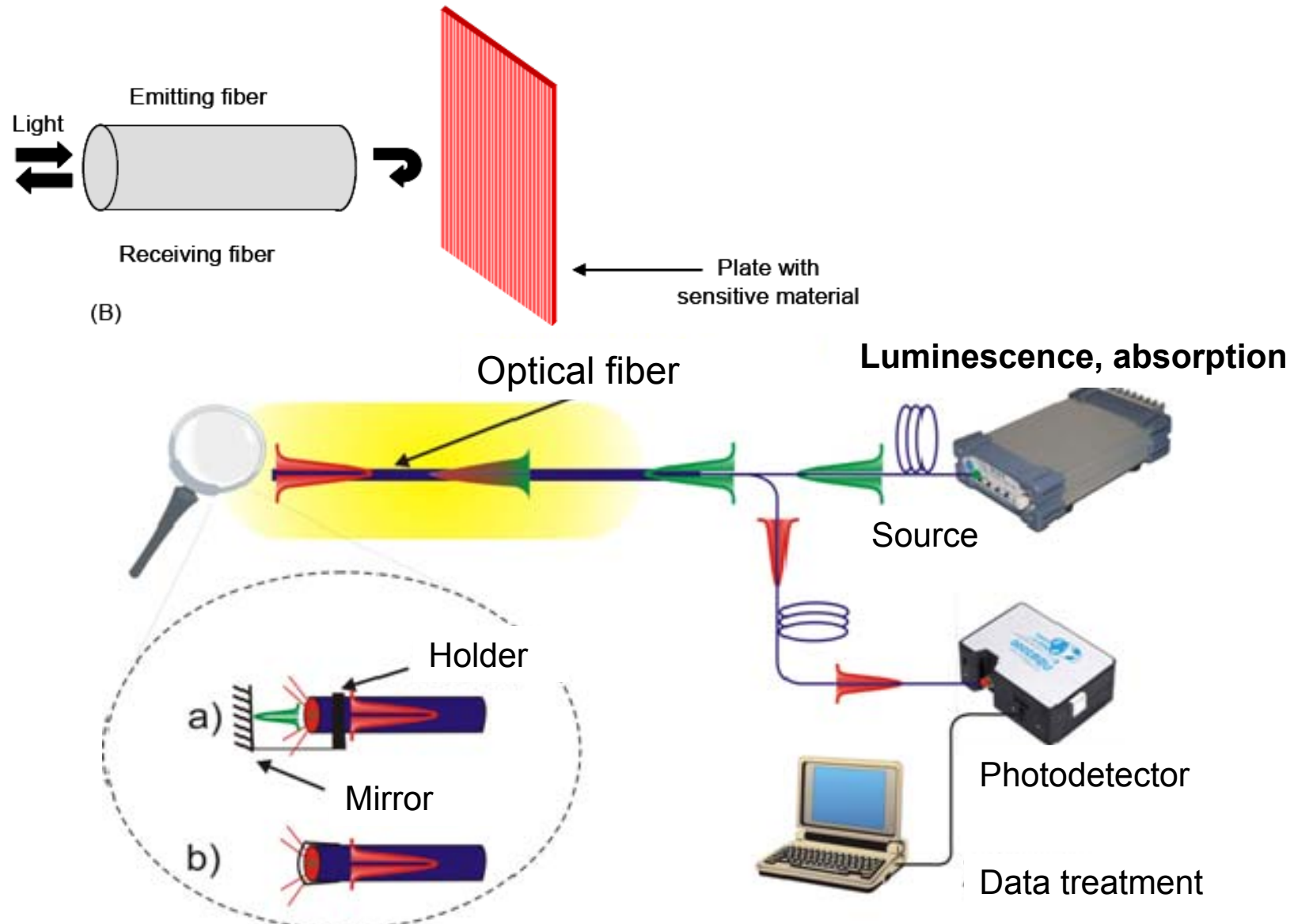
# SENSORS OF TOLUENE ON PCS FIBERS

- Gaseous toluene – 0.003%
- Toluene in water – 1-2 mg/l

Practical use – detection of leakages of petrol from tanks, petrol lines

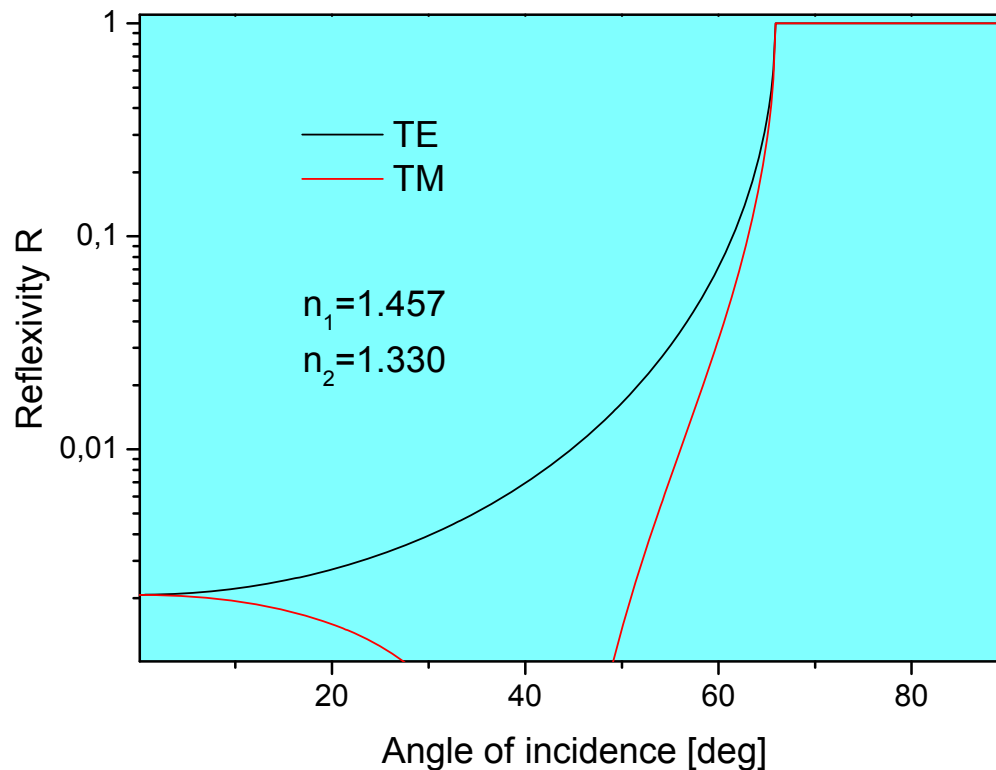
Control of quality of waste water from petrol refinery

# REFLECTION FOCUS - PRINCIPLE



# REFLECTION SENSOR - LIMITATIONS

Low reflectivity due at small angle  $\theta$  ( %) – necessary to use approaches for R increase



**Evanescent sensor**  
 $\theta > 80\text{deg}$ ,  $R > 0.9$

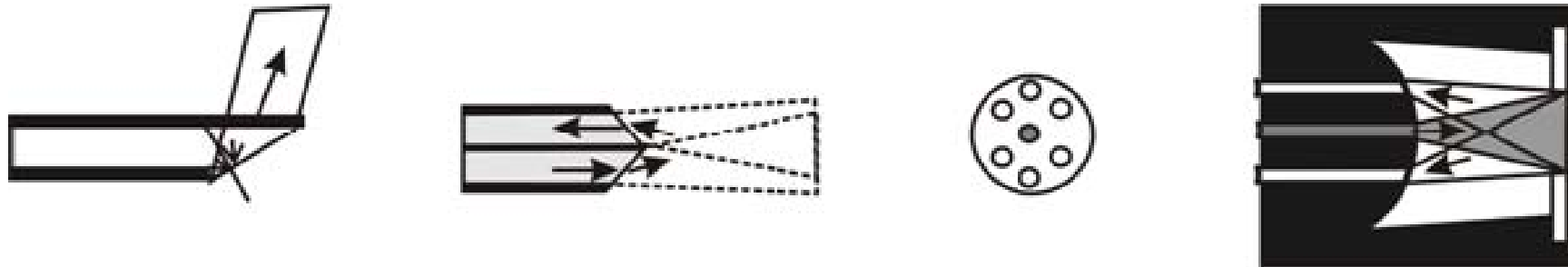
**Reflection sensor**  
 $\theta \sim 0 \text{ deg}$ ,  $R = 0.04$

$$R = \left( \frac{n_1 - n_2}{n_1 + n_2} \right)^2$$

# IMPROVEMENT OF REFLECTIVITY

**Small R + Small reflection area  $\rightarrow$  low intensity of reflected light**

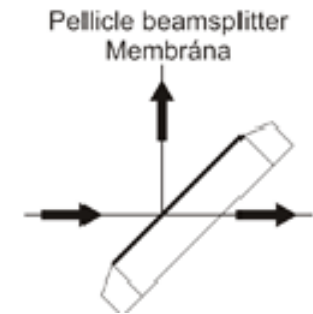
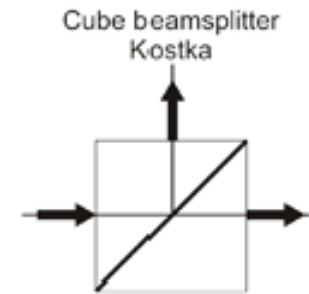
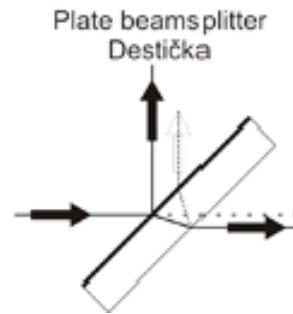
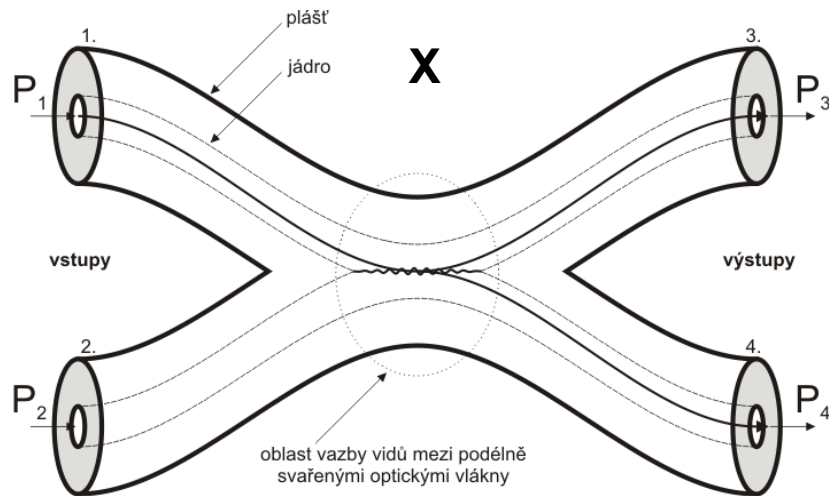
**Different approaches are used for improving intensity of reflected light – beveled fiber ends, fiber bundles, mirrors**



**U. Utziger et al., „Fiber-optic probes for biomedical optical spectroscopy“, J. Biomedical Optics 8(1) (2003) 121–147**

# INSTRUMENTATION

## Fiber couplers (X, Y), beam splitters, connectors



SMA

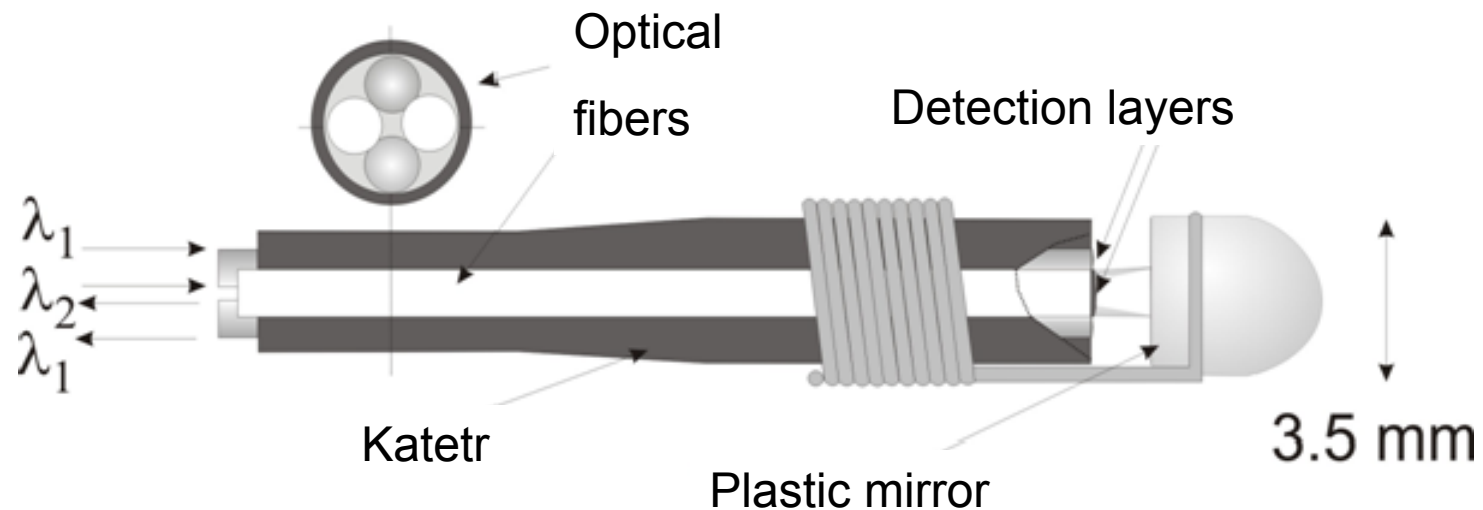
FC



Commercially  
available

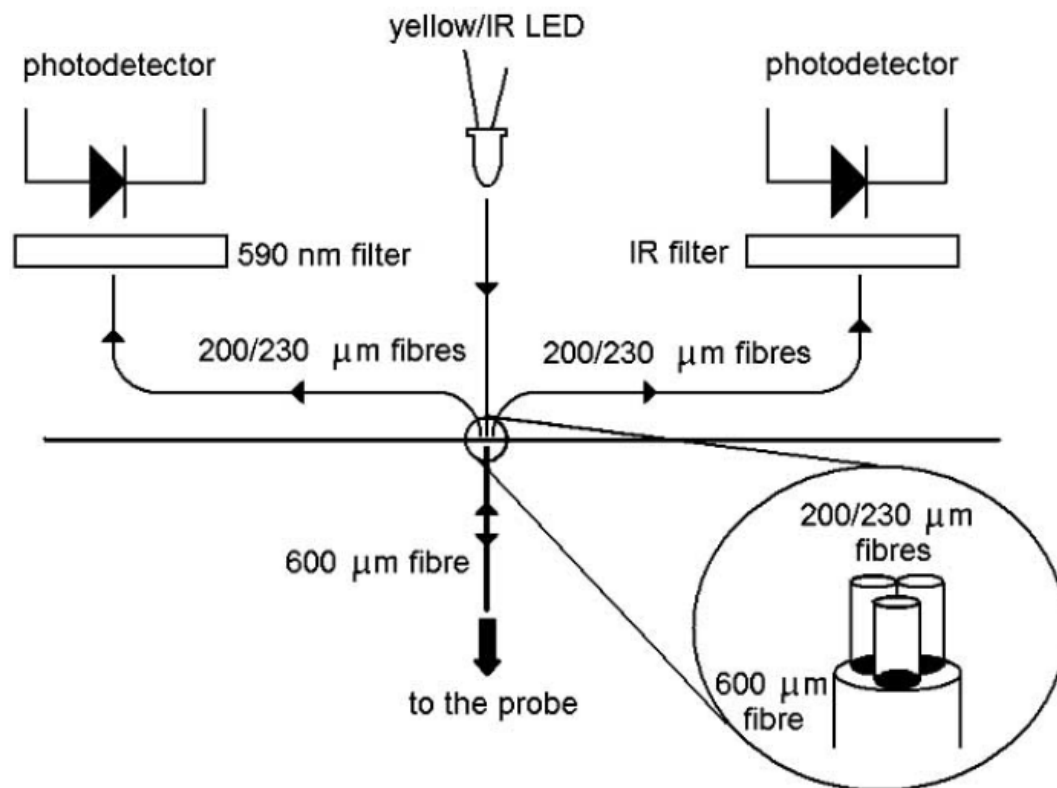
# REFLECTION SENSOR FOR pH IN STOMACH

Absorption pH transducers bromophenol blue and thymol blue (pH range 1-8)



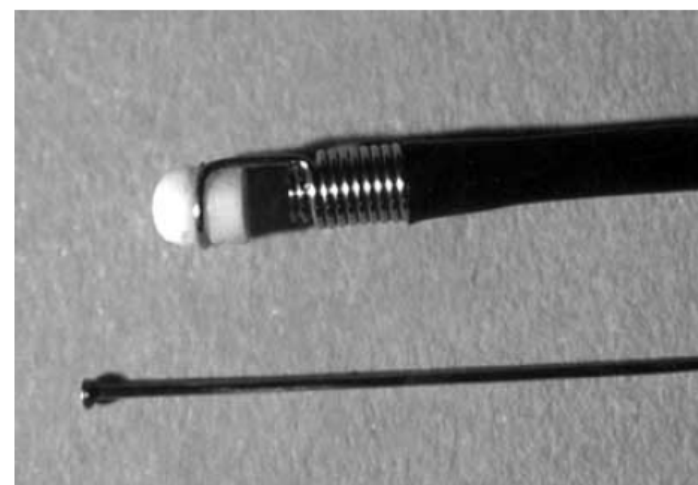
CNR Florence, Italy, F. Baldini, A. N. Chester, J. Homola, S. Martelluci: „*Optical chemical sensors*“, Springer (2006)

# REFLECTION SENSOR OF BILIRUBIN



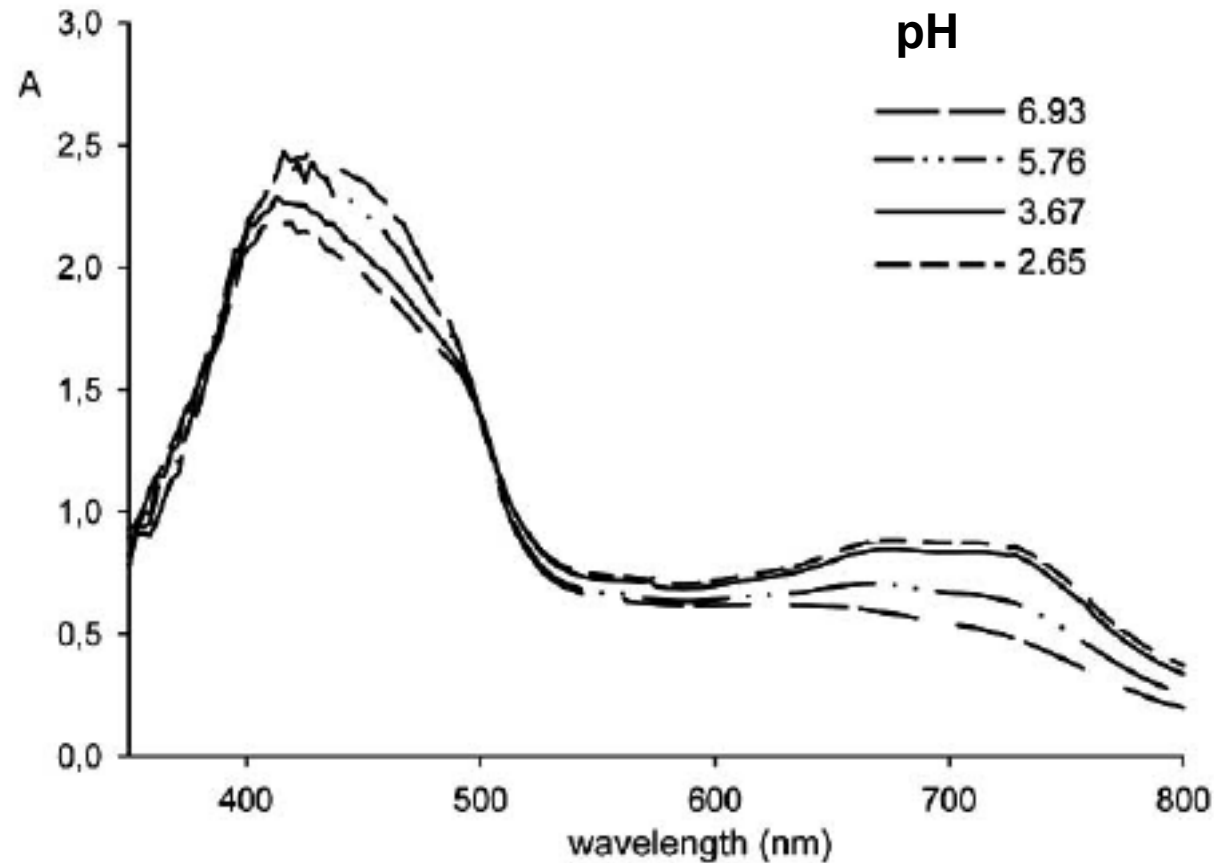
Measurement of  
absorption spectra  
of bilirubin

No transducer



F. Baldini et al., *Current Analytical Chemistry*, 2008, 4, 378-390 ( for  
detection of carbon dioxide in stomach)

# ABSORPTION SPECTRUM OF BILIRUBIN

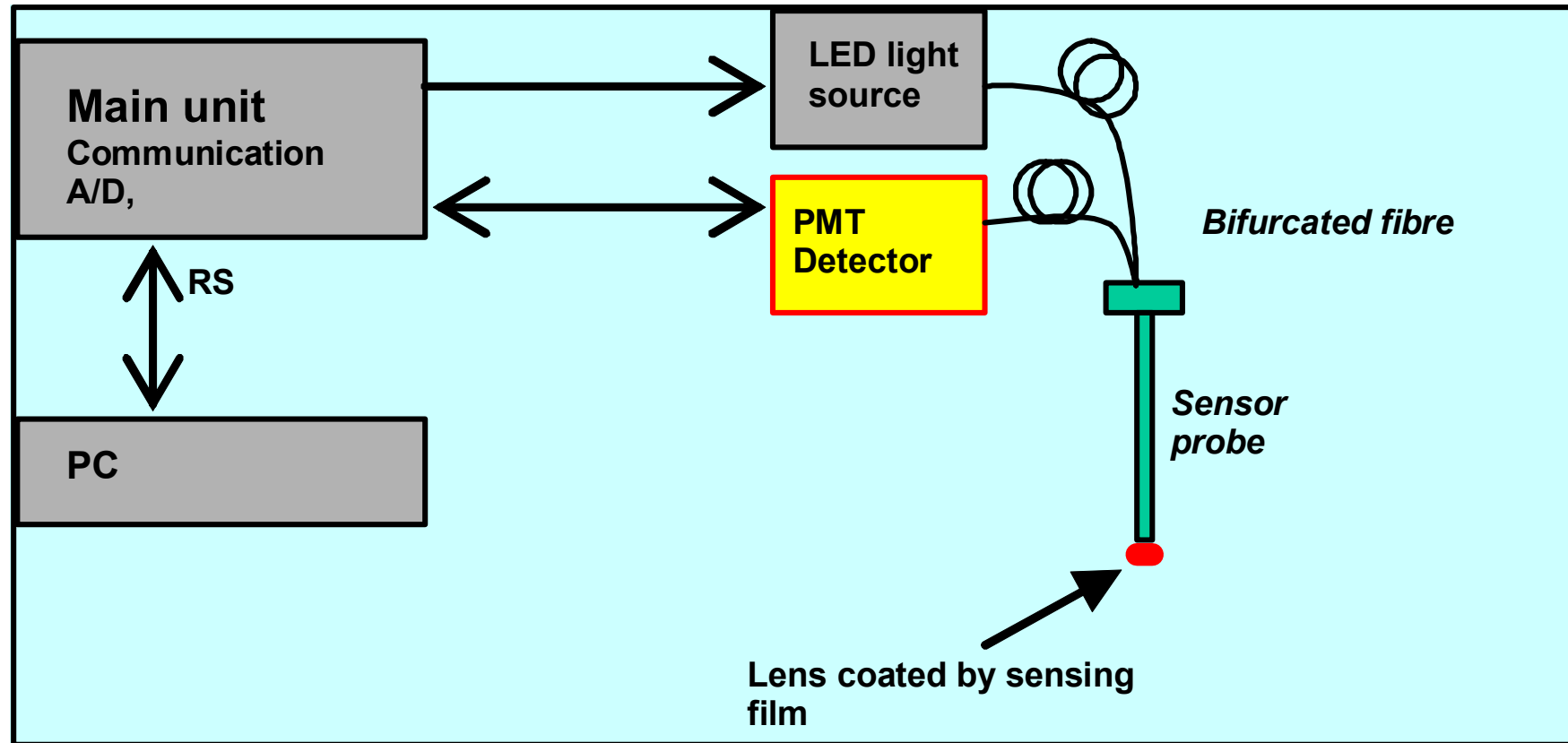


**Calibration:**

**absorbance 470nm (measurement)/ 590 nm (reference)**

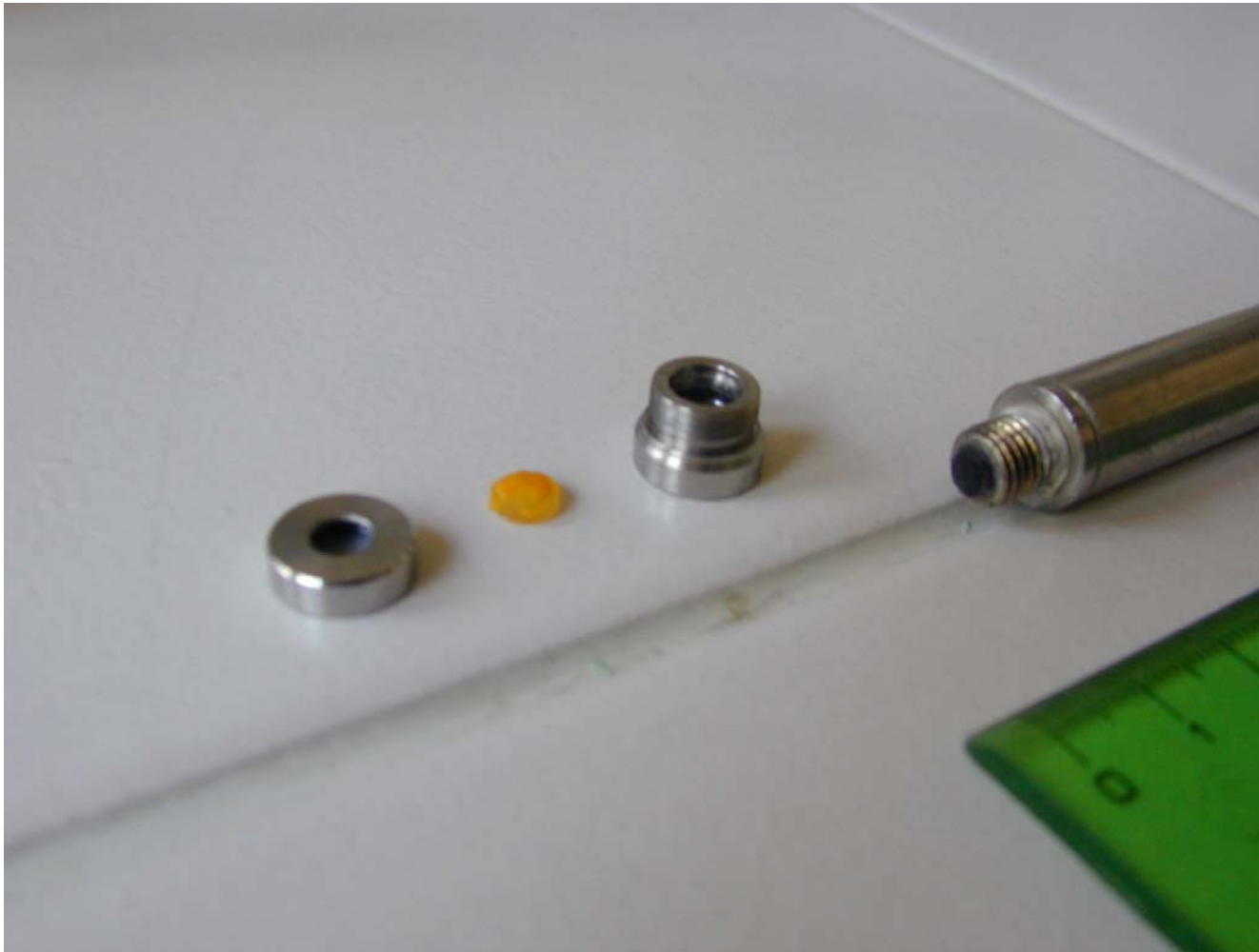


# IPE REFLECTION SENSOR OF OXYGEN, GLUCOSE



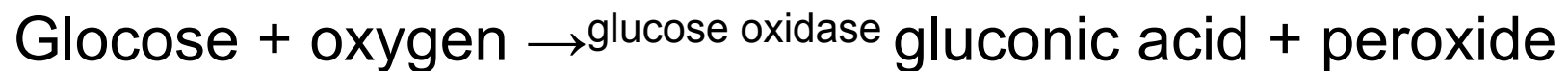
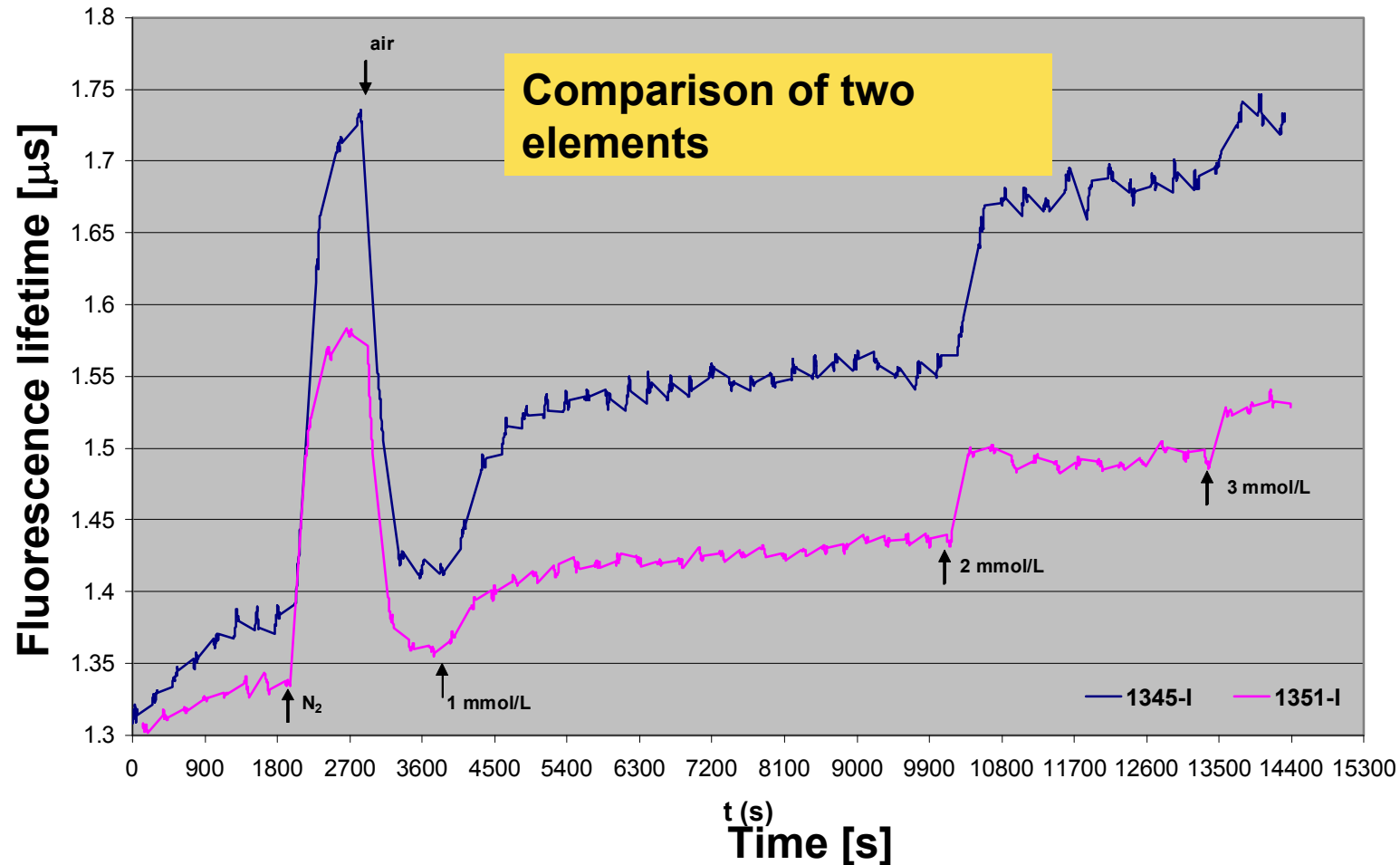
**Oxygen – quenching of fluorescence Ru complex**  
**Glucose – oxygen detection in enzymatic reaction**

# SENSING HEAD AND DETECTION LENS

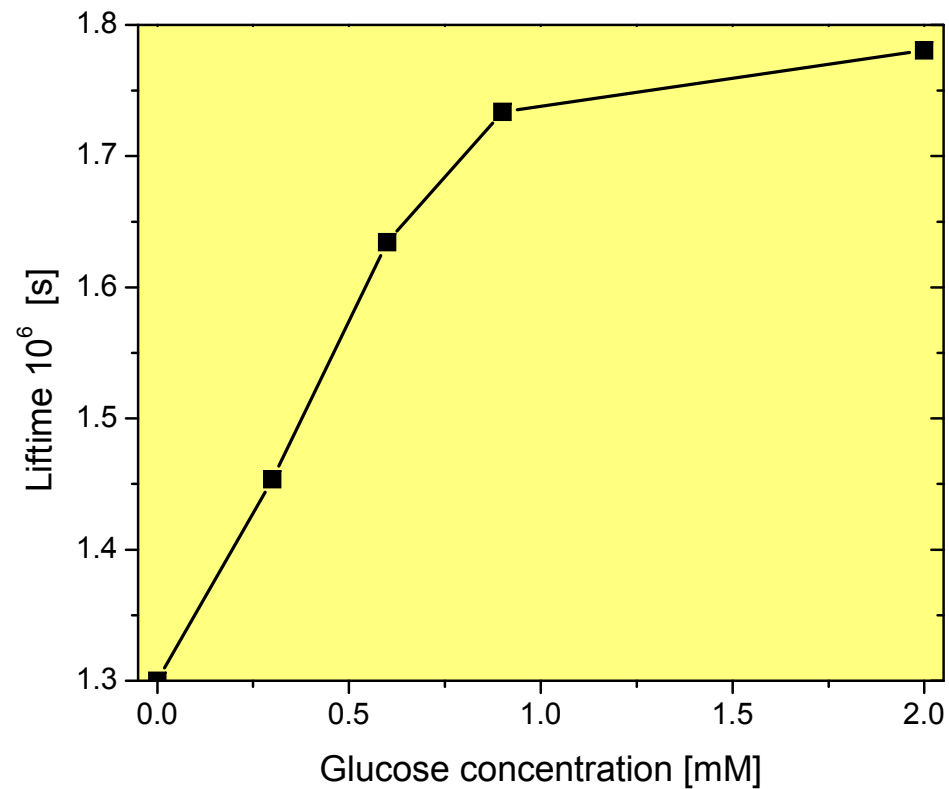


## Fluorescence lifetime measurements

# TIME RESPONSE TO OXYGEN AND GLUCOSE

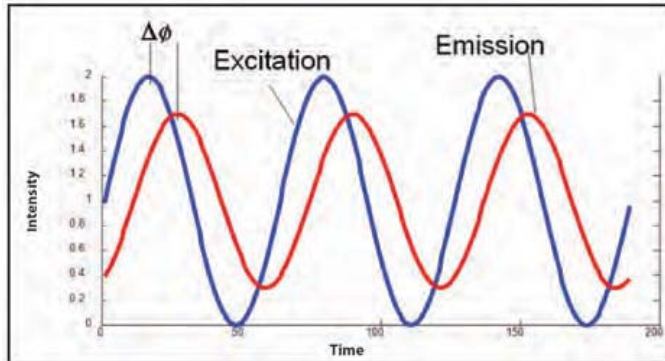


# GLUCOSE SENSOR - CALIBRATION CURVE



Detection limit 0.2 mM

# COMMERCIAL OXYGEN SENSORS



**Fluorescence lifetime measurements, phase modulation**

[www.oceanoptics.com/Products](http://www.oceanoptics.com/Products)

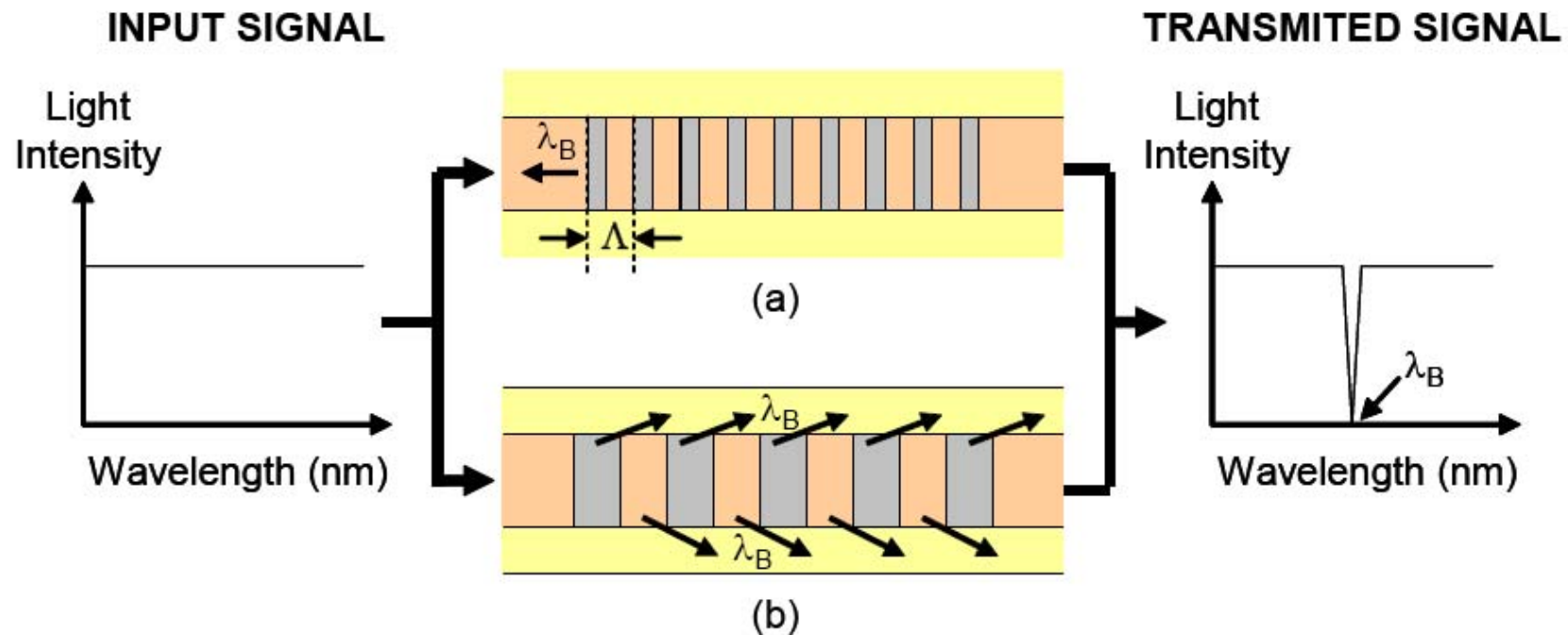


**Detection elements**



**Detection of oxygen in plants**

# OPTICAL GRATINGS IN FIBERS



Longitudinal periodic changes of refractive index in fiber  $\rightarrow$  light wave is reflected on grating at resonance condition:

(a) Bragg gratings (FBG)  $\lambda_B = 2n_{\text{eff}}\Lambda_B$  ( $\Lambda_B \ll \lambda$ )

(b) „Long Period Gratings (LPG)“  $\lambda_{\text{LPG}} = (n_{\text{CO}} - n_{\text{cl}}^m)\Lambda_{\text{LPG}}$  ( $\Lambda \sim 10^2 \mu\text{m}$ )

# GRATING PREPARATION AND PROPERTIES

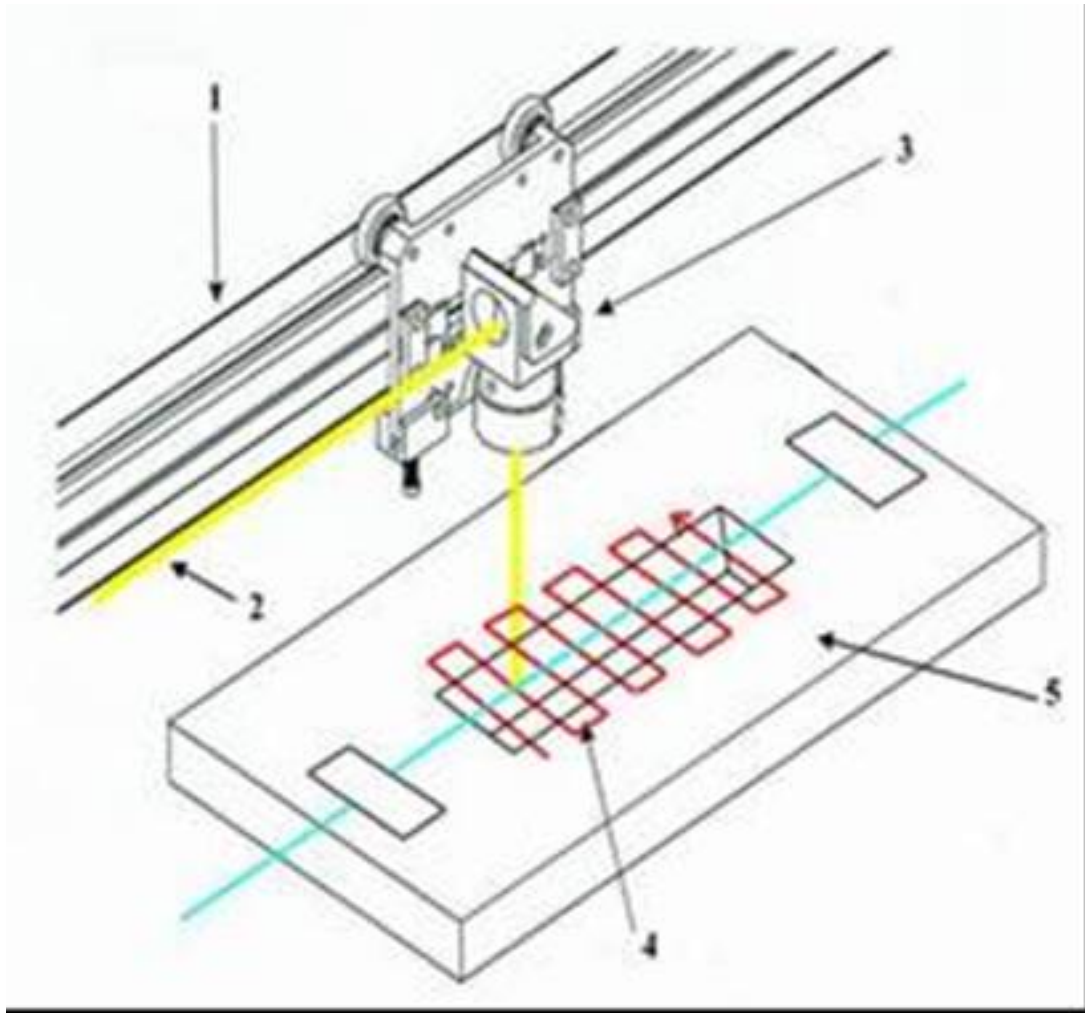
- FBGs - inscription by UV lasers into fibers saturated by hydrogen  
Sensitive to **effects influencing the core** – temperature, strains
- LPGs - inscription by CO<sub>2</sub> laser  
Sensitive to **effects influencing both core and cladding** – temperature, stresses, refractive index – chemical detection

K.O. Hill, Gerald Meltz, J. Lightwave Technol. 15, 1263-1276, 1997

J. Canning, J. Sensors 2009, Article ID 871580, 17 stran - vlákna

I. J. G. Sparrow, J. Sensors 2009, Article ID 607647, 12 stran - planární

# IPE PREPARATION OF LPGs

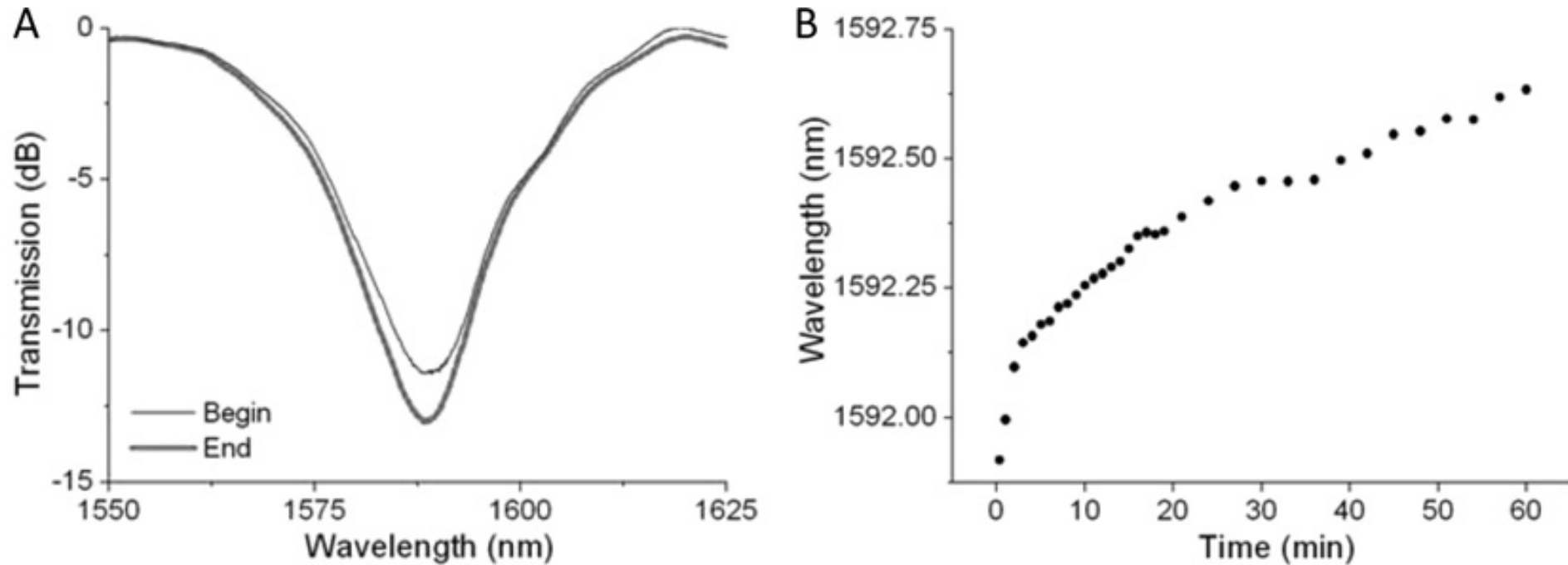


## CO<sub>2</sub> laser

Beam crossing through the fiber heats up the fiber → changes of mechanical stresses → change of RI due to photoelasticity



# DNA DETECTION BY LPG IN SM FIBER

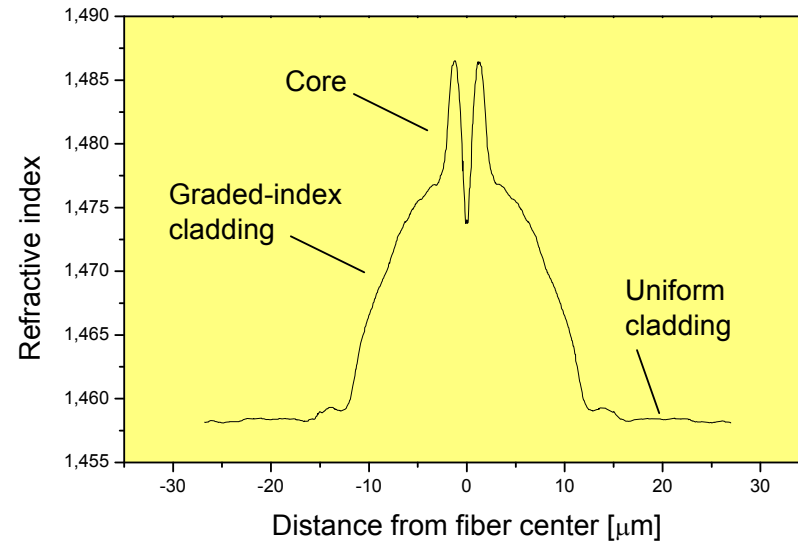
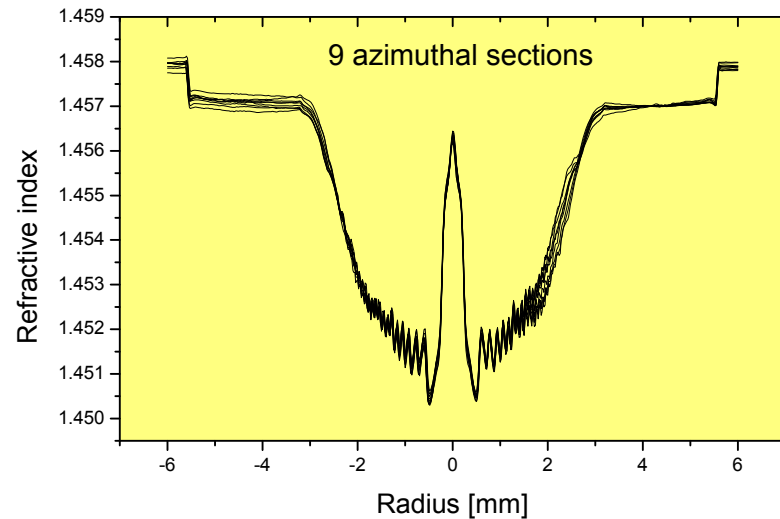


$$\Lambda = 161 \mu\text{m}$$

1  $\mu\text{M}$  ssDNA (5-GCACAGTCAGTCGCC-NH<sub>2</sub>-3) in PBS buffer

A.V. Hine et al., *Biochem. Soc. Trans.* (2009) 37, 445–449

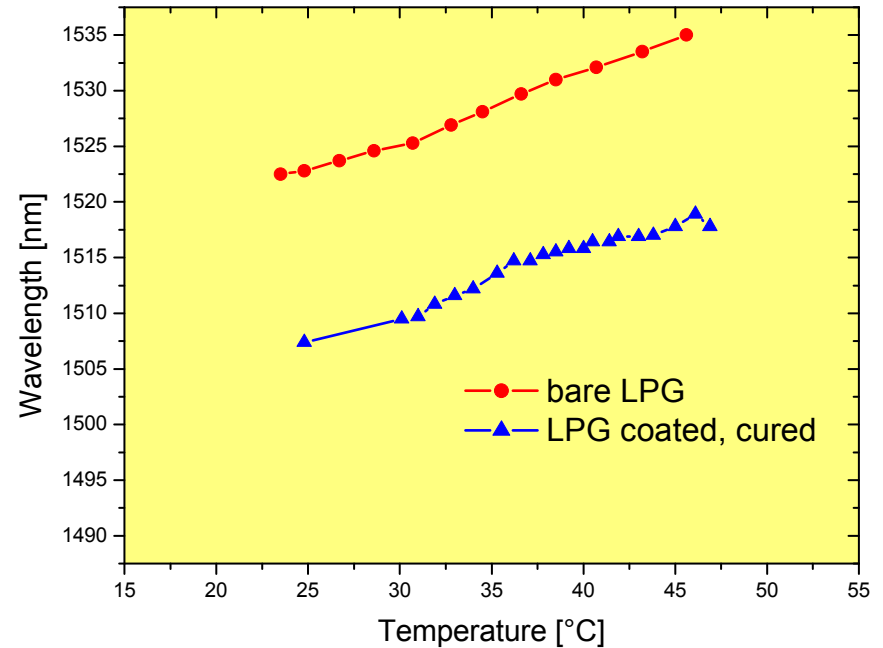
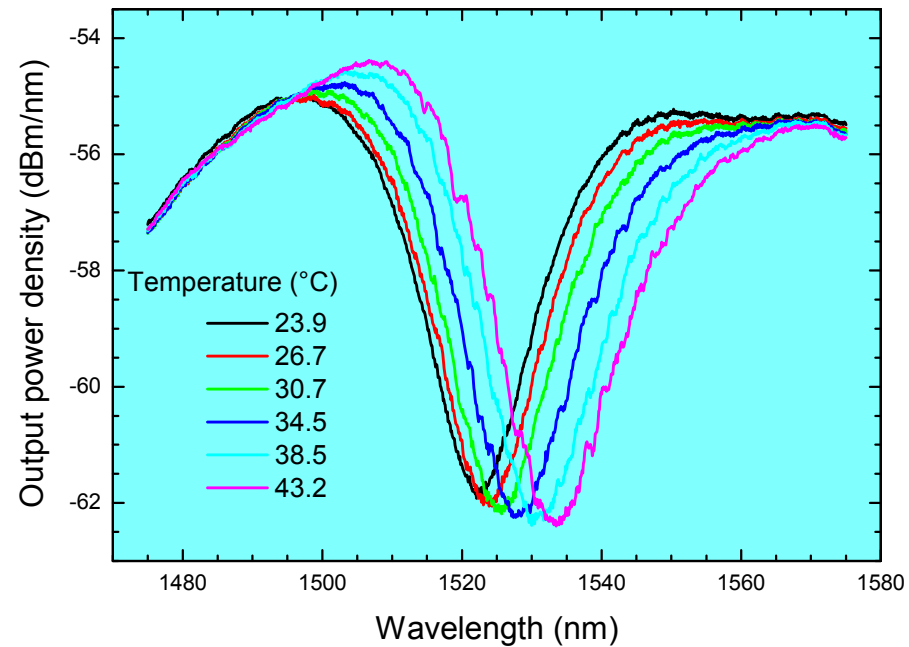
# IPE LPGs IN GI AND IGI FIBERS



SM core, IGI, GI cladding allow to control detection selectivity to temperature, stresses  
LPGs in standard SM fibers sensitive to a number of effects

**F. Todorov et al., Sensor Lett. 7, 979-983 (2009)**

# LPG TEMPERATURE SENSITIVITY

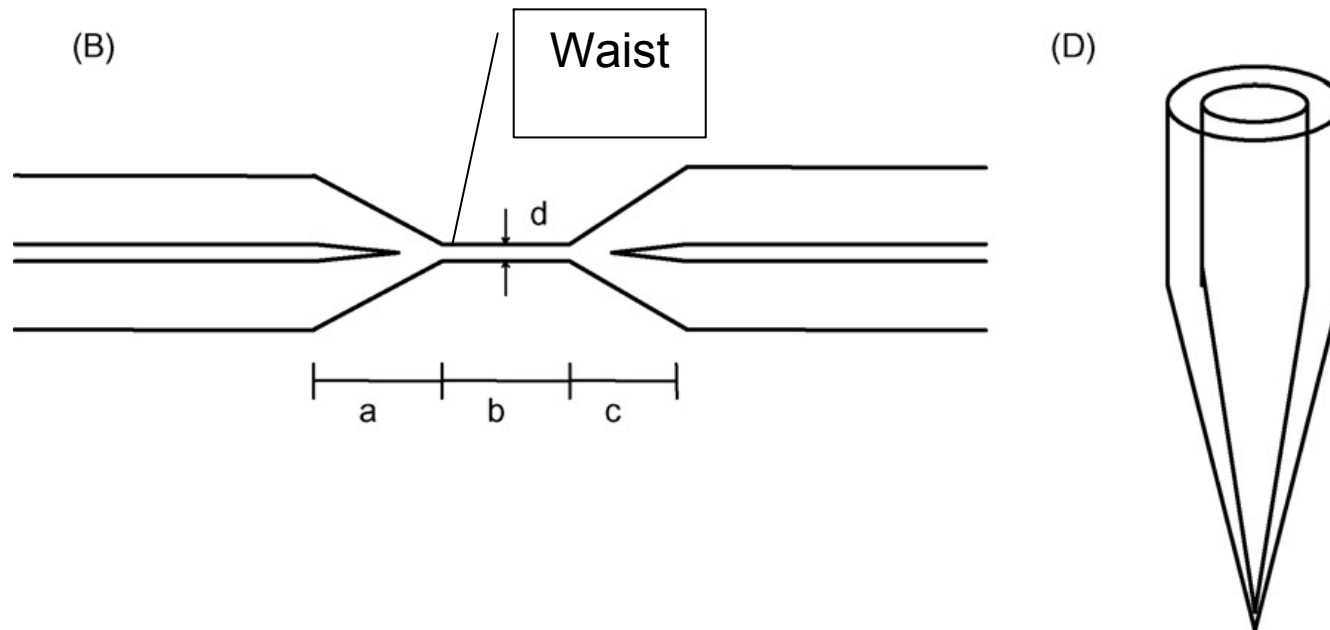


$\Lambda_B = 500 \mu\text{m}$ ; Sensitivity 0,5 nm/°C

# OPTICAL MICROSENSORS

- **Fiber-optic tapers**
- **Optical nanoparticles – PEBBLE** - „*probes encapsulated by biologically localized embedding*“;
- **Microresonators with whispering gallery modes (WGM)**

# FIBER-OPTIC TAPERS

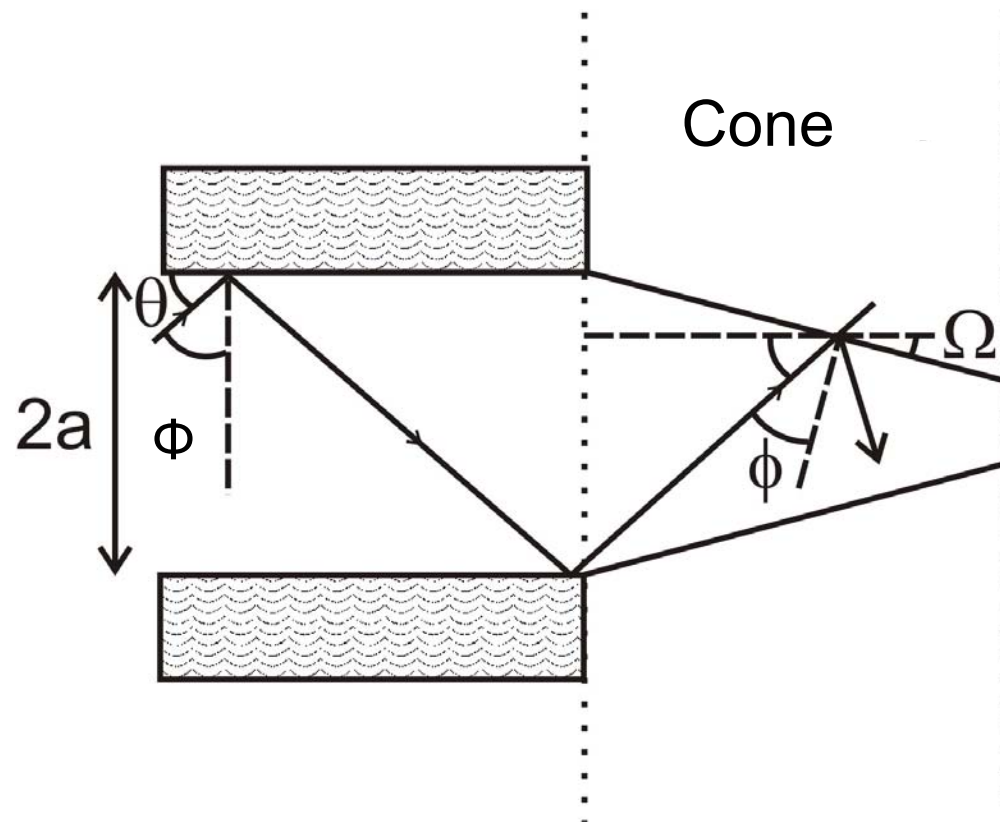


**Make possible to decrease core diameter and angle  $\theta$  on the core/cladding boundary  $\rightarrow$  sensitivity increase.**

**Tapers with two cones (B) – “biconical tapers”**

**Tapers with one cone (D)- “fiber tips”**

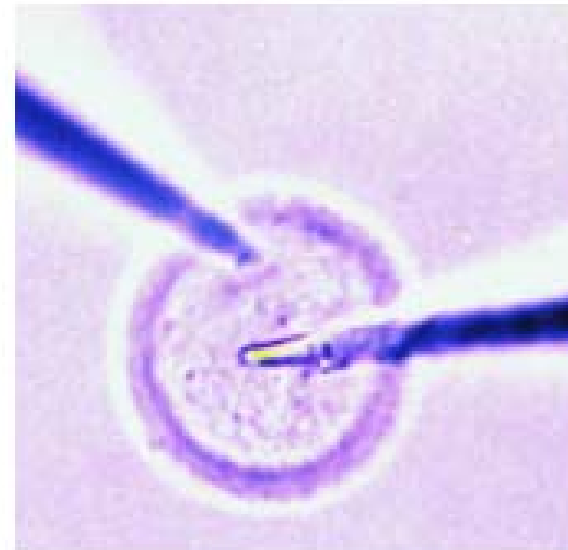
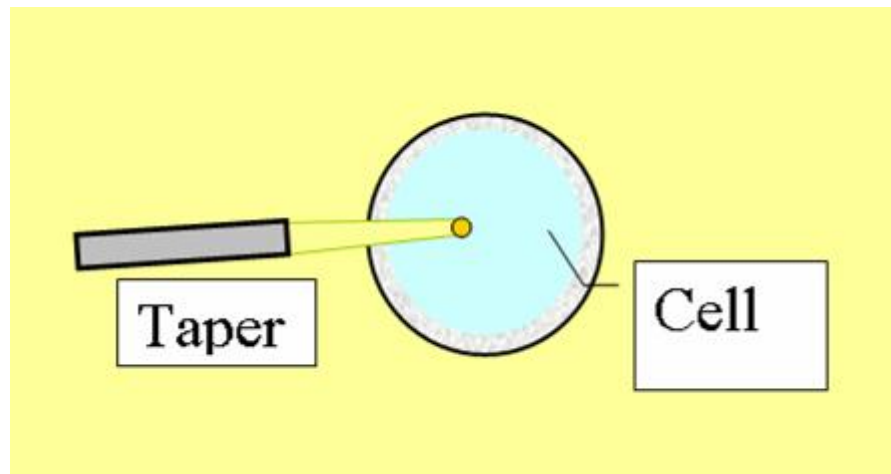
# TAPER – PRINCIPAL SCHEME



**$d < 2a$ ,  $\Phi < \theta \rightarrow$  increased power in evanescent wave**

# SENSORS ON FIBER TIPS

Optical fiber elongated into a tip that is coated with optical transducer



Tips sensitive to potassium in mouse oocyte cells  
(dimension  $100\mu\text{m}$ )

S.M. Buck et al. , *Talanta* 63 (2004) 41

# SENSORS ON FIBER TIPS - REFERENCES

## - R. Kopelman et al. (USA)

E. J. Park et al., J. Mater. Chem. **15** (2005) 2913 – oxygen detection

M.R. Shortreed et al., Sens. Actuators **B 38-39** (1997) 8 – K detection

## -T. Vo-Dinh et al., (USA)

T. Vo-Dinh et al., J. Nanoparticle Research **2** (2000) 17-

B.M. Cullum et al., Tibtech September **18** (2000) 388-review

T. Vo-Dinh et al., Anal Bioanal Chem **382** (2005) 918 -review

B. M. Cullum et al., Analytical Biochemistry **277** (2000) 25– benzo[a]pyrene tetrol in cancer cells

## *Optical sensors*

O.S. Wolfbeis, J. Mater. Chem. **15** (2005) 2657– fluorescence sensors

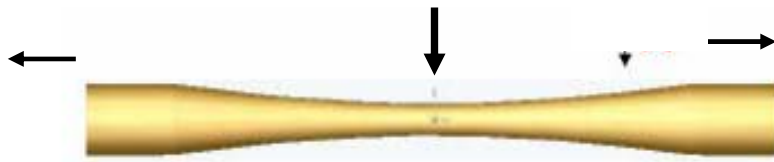


# FIBER TIP PREPARATION – THERMAL ELONGATION



Input optical fiber

Heating



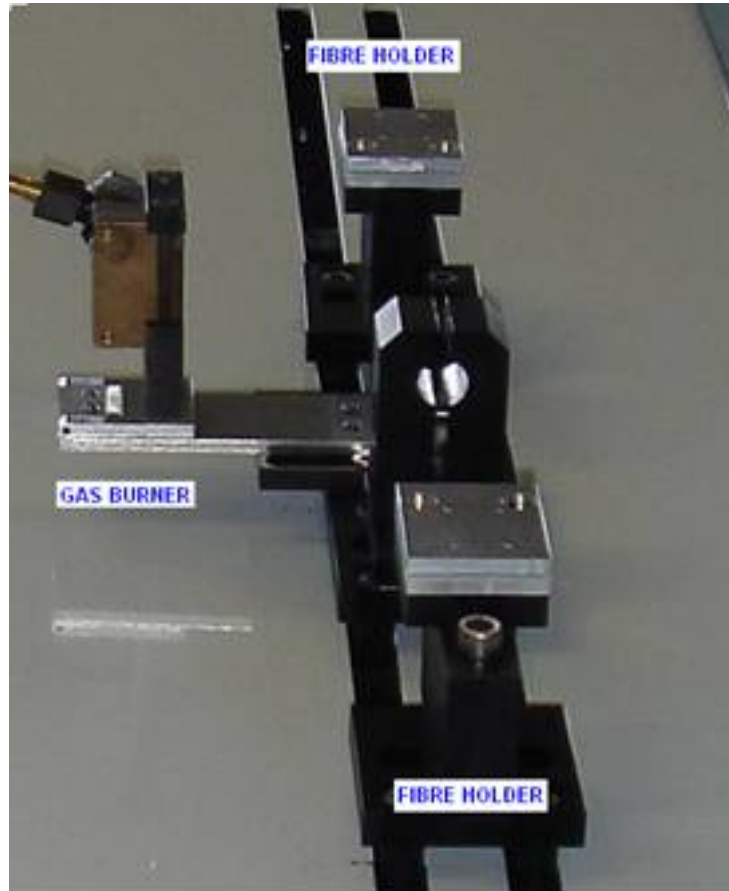
Heating up and elongation of fiber by a burner or CO<sub>2</sub> laser

Coating the taper with a metallic protective layer

Precise cutting

T. Martan et al., Proc. SPIE 7138 (2008), Article 71380Z DOI:  
10.1117/12.818000

# IPE TAPERIN APPARATUS



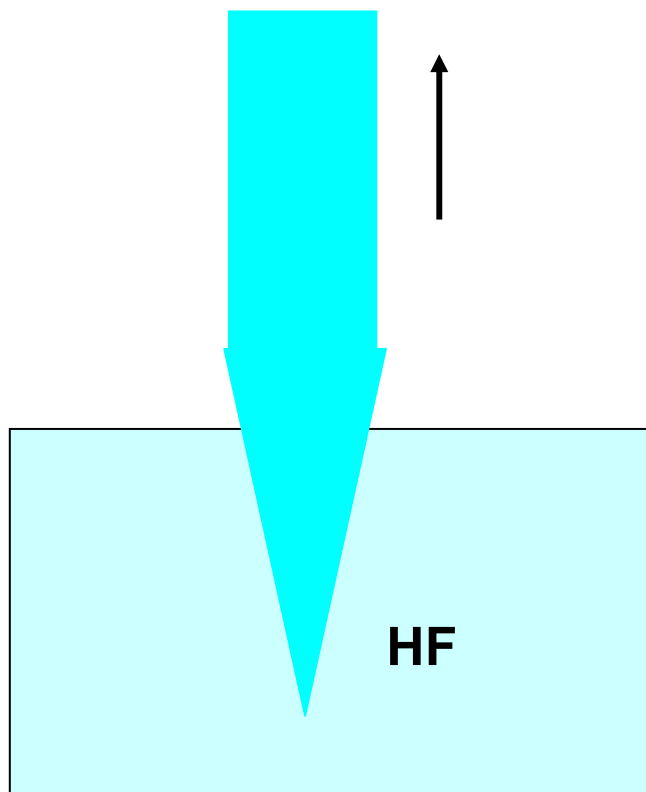
**Gas burner**

**PC-controlled elongation**

**Al coating prepared by  
sputtering**

**T. Martan et al., Proc. SPIE 7138  
(2008), Article 71380Z DOI:  
10.1117/12.818000**

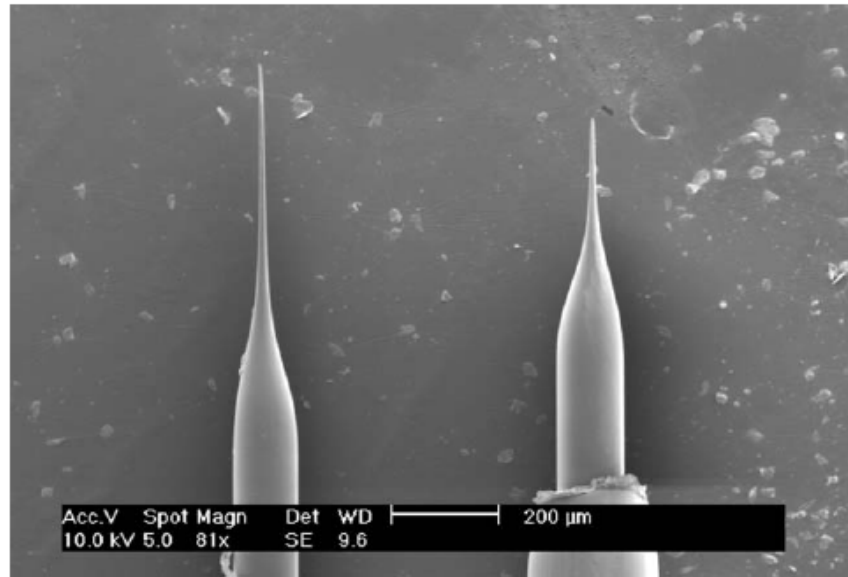
# FIBER TIPS PREPARED BY CHEMICAL ETCHING



Immersing silica fiber in a solution of hydrofluoric acid and its slow withdrawal a tip at the fiber end is formed

N. Nath et al., J. Anal. Toxicology 23 (1999), 460-467

# TIP PHOTOS - LITERATURE



**Tip diameter 50 nm**

**B. Cullum et al., Tibtech September 18 (2000) 388-review**

**E. J. Park et al., J. Mater. Chem. 15 (2005) 2913 – detection of oxygen**

# APPLICATION OF DETECTION LAYER ON TIP

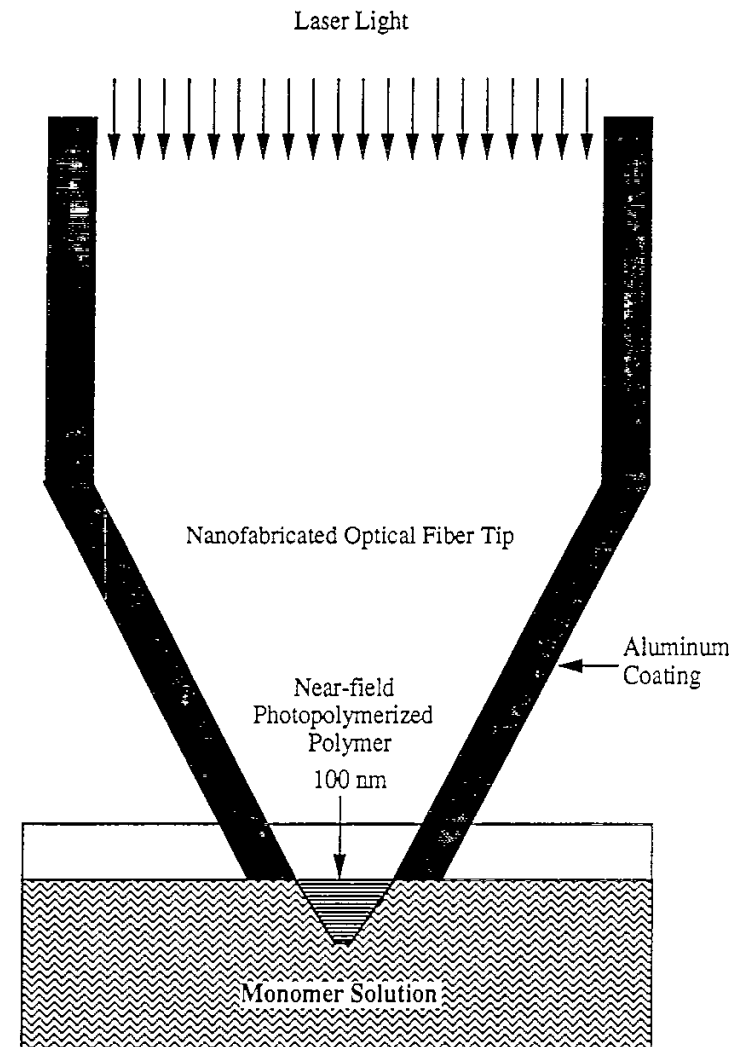
## Photopolymerisation

Monomer (UV acrylate) solution containing transducer

Launching UV radiation into the fiber a small amount of monomer is cured at the tip end

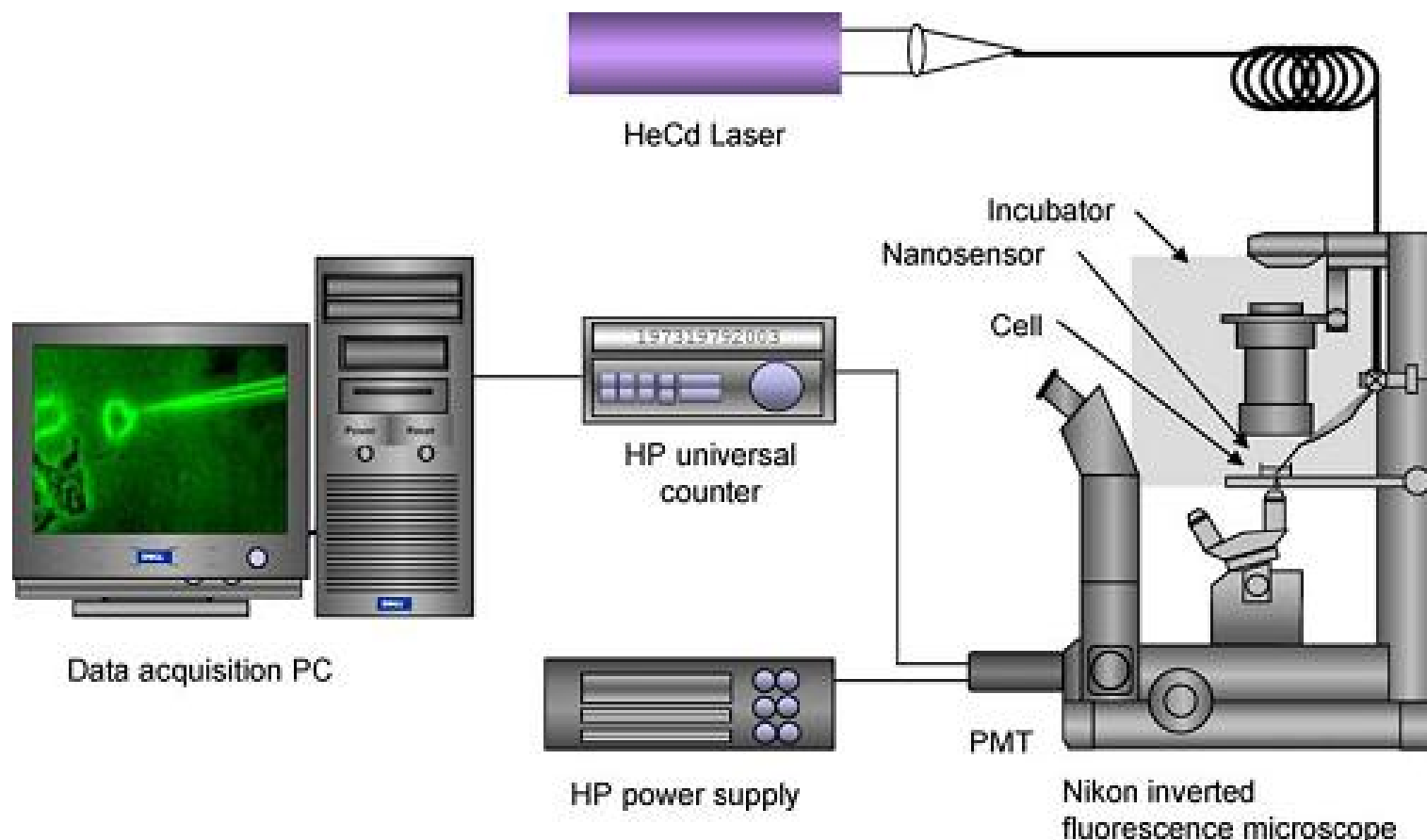
Only end of the tip coated

W. Tan et al., *Sens. Actuators B28 (1995) 157 - pH*



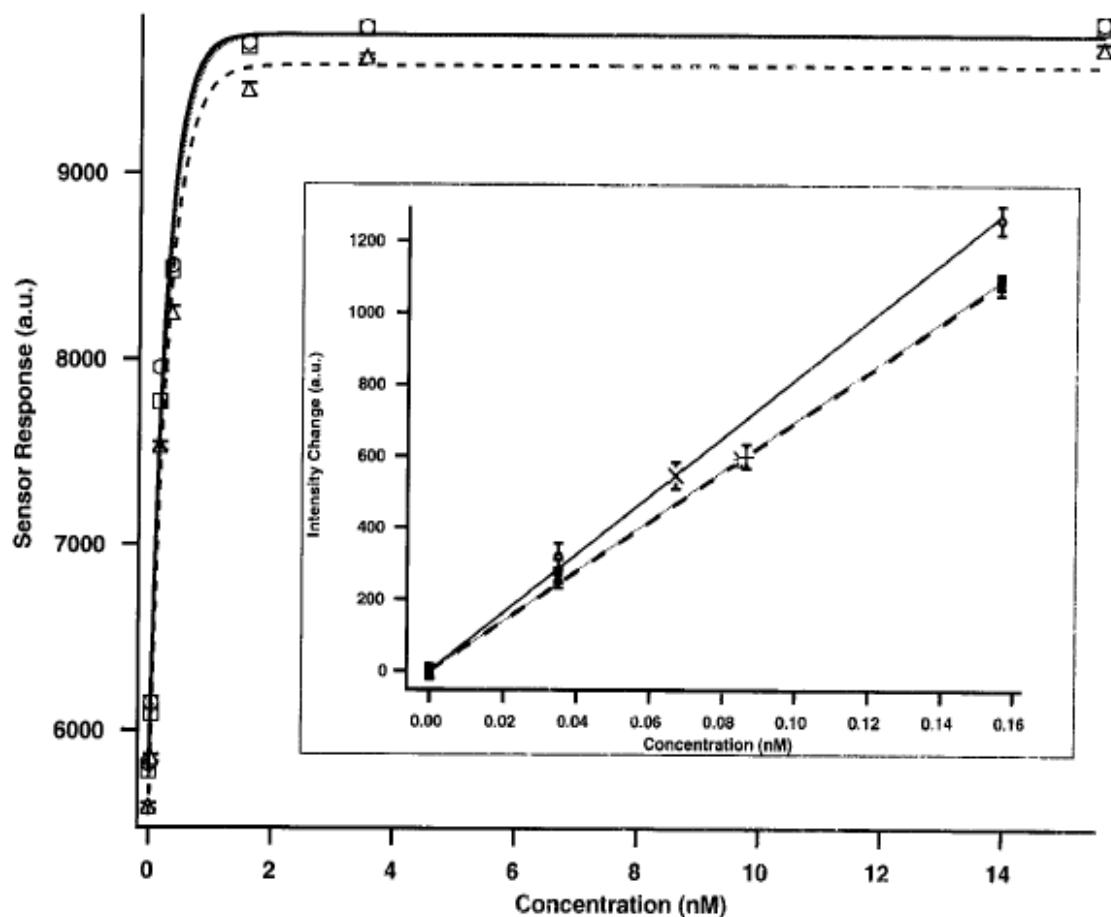
# MEASUREMENTS WITH TIPS

## Inverted fluorescence microscope



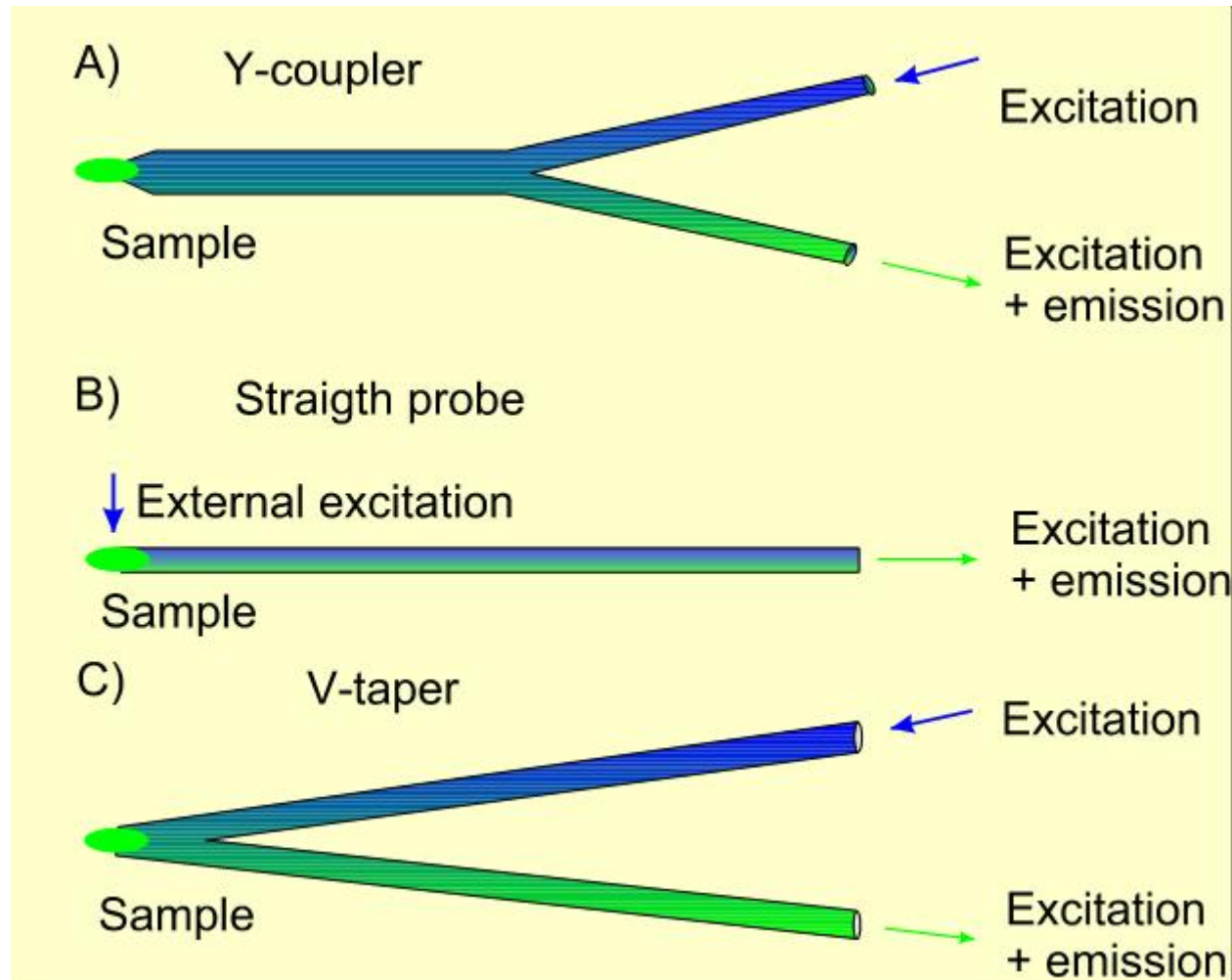
T. Vo-Dinh et al., *Anal Bioanal Chem* 382 (2005) 918

# DETECTION OF CANCER MARKERS



**B. M. Cullum et al., Analytical Biochemistry 277 (2000) 25** – Auto fluorescence of benzo[a]pyrene tetrol in rat liver cell

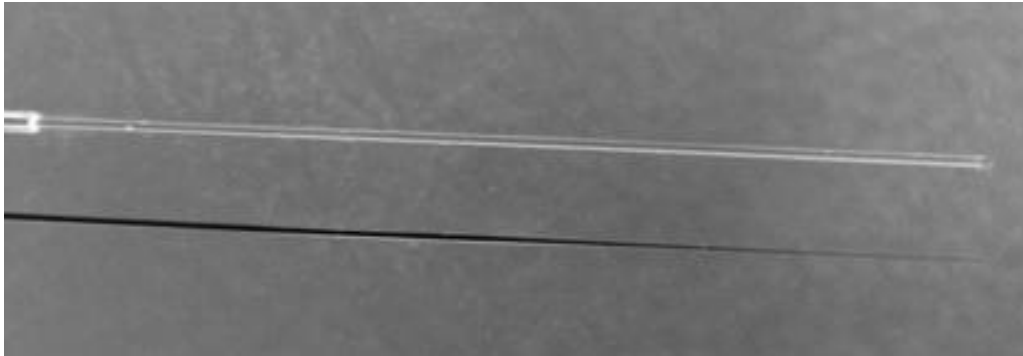
# IPE APPROACH - REFLECTION ARRANGEMENT



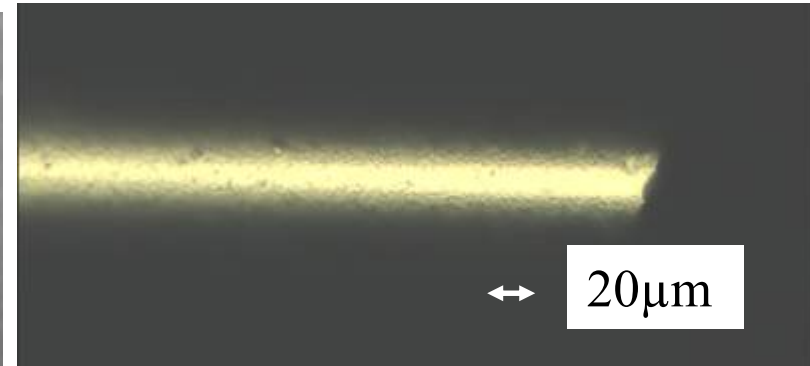


# IPE TAPERS

Taper coated with ITO coating and input fiber



Taper coated with Al



Taper coated with ITO, tip diameter 2 μm



# pH DETECTION MEMBRANES APPLIED ON TIP

**Porous membrane with fluorescence pH transducers  
BCECF or HPTS**

**HPTS= 8-hydroxypyrene-1,3,6-trisulfonic acid trisodium salt ( $\lambda_{exc} = 430$  nm,  $\lambda_{em} = 480$  nm)**

**BCECF=2',7'-Bis(2-carbonylethyl)-5(6)-carboxyfluorescein (Aldrich 14560) ( $\lambda_{exc} = 473$  nm,  $\lambda_{em} = 540$  nm)**

## Porous membrane

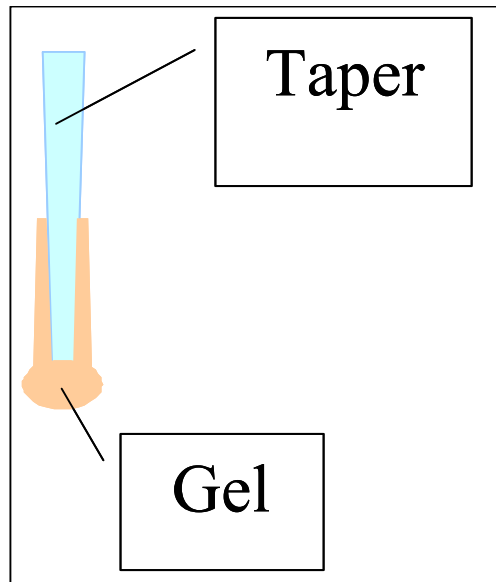
– TEOS, HCl,  $R_w=2$ , BCECF

– Propyltriethoxysilane + (3-glycidoxy)trimethoxysilane, HCl,  $R_w=2$ , HPTS, Ru phenanthroline chloride

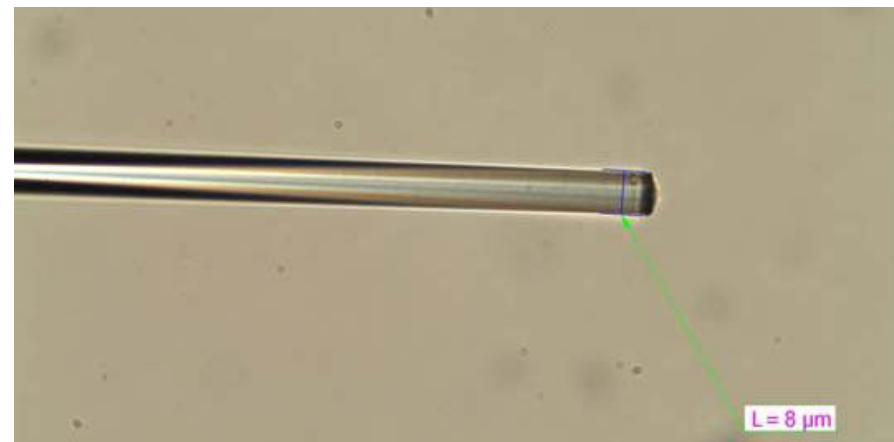
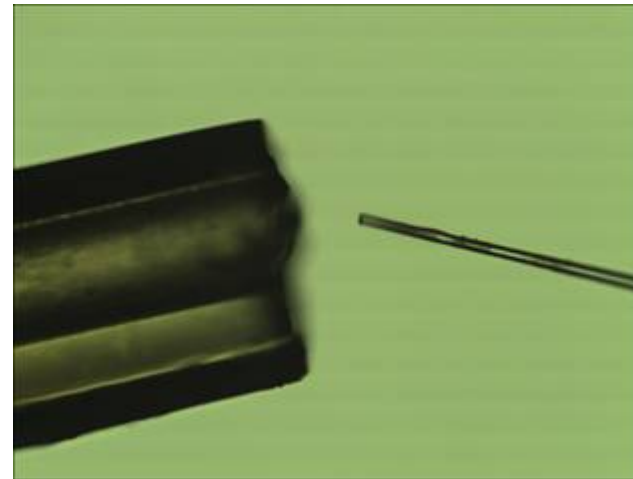
**Gel layers dried at 70 °C, 24 h**

# APPLICATION OF INPUT SOLUTIONS ON TIPS

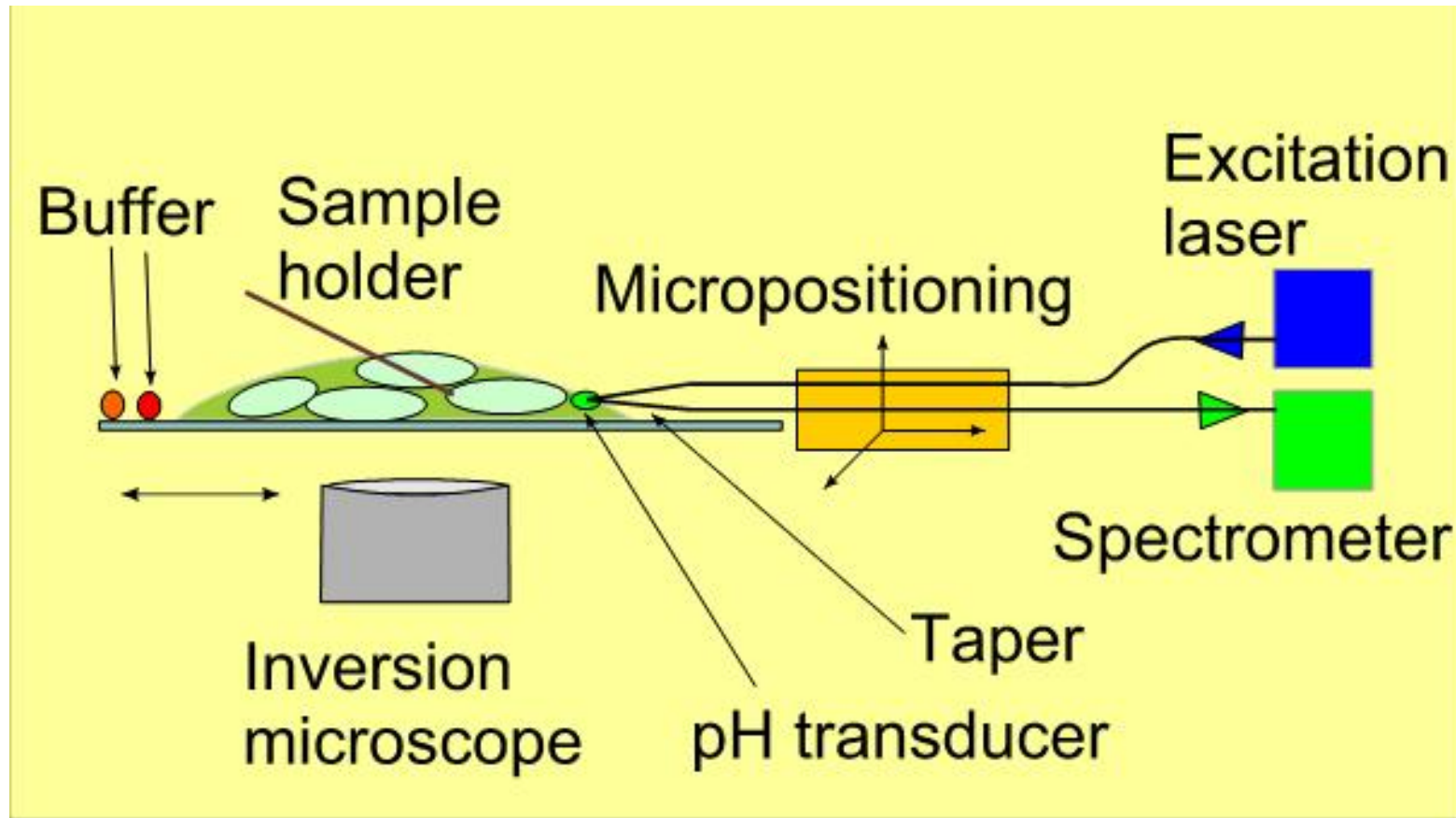
## Dip Coating



## Capillary effects

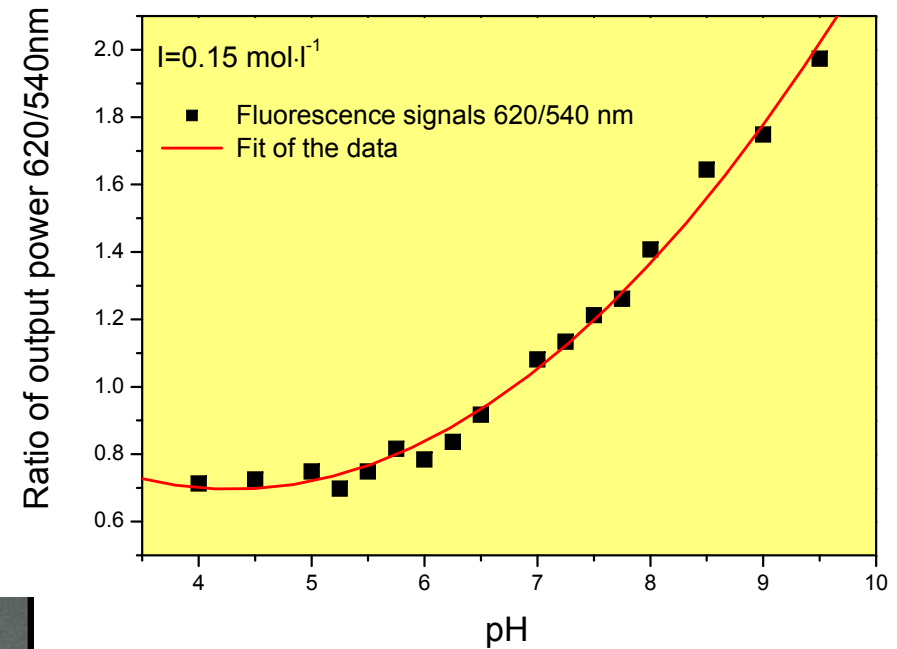
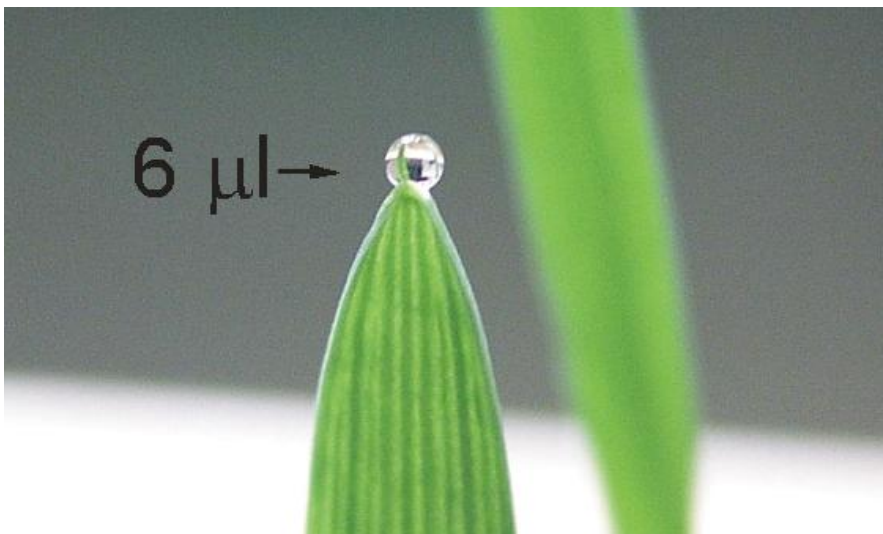


# SCHEME OF SET-UP IPE



# MEASUREMENTS

Change of fluorescence intensity of transducers due to pH changes



**Drops of exudate from tobacco leaves**

# pH MEASUREMENTS RESULTS

	Exudate from leaf tip	Exudate from a leaf center	Exudate from leaf basis
Average	5.0	5.6	5.5
St. deviation	0.3	0.3	0.1
Electrochem.	5.4	5.4	6.0

No pH gradient through leaves

I. Kašík et al., *Anal. Bioanal. Chem.* 398 (2010) 1883-1889

# CONCLUSIONS

- **Several original approaches** for increasing the **detection sensitivity** of multimode PCS (PCG) or plastic fibers to chemical developed in IPE.
- Optical fiber sensors can be used for detection in small volumes (cells), in remote or hardly accessible places
- They can be used for distributed detection - useful for monitoring integrity of buildings, bridges, airplanes, preventing leakages of dangerous substances into environment