

Changes of MicroRNA-1, -15b and -21 Levels in Irradiated Rat Hearts After Treatment With Potentially Radioprotective Drugs

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Summary

The aim of this study was to measure expression levels of microRNAs (miRNAs) (miRNA-1, -15b and -21) in the rat myocardium after a single dose of ionizing radiation (6-7 Gy/min, total 25 Gy). The rats were treated with selected drugs (Atorvastatin, acetylsalicylic acid (ASA), Tadalafil, Enbrel) for six weeks after irradiation. MiRNAs levels were measured by RT-qPCR. Irradiation down-regulated miRNA-1 in irradiated hearts. In Tadalafil- and Atorvastatin-treated groups, miRNA-1 expression levels were further decreased compared with irradiated controls. However, Enbrel increased miRNA-1 level in irradiated hearts similarly to that in non-irradiated untreated group. Increase of miRNA-15b is pro-apoptotic in relationship with ischemia. Irradiation caused down-regulation of miRNA-15b. Administration of ASA in the irradiated group resulted in the increase of miRNA-15b expression compared to non-treated controls without irradiation. After Enbrel administration, miRNA-15b levels were overexpressed compared to non-treated normal group. MiRNA-21 belongs to the most markedly up-regulated miRNAs in response to cardiogenic stress. MiRNA-21 was increased nearly 2-fold compared to non-treated hearts whereas Tadalafil reduced miRNA-21 levels (about 40 %). Our study suggests that Enbrel and Tadalafil changed miRNAs expression values of the irradiated rats to the values of non-irradiated controls, thus they might be helpful in mitigation of radiation-induced toxicity.

Key words

Irradiation • Toxicity • MicroRNA • qPCR • Heart • Treatment

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Introduction

Radiotherapy in the mediastinum area may often involve incidental exposure of the heart to ionizing radiation. In fact, epidemiological evidence has established a clear link between cardiovascular disease and exposure of the heart and major vessels to radiation doses (e.g. radiation therapy) (Darby *et al.* 2013).

Ionizing radiation may cause severe cellular damage both, directly by energetic disruption of DNA integrity, and indirectly, as a result of the formation of intracellular free radicals (Slezak *et al.* 2015). Radiation exposure consequences if are not intensive enough, may require longer period of time to develop (Donnelly *et al.* 2010).

MicroRNAs (miRNAs) are small, non-coding RNA molecules approximately 22 nucleotides in length which act as post-transcriptional regulators of gene expression. Each miRNA regulates dozens to hundreds of distinct target genes. Thus, miRNAs are estimated to regulate the expression of more than a third of human protein-coding genes (Lewis *et al.* 2005). To date, more than 2,500 miRNAs have been reported in humans (<http://www.miRbase.org/>). They play a crucial role in diverse cellular pathways. Many studies have suggested that miRNAs play a significant role in post-

transcriptional gene regulation in irradiated cells (Templin *et al.* 2012, Lee *et al.* 2014). Moreover, miRNAs show different expression patterns in the normal and diseased heart and some data indicate that miRNA expression may represent an efficient diagnostic marker of the heart disease (van Rooij *et al.* 2006, Li *et al.* 2012).

The current strategy to prevent radiation-induced diseases is mostly based on administration of treatments with mitigation effects. Acetylsalicylic acid (ASA), well-known as aspirin, is non-selective inhibitor of cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2), which prevents formation of pro-inflammatory prostaglandins and thromboxanes, thereby preventing clotting and formation of microthromboses (De Caterina *et al.* 1985, Perrone *et al.* 2010). Since formation of microthromboses was observed in irradiated rats (Slezak *et al.* 2014), acetylsalicylic acid can be useful as a possible drug for mitigation of radiation side effects. Atorvastatin (Lipitor) is widely used to inhibit 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMG-CoA) to decrease LDL. In many recent studies it was shown that Atorvastatin has also anti-inflammatory and antioxidative effects (Zhao *et al.* 2005). Important is it has positive effect on endothelial function increasing expression of endothelial nitric oxide synthase (eNOS) and increasing bioavailability of nitric oxide (NO) (Laufs *et al.* 2000, Lefer *et al.* 2001, Treasure *et al.* 1995). Tadalafil (Cialis) is the inhibitor of phosphodiesterase 5 (PDE5 inhibitor) and beside the effect on erectile dysfunction it is used for treatment of pulmonary arterial hypertension (like sildenafil). Tadalafil prevents the degradation of cGMP by PDE5 what leads to binding nitric oxide (NO) to guanylate cyclase receptors resulting in increased levels of cGMP and in the vasodilatation (Webb *et al.* 1999). Tadalafil can be used for its vasodilatation effect in the lungs, because some of the applied radiation dose is absorbed by lungs too. This causes constriction of blood vessels and lung fibrosis. Enbrel (etanercept) is an inhibitor of tumor necrosis factor alpha (TNF- α). After irradiation inflammatory reactions develop in the organisms (Khan *et al.* 2013). It is supposed that inflammation arises from activation of TNF- α , so its inhibition can be very useful in the treatment of radiation-induced disease.

In the present study, we measured the expression levels of miRNA-1, miRNA-15b and miRNA-21 in the irradiated rat myocardium exposed to a single dose of 25 Gy. The main goal of our study was to examine whether expression of assessed microRNAs will be affected in the hearts of the irradiated rats after

administration of potentially radioprotective drugs (acetylsalicylic acid, Atorvastatin, Tadalafil and Enbrel).

Methods

Experimental models

The experiments on animals were performed in accordance with the Ethics Committee of the Institute for Heart Research SAS and protocols approved by the State Veterinary and Food Administration of the Slovak Republic. We used 12 weeks old male Wistar rats (250 g weight) that were purchased from Department of Toxicology and Laboratory Animals Breeding, Slovak Academy of Sciences, Slovakia. All animals were housed under standard environmental conditions and maintained on commercial rat chow and tap water *ad libitum*.

Forty rats were randomly divided into two groups, irradiated (IR, n=25) and non-irradiated (C, n=15). Rats from IR group were anesthetized and irradiated by a single dose of ionizing radiation with a total dose 25 Gy in 6-7 Gy/min ratio to the mediastinum area using 5 MeV/1 kW electron linear accelerator UELR 5-1S equipped with tungsten converter to X-rays (NIIIEFA St. Petersburg, RF).

Both groups were given specific symptomatic treatment for mitigation of a negative impact of irradiation – Atorvastatin (ATOR, 1 mg/kg), acetylsalicylic acid (Aspirin – ASA, 12.5 mg/kg), Tadalafil (Cialis – TAD, 0.25 mg/kg) and Etanercept (Enbrel – E, 312.5 μ g/kg) – for six weeks after irradiation. Doses of drugs were calculated from the maximal therapeutic dose for humans in relation to the rat weights. Six weeks after irradiation animals were sacrificed. Hearts were rapidly removed, divided into atria, ventricles and septum, placed in the liquid nitrogen and stored at -80 °C. Left ventricular tissue was used for the miRNAs assessments.

Quantitative real-time PCR

The total RNA was extracted by miRNA mini kit (QIAGEN Sciences, MD, USA). Concentration of isolated total RNA was measured by Nanodrop ND-1000 spectrophotometer (Agilent technologies, CA, USA). Ten (10 ng) of the total RNA was reverse transcribed to cDNA with TaqMan® MicroRNA Reverse Transcription Kit (Applied Biosystems, Carlsbad, CA, USA), cDNA was amplified using the TaqMan® Universal MasterMix II (Applied Biosystems, Carlsbad, CA, USA) on Roche Light cycler 480 II (Roche Applied Science, IN, USA). RT-qPCR was performed using specific

TaqMan MicroRNA assays (Applied Biosystems, Carlsbad, CA, USA) for detection of miRNA-1 (assay ID 002064), miRNA-15b (assay ID 000390) and miRNA-21 (assay ID 000397) according to the manufacturer's instructions. Amplification data were normalized to U6 small RNA (assay ID 001973).

Statistical analysis

All values of experiments were expressed as means \pm SD. Statistical analysis was performed using one-way two-tailed ANOVA test, followed by Bonferroni *post-hoc* analysis (GraphPad Prism 7). A probability value less than 0.05 was considered to reflect statistically significant differences.

Table 1. Biometrical parameters of experimental rats.

| | Body weight (g) | Heart weight (g) | Systolic blood pressure (mm Hg) |
|-------------------------|--------------------|---------------------|------------------------------------|
| <i>Control (n=3)</i> | 397 \pm 13 | 0.94 \pm 0.06 | 90.38 \pm 32.6 |
| <i>C ATOR (n=5)</i> | 409.67 \pm 38 | 1.02 \pm 0.07 | 105.85 \pm 8.5 |
| <i>C ASA (n=5)</i> | 396 \pm 3 | 1.04 \pm 0.10 | 110.68 \pm 7.8 |
| <i>C TAD (n=5)</i> | 415.67 \pm 31.5 | 1.00 \pm 0.06 | 92.69 \pm 19.4 |
| <i>C E (n=5)</i> | 382.33 \pm 13.5 | 1.05 \pm 0.09 | 113.24 \pm 11.7 |
| <i>Irradiated (n=3)</i> | 322.20 \pm 31.3 | 1.26 \pm 0.15 | 76.44 \pm 8.4 |
| <i>IR ATOR (n=5)</i> | 342 \pm 27.7 | 1.26 \pm 0.06 | 98.43 \pm 19.3 |
| <i>IR ASA (n=5)</i> | 335.60 \pm 17.08 | 1.27 \pm 0.13 | 103.09 \pm 9.3 |
| <i>IR TAD (n=5)</i> | 321.80 \pm 25.07 | 1.27 \pm 0.13 | 74.90 \pm 8.3 |
| <i>IR E (n=5)</i> | 379.60 \pm 33.6 | 1.21 \pm 0.19 | 96.55 \pm 29.7 |

Data are expressed as mean \pm SD. Control – rats without irradiation (n=3), C ATOR – rats treated with atorvastatin without irradiation (n=3), C ASA – rats treated with acetylsalicylic acid without irradiation (n=3), C TAD – rats treated with Tadalafil without irradiation (n=3), C E – rats treated with Enbrel without irradiation (n=3), Irradiated – rats irradiated with 25 Gy (n=5), IR ATOR – irradiated rats treated with atorvastatin (n=5), IR ASA – irradiated rats treated with acetylsalicylic acid (n=5), IR TAD – irradiated rats treated with Tadalafil (n=5), IR E – irradiated rats treated with Enbrel (n=5).

MiRNA-1

Expression of miRNA-1 was measured in rat left ventricles (LV) six weeks after exposure to ionizing radiation on mediastinum (Fig. 1). Irradiation caused a decrease in miRNA-1 in LV (1.605 \pm 0.2206 in irradiated animals, P=0.1165 and 2.740 \pm 0.7375 in non-irradiated control hearts). Treatment with ASA decreased miRNA-1 levels in irradiated hearts (1.830 \pm 0.07024) to the same level as in irradiated rats without treatment. Treatment with ATOR and TAD further decreased miRNA levels (ATOR C=2.290 \pm 0.7823, ATOR IR=1.478 \pm 0.1353, P=0.2241 and TAD C=2.593 \pm 0.09684, TAD IR=1.140 \pm 0.03979, P=0.0001). However,

Results

Biometric parameters of experimental rats were measured six weeks after irradiation. The data are shown in Table 1. Irradiation decreased the body weight of rats after six weeks. A similar trend was seen in the measured blood pressure. Irradiated rats revealed lower blood pressure values than the non-irradiated ones.

Selected drugs (ATOR, ASA, TAD, E) were tested to alleviate the negative impact of radiation on rat hearts. In addition, we monitored the changes in the expression of miRNAs (miRNA-1, miRNA-15b and miRNA-21) as follows.

E increased miRNA-1 level in irradiated hearts to the level of non-irradiated untreated group (2.094 \pm 0.6295 in IR E group, P=0.8611 and 1.605 \pm 0.2206 in irradiated control hearts, P=0.1165).

MiRNA-15b

The expression of miRNA-15b was higher in control hearts (0.1678 \pm 0.04371) as compared to the irradiated groups (0.1460 \pm 0.01806, P=0.6121) (Fig. 2). Treatment with ATOR had no significant effect on the expression levels of miRNA-15b as compared to non-treated irradiated controls (0.1213 \pm 0.01037 in IR ATOR group, P=0.5592 and 0.1460 \pm 0.01806, P=0.6121 in

IR C). On the other hand, administration of ASA had opposite effect on miRNA-15b expression levels (ASA=0.1200±0.002517 in C, 0.1615±0.03175 after irradiation, P=0.3654). TAD group had comparable miRNA-15b expression levels in irradiated group (TAD=0.1300±0.001528 in C, 0.1438±0.04934 after

irradiation, P=0.8403) compared with non-treated irradiated group. Expression level of miRNA-15b in irradiated E group was increased more than in all other groups (E=0.1317±0.03963 in C, 0.1900±0.04926 after irradiation, P=0.4474).

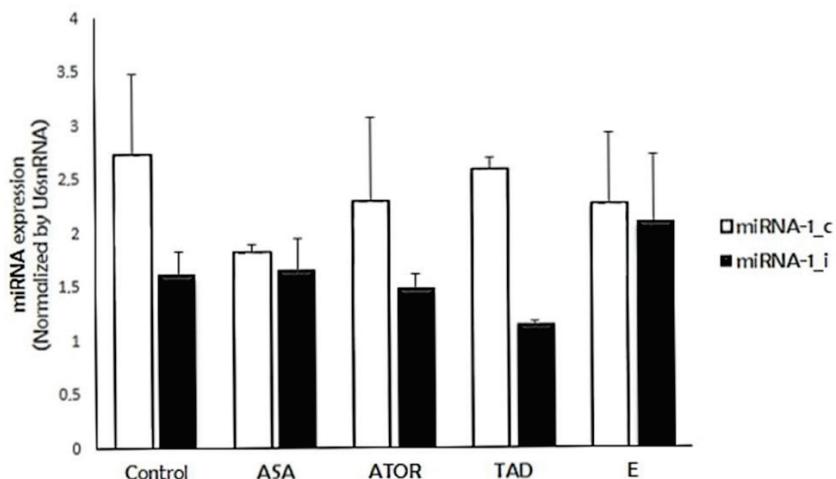


Fig. 1. MiRNA-1 expression levels in irradiated rat myocardium after treatment with acetylsalicylic acid (ASA), atorvastatin (ATOR), Tadalafil (TAD) and Enbrel (E) were expressed as mean ± SD. White columns – non-irradiated (n=3), dark columns – irradiated (n=5).

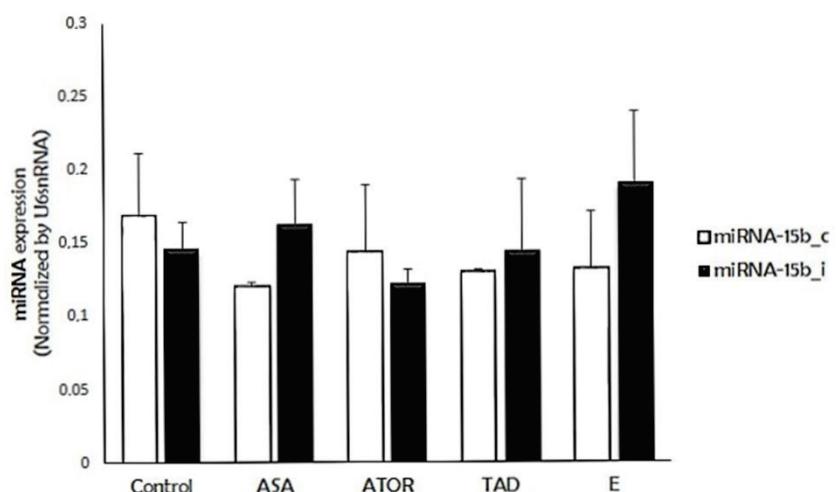


Fig. 2. MIRNA-15b expression levels in irradiated rat myocardium after treatment with acetylsalicylic acid (ASA), atorvastatin (ATOR), Tadalafil (TAD) and Enbrel (E) were expressed as mean ± SD. White columns – non-irradiated (n=3), dark columns – irradiated (n=5).

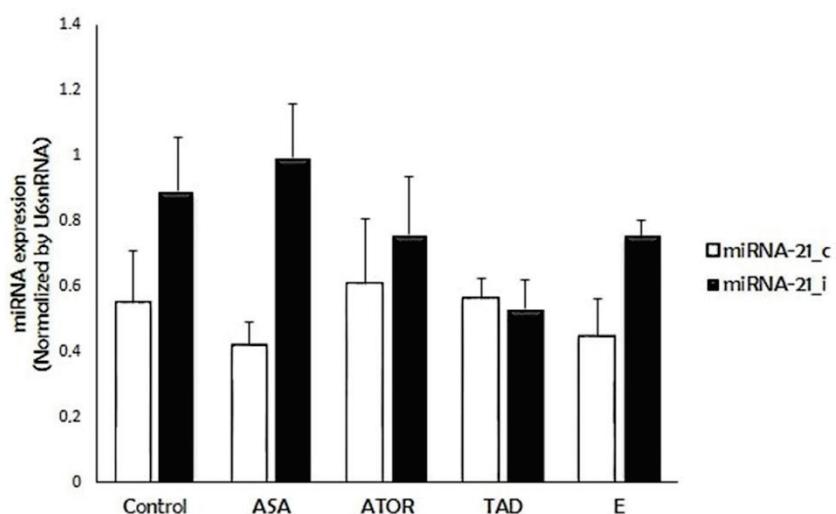


Fig. 3. miRNA-21 expression levels in irradiated rat myocardium after treatment with acetylsalicylic acid (ASA), atorvastatin (ATOR), Tadalafil (TAD) and Enbrel (E) were expressed as mean ± SD. White columns – non-irradiated (n=3), dark columns – irradiated (n=5).

MiRNA-21

Irradiation caused up-regulation of miRNA-21 in non-treated irradiated hearts (0.8915 ± 0.1645 IR group, 0.5473 ± 0.1616 C group, $P=0.1928$) (Fig. 3). The most effective, according to miRNA-21 expression levels, was administration of TAD (0.8915 ± 0.1645 IR control group, $P=0.1928$ and 0.5285 ± 0.09128 ATOR IR, $P=0.7992$). Treatment with ATOR had slight effect in increasing miRNA-21 expression (0.6043 ± 0.2032 in C group and 0.7578 ± 0.1793 after irradiation, $P=0.6051$). Similarly, Enbrel slightly increased miRNA-21 (0.4480 ± 0.1159 in C group and 0.7544 ± 0.04648 after irradiation, $P=0.0266$). Treatment with ASA elevated miRNA-21 expression after irradiation (0.4187 ± 0.07044 in C group and 0.9926 ± 0.1683 after irradiation, $P=0.047$).

Discussion

This work was focused on the changes in the expression of miRNAs (miRNA-1, -15b and -21) in rat hearts after 25 Gy irradiation. The dose of 25 Gy was selected according to our previous screening results (not published). We wanted to achieve the maximum damage of the heart with minimal mortality in our experiments. Cytotoxicity, oxidative stress, inflammation, fibrosis and the cardiac hypertrophy represent common consequences of radiation exposure (Pathak *et al.* 2015, Slezak *et al.* 2015).

Recently, various studies showed that radiation can modulate the expression levels of the miRNAs (Chaudhry 2014, Wang *et al.* 2015). MiRNAs have an important function in remodeling of the heart tissue in adaptation to stressors (Romaine *et al.* 2015). In this study, miRNAs were used as plausible biomarkers of radiation damage and repair. MiRNAs were selected due to their known association with cardiac hypertrophy (miRNA-1), apoptosis (miRNA-15b, -21), fibrosis (miRNA-21) (Zhu and Fan 2012) and oxidative stress (miRNA-15b, -21) (Simone *et al.* 2009). Used miRNAs were selected based on its functions in many cardiovascular diseases and its connection with pathological processes evolving in the heart after irradiation (see results of previous experiments) (Slezak *et al.* 2013, 2014).

In the present study, we evaluated the changes in miRNAs expression levels in irradiated rat heart before and after administration of drugs that might be involved in symptomatology of radiation injury. In normal and

irradiated rat LV, the expression of the above mentioned miRNAs was measured after six weeks of treatment. This time period was chosen as the time when all the changes are manifested and are maximally expressed. We used selected drugs with possible mitigating effect (acetylsalicylic acid – Aspirin (ASA), Atorvastatin – Lipitor (ATOR), Tadalafil – TAD and Etanercept – Enbrel (E)). These drugs were selected mainly due to their involvement in antioxidative, antithrombotic, anti-inflammatory and cardioprotective pathways of cardiomyocytes and endothelium that is important in suppressing of the negative radiation effects.

MiRNA-1 expression levels

MiRNA-1 belongs to the most abundant miRNAs in normal rat heart tissue. Available studies showed that miRNA-1 was frequently down-regulated in various types of cardiac diseases (Duan *et al.* 2014). Our results showed that exposure to the radiation (single dose of 25 Gy) decreased miRNA-1. Similarly Ikeda *et al.* (2007) reported that miRNA-1 was down-regulated in dilated cardiomyopathy and aortic stenosis. A decreased expression of miRNA-1 was observed in several models of cardiac hypertrophy and heart failure (Ikeda *et al.* 2009, Sayed *et al.* 2007). These results were also confirmed in humans. Seronde *et al.* (2015) found that levels of miRNA-1 were significantly lower in acute heart failure and chronic heart failure patients compared to healthy subjects.

The expression of miRNA-1 in the irradiated heart after the administration of ASA, ATOR, TAD and E is currently unknown. Our results show that treatment with ASA, ATOR and TAD had no protective effect on the heart exposed to irradiation. On the other hand, treatment with E increased miRNA-1 value in irradiated hearts to the level of non-irradiated controls. Thus E appears to be the most effective drug in restoring the miRNA-1 as compared to other administered drugs. These results suggest that inhibition of TNF- α inflammatory pathway with E might be very effective in irradiation injury which is supported by the fact that TNF- α is activated by calmodulin (Wang *et al.* 2012) and it is inversely correlated with miRNA-1 expression (Ikeda *et al.* 2009). These results indicate that calcium plays a critical role in the myocardial hypertrophy induced by TNF- α , suggesting that the TNF- α -induced hypertrophic response may be linked to Ca^{2+} homeostasis (Wang *et al.* 2012).

MiRNA-15b expression levels

MiRNA-15b has its role in the cardiac development and disease (Porrello 2013). Moreover, it has been reported that the expression of miRNA-15b may be altered following exposure to various genotoxic stressors including radiation (Simone *et al.* 2009). In the present study, we observed that miRNA-15b was down-regulated in irradiated rat hearts as compared to non-irradiated ones. Similarly, Guo *et al.* (2009) reported that miRNA-15b was down-regulated in rat activated hepatic stellate cells and which correlated to apoptosis by targeting Bcl-2. However, most of the studies revealed opposite effect of miRNA-15b. Up-regulation of the miRNA-15 family was observed following cardiac ischemia and heart failure (Latronico *et al.* 2007). In another study, Tijssen *et al.* (2014) showed up-regulation of miRNAs from miRNA-15 family in the overloaded heart. It was suggested that miRNA-15 family can inhibit the TGF β -pathway which is important in the development of cardiac fibrosis and hypertrophy. Quantitative real time-PCR data showed that the expression of miRNA-15a and miRNA-15b was up-regulated in mice hearts subjected to ischemia/reperfusion and overexpression of miRNA-15b enhanced cell apoptosis (Liu *et al.* 2012). Rahman *et al.* (2014) identified miRNA-15b up-regulation in a dose- and time-dependent manner in human bronchial epithelial cells (HBEC).

To our knowledge, the expression of miRNA-15b in the irradiated heart after the treatment with ASA, ATOR, TAD and E has not been reported yet. Treatment with ASA and E caused up-regulation of miRNA-15b in irradiated rat ventricles to the level of non-irradiated non-treated hearts. This indicates that ASA and E may protect the irradiated heart from fibrosis and hypertrophy, probably through their anti-inflammatory actions. In irradiated groups, administration of ATOR and TAD had no significant effects on miRNA-15b expression levels. These results suggest that inflammatory pathways are important in mitigating the cardiac fibrosis and hypertrophy. Protective effects of ASA and E can also be mediated through targeting Bcl-2 gene expression, because ASA (Ding *et al.* 2014) and E (Malaviya *et al.* 2006) are pro-apoptotic as well as miRNA-15b, so they can alleviate hypertrophy and fibrosis (Slezak *et al.* 2015).

MiRNA-21 expression levels

Dong *et al.* (2014) found that miRNA-21

expression was significantly higher in cardiac fibroblasts than in cardiomyocytes. Cardiac fibrosis is an important factor involved in the development of heart insufficiency and miRNA-21 is highly expressed in fibrotic cardiac muscles (Thum *et al.* 2008). In the present study, we showed that miRNA-21 levels were up-regulated in the irradiated hearts compared to the non-irradiated controls. These results are consistent with the finding by Valkov *et al.* (2015). MiRNA-21, a well-studied miRNA in cardiac fibroblasts and fibrosis development, was increased 3.1-fold upon fibroblast activation. Halimi *et al.* (2014) found out that miRNA-21 level before radiotherapy was comparable with that in healthy volunteers ($P=0.10$) and increased significantly after radiotherapy ($P<0.001$). MiRNA-21 was shown to be altered under conditions of various oxidative stressors including radiation (Simone *et al.* 2009). Viczenczova *et al.* (2016) observed increased levels of miRNA-21 and total Cx43 protein expression in rat hearts after chest irradiation.

MiRNA-21 levels were not previously measured in irradiated rat hearts after administration of ASA, TAD, ATOR and E. Based on the obtained results, the most protective effect proved to be exerted by TAD, because it lowered miRNA-21 level to its value in untreated non-irradiated heart. Slight mitigating effect was also shown by ATOR and E.

Conclusion

MiRNA molecules are considered to be important prognostic and therapeutic factors in many diseases, including radiation-induced heart disease or heart failure. They are necessary in post-transcriptional regulation of gene expression and also in communication among cells. Based on miRNA expression levels, our results suggest that among the selected drugs, particularly Tadalafil and Enbrel changed miRNA expression values of the irradiated rats to the values of non-irradiated controls. Referring to these results we can suppose that Tadalafil and Enbrel might be helpful in mitigation of damaging effects of irradiation.

Conflict of Interest

There is no conflict of interest.

Acknowledgements

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