

Blood Vessel Replacement: 50 years of Development and Tissue Engineering Paradigms in Vascular Surgery

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Summary

The gold standard material in bypass surgery of blood vessels remains the patient's own artery or vein. However, this material may be unavailable, or may suffer vein graft disease. Currently available vascular prostheses, namely polyethylene terephthalate (PET, Dacron) and expanded polytetrafluoroethylene (ePTFE), perform well as large-caliber replacements, but their long-term patency is discouraging in small-caliber applications (<6 mm), such as in coronary, crural or microvessel surgery. This failure is mainly a result of an unfavorable healing process with surface thrombogenicity, due to lack of endothelial cells and anastomotic intimal hyperplasia caused by hemodynamic disturbances. An ideal small-diameter vascular graft has become a major focus of research. Novel biomaterials have been manufactured, and tissue-biomaterial interactions have been optimized. Tissue engineering technology has proven that the concept of partially or totally living blood vessels is feasible. The purpose of this review is to outline the vascular graft materials that are currently being implanted, taking into account cell-biomaterial physiology, tissue engineering approaches and the collective achievements of the authors.

Key words

Small-caliber vascular grafts • Synthetic polymers • Biomaterials • Tissue engineering • Stem cells • Dynamic bioreactor • Shear stress

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Introduction

Atherosclerosis accounts for almost one half of all deaths in Europe (Stehouwer *et al.* 2009). Although advanced pharmacological and minimally-invasive techniques offer a growing therapy option (Met *et al.* 2008), a surgical bypass of blood vessels on the heart or on a lower extremity remains the procedure of choice in a number of patients (Fig. 1) (Guyton 2006, Norgren *et al.* 2007). This approach is also more cost-effective, and in particular preserves the quality of the patient's life better than primary amputation of a limb (Cheshire *et al.* 1992). A synthetic tube or a vascular prosthesis has to be implanted when the patient's own artery or vein is not available. After more than half a century of development work, the results achieved with currently available materials are not optimal in terms of healing and tissue regeneration.

Long-term patency rates of prosthetic grafts are satisfactory in large-caliber arteries (>8 mm), where thrombogenicity (i.e. disposition to blood coagulation) may be overcome by massive blood flow, and the 5-year patency of aorto-iliac substitutes is 90 % (Brewster 1997). There is little difference between the results for prosthetic and autogenous (i.e. patient's own) material in medium-caliber replacements (6-8 mm), e.g. in carotid or common femoral arteries (Ricotta 2005). However, in small-caliber vessels (<6 mm), such as coronary arteries (heart), infrainguinal arteries (below the inguinal ligament), and particularly in low-flow infrageniculate

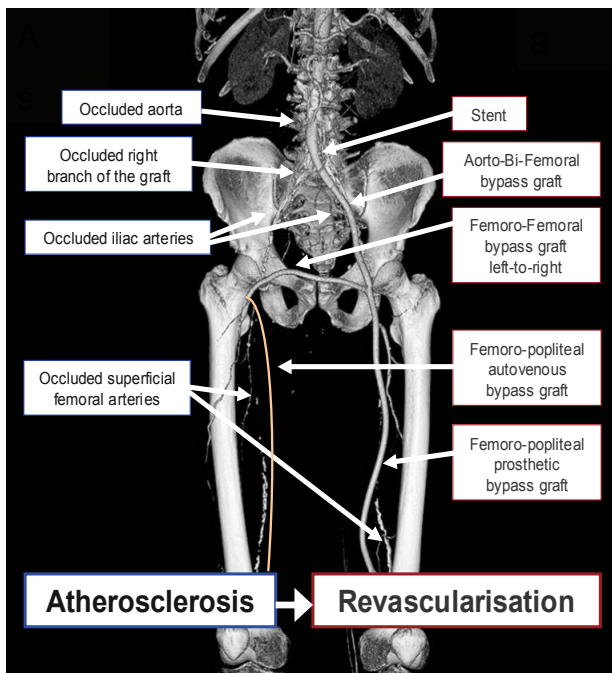


Fig. 1. Surgical repair of multilevel peripheral atherosclerotic disease.

arteries (below the knee joint), the outcomes of vascular prostheses are rather disappointing. Five-year primary patency of prosthetic (ePTFE) above-the-knee femoro-popliteal bypass grafts is as low as 39 %, whereas autovenous bypasses (i.e. performed with own vein) have a rate of 74 % (Klinkert *et al.* 2004). A summary of currently-used vascular replacements is shown in Table 1.

Currently available vascular grafts fail due to the thrombogenicity of the artificial surface and intimal hyperplasia (IH), which is located at distal anastomosis (Fig. 2) of prosthetic grafts (i.e. the site of the junction to the artery). Unlike animal models, most of the prosthetic blood-contacting surfaces remain uncovered by tissue in humans (Berger *et al.* 1972). The etiology of IH, developing usually 2-24 months post implantation, is multifactorial and includes a compliance mismatch between a relatively rigid prosthesis and the more elastic native artery (Sarkar *et al.* 2006), graft/artery diameter mismatch, lack of endothelial cells (EC), surgical trauma and flow disturbances resulting in adaptive changes in the sub-endothelial tissue, characterized by proliferation and migration of vascular smooth muscle cells (VSMC) from media to intima, and synthesis of extracellular matrix (ECM) proteins (Haruguchi and Teraoka 2003).

To overcome these fundamental inconveniences, novel biomaterials research (Shin and Mikos 2003) and particularly tissue engineering modalities are increasingly being adopted (Isenberg *et al.* 2006).

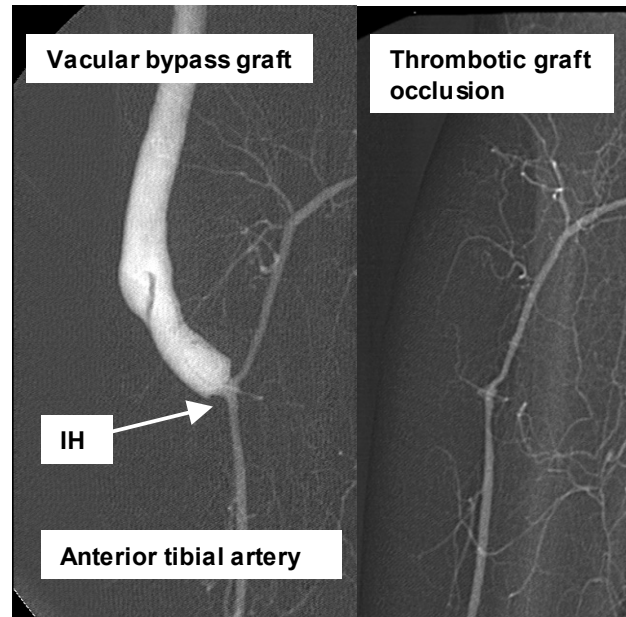


Fig. 2. Intimal hyperplasia (IH) in distal anastomosis of a vascular graft, causing thrombosis and occlusion.

Promising and clinically proven outcomes have been achieved with biohybrid and tissue-engineered vascular grafts.

An ideal vascular graft would possess the following characteristics: mechanical strength and compliance to withstand long-term hemodynamic stresses; non-toxicity; non-immunogenicity; biocompatibility; “off-the-shelf” availability in various sizes for emergency care; operative suturability and simplicity of surgical handling; resistance to *in vivo* thrombosis; ability to withstand infection; complete incorporation into the host tissue with satisfactory graft healing and ability to grow when placed in children (Kakisis *et al.* 2005), and, last but not least, reasonable manufacturing costs. The task is so demanding and the potential rewards are so great that research in the field of small-caliber vascular substitutes has been compared with the search for the Holy Grail (Conte 1998).

The aim of this review is to outline the characteristics and drawbacks of currently-used vascular substitutes, with special emphasis on the physiological cellular response to materials and on vascular tissue engineering approaches, based on the collective experience of the authors.

Historical background

The history of vascular surgery tracks progress made in suturing and replacing damaged vessels. In

Table 1. Vascular substitutes in clinical use according to body region.

Vascular substitute choice	Vascular regions				
	Large-caliber arteries (≥ 8 mm)	Medium-caliber arteries (6-8 mm)	Small-caliber arteries (≤ 6 mm)	Venous reconstructions	Hemodialysis arterio-venous access
	Aorta, arch vessels, iliac and common femoral arteries	Carotid, subclavian, common femoral, visceral and above-the-knee arteries	Coronary, below-the-knee, tibial and peroneal arteries	Superior and inferior vena cava, ilio-femoral veins, portal vein, visceral veins	Upper > lower extremity
1 st choice	Prosthesis (Dacron, ePTFE)	Prosthesis or autograft (equal)	Arterial or venous autograft	Saphenous spiral vein graft, deep venous autograft	Native material
2 nd choice	Allograft, deep venous autograft	Prosthesis or autograft	Composite graft, vein interposition, prosthesis (ePTFE, Dacron), allograft, biosynthetic	Allografts, ePTFE, Dacron, biografts	ePTFE, PU, xenografts, biografts, TEBV (clinical trial)

ePTFE (expanded polytetrafluoroethylene), PU (polyurethane), TEBV (totally-engineered blood vessels).

ancient times, vascular interventions were limited to compressing and cauterizing injured vessels to control bleeding. Ambrose Paré developed a vessel ligation in the 15th century, but surgical repair of vessels did not begin until 1759, when Hallowell and Lambert repaired a brachial artery injury with a suture. The first vascular anastomosis (connection of blood vessels) was performed by Nicholas Eck as a porto-caval shunt in dogs in 1877 (Starzl 2003). Carrel and Guthrie optimized the vascular anastomosis transplantation technique and even tissue culture for organ replacement in the early 1900s, and Alexis Carrel received the Nobel Prize for Physiology or Medicine in 1912.

Goyannes first used an autogenous popliteal vein graft for popliteal aneurysm repair in 1906. A femoro-popliteal bypass with a reversed saphenous vein graft was first performed by Kunlin in 1948, initiating a very successful era for this type of graft that has lasted until the present day (Lopez and Ginzberg 2008). At the same time, the first fresh arterial allografts (foreign tissue of the same species) began to be used in human vascular reconstructive surgery (Gross *et al.* 1948). An artificial vascular prosthesis was first implemented as an aortic replacement with a Vinyon "N" tube in a dog experiment (Voorhees *et al.* 1952). The same material was implanted in humans to replace an aneurysm (dilation) of the abdominal aorta (Blakemore and Voorhees 1954), leading to rapid progress in vascular surgery and prostheses research and use (Sauvage 1986).

Arterial replacements

Biological vascular grafts

The gold standard for vascular replacement remains the autologous native vessel, which possesses the most physiological properties. In coronary artery bypass grafting (CABG), the internal mammary artery and the radial artery are superior to a greater saphenous vein graft (SVG), which is also often used (Beghi *et al.* 2002). In the case of lower limb or peripheral bypass surgery, the material of choice is also greater saphenous vein (*vena saphena magna*). It can be cleansed of valves and anastomosed *in situ*, so that the *vasa vasorum* remain intact (Kachlik *et al.* 2007) and the blood flow direction is reversed. In this type of graft, preservation of desired endothelial properties can be expected; however, several studies have failed to confirm superiority to the more widespread *ex vivo* reversed setting of the SVG (Lawson *et al.* 1999). It should be noted that, despite good clinical performance, SVG is also liable to atherosclerosis and intimal hyperplasia occurring throughout the length (Sarjeant and Rabinovitch 2002). In addition, almost 30-40 % of patients lack an appropriate saphenous vein (Faries *et al.* 2000) due to previous phlebitis, vessel removal, varicosities, hypoplasia or anatomical unsuitability. Additionally, any surgical vessel harvest is associated with indispensable donor site morbidity (Swenne *et al.* 2006).

Alternatives include the use of other native vascular materials only in carefully selected indications,

and sometimes with limited clinical performance: arteries, such as the right gastroepiploic artery for coronary application (Sasaki 2008), and veins, such as the lesser saphenous vein (*vena saphena parva*) (Chang *et al.* 1992), arm veins (Calligaro *et al.* 1997) for coronary and peripheral bypasses, or deep leg veins (Ali *et al.* 2009) for infected aortic graft replacement, visceral revascularization or even primary lower limb bypass.

Although the use of fresh (cold-stored) or cryopreserved homografts (i.e. human allografts from cadaver donors) was abandoned in the early 1960s because of difficulties in preserving them, late graft deterioration, aneurysm formation and the anticipated availability of synthetic prostheses, they have been reintroduced for managing aortic prosthetic graft infection (Kieffer *et al.* 2004), lower extremity primary revascularization (Dardik *et al.* 2002, Fahner *et al.* 2006, Matia *et al.* 2007) and simultaneous or sequential revascularization surgery in solid organ transplant-recipients (Matia *et al.* 2008). Although allografts are not routinely used (usually in limb-threatening situations, in redo surgery or in an infected field), some studies suggest improved patency of these grafts, e.g. in the case of preserved externally-supported human umbilical vein (harvested from newborns) when compared to ePTFE (Johnson and Lee 2000).

The application of heterografts (xenografts, i.e. tissue from different species) (Schmidt and Baier 2000) involves mainly studies on alternative hemodialysis vascular access. Although bovine carotid artery heterografts did not show superiority to PTFE (Hurt *et al.* 1983), and decellularized bovine ureteric grafts have been implanted with ambiguous results (Chemla and Morsy 2009), glutaraldehyde-crosslinked bovine mesenteric vein provided a considerable reduction in infection, thrombosis and reintervention rate (Katzman *et al.* 2005). Importantly, decellularized natural tissue of allogenic (human) or xenogenic (animal) origin serves as a scaffold for cell seeding within the scope of tissue engineering of vascular grafts (Dahl *et al.* 2003). A summary of biological vascular conduit materials is presented in Table 2.

Synthetic vascular grafts

For more than 50 years, two polymers have been used for synthetic vascular prostheses: 1) polyethylene terephthalate (PET), Terylene or Dacron, and 2) polytetrafluoroethylene (PTFE), Teflon or Gore-Tex. Both of these molecules are highly crystalline and

hydrophobic. Prosthetic rings or coils can be applied to the external surface of both materials to resist kinking and compression in anatomically required positions (long bypasses, grafts crossing the joint or body midline).

PET/Dacron (polyethylene terephthalate, [-O-C=O-C₆H₄-O-C=O-CH₂-CH₂-]) was introduced in England in 1939, and was further developed and patented as Dacron by DuPont in 1950. It is a thermoplastic polymer resin of the polyester family and is used in synthetic fibers of round cross-section. These fibers are bundled into multifilament yarns, which can be woven (over-and-under pattern) or knitted (looped fashion) into textile vascular graft fabrics and tubes. A crimping technique (an undulating surface) is sometimes used to increase distensibility and kink-resistance. The porosity of a textile Dacron graft is defined by water permeability, which is greater for knitted Dacron. Knitted Dacron is impregnated with albumin, collagen or gelatin to make it more impervious, to decrease the porosity/permeability, and to avoid the need for blood preclotting prior to implantation. Depending on the cross-linking agent (formaldehyde or glutaraldehyde), the albumin impregnation is degraded 2 to 8 weeks after implantation (Marois *et al.* 1996). Although collagen impregnation increased platelet deposition and delayed the healing process in a dog experiment (Guidoin *et al.* 1996), a prospective randomized trial with aorto-iliac prostheses indicated that collagen impregnation does not stimulate the coagulation cascade more than conventional Dacron (De Mol Van Otterloo *et al.* 1991). There are no differences in clinical graft patency between woven and knitted Dacron, when used as an aorto-iliac bypass graft (Quarmby *et al.* 1998). Dacron is reported to dilate over time, but direct association with graft complications and failure has been rare (Blumenberg *et al.* 1991).

Host reactions to the vascular prosthesis start immediately after restoration of blood circulation. The tissue-prosthesis and blood-prosthesis interfaces are complex microenvironments, and the physico-chemical properties of the surface of the prosthesis, such as charge, energy, wettability and roughness, may be responsible for the graft patency. The first step is the plasma protein adsorption/desorption process typical for any blood/material interface (Vroman and Adams 1969), followed by platelet deposition, white blood cell and erythrocyte adhesion, and eventually endothelial and smooth muscle cell migration. Fibrin deposits, which contain platelets and blood cells, build up during the first few hours to days after implantation. They are stabilized

Table 2. Biological vascular grafts in clinical use.

	Biological vascular grafts				
	Autografts		Allografts (homografts)		Xenografts (heterografts)
	Arterial	Venous	Arterial	Venous	
<i>Advantages</i>	Closest approximation, less diameter mismatch, internal mammary artery anatomically nearby, excellent function	Durable and versatile, good results, infection resistance, relative availability	Off the shelf availability, better resistance to infection, transplant-recipient patients		
<i>Disadvantages</i>	Availability, vasospasm (radial artery), donor site morbidity	Availability, harvest injury, vein graft disease	Antigenicity, graft deterioration, early occlusions, chronic rejection, intake of drugs, infection risk		
<i>Healing</i>	Intimal thickening, myointimal hyperplasia (radial artery)	Endothelial desquamation, vein dilation, wall thickening, arterialization, re-endothelialization	Endothelial denudation, immune response, fibrotization		
<i>First use</i>	Jaboulay and Briau 1896	Goyannes 1906	Gross <i>et al.</i> 1948	Linton 1955	
<i>Review e.g.</i>	Nezic <i>et al.</i> 2006	Cooper <i>et al.</i> 1996	Fahner <i>et al.</i> 2006	Dardik <i>et al.</i> 2002	Schmidt and Baier 2000

over a period of up to 18 months and form an inner compacted fibrin layer. Fibrin also fills the interstices within the graft wall. Generally, this fibrin/platelet pseudointima remains acellular; however, after 5 months, capillaries and fibroblasts can grow into the tight interstitial spaces even in humans (Stewart *et al.* 1975) and reach the inner fibrin layer. Only a few sparse small islands of endothelialization appeared in areas remote from the anastomosis region on woven excised Dacron grafts (Wu *et al.* 1995). External fibrin matrix surrounding the graft is gradually invaded by macrophages, and granulation tissue and foreign-body giant cells (FBGC) are usually seen under the external surrounding connective tissue capsule (Rahlf *et al.* 1986). In the case of knitted Dacron, endothelial islands can occasionally be observed on human explants during redo surgery or autopsy 1-11 years after implantation (Shi *et al.* 1997). Mature endothelium and a well-developed smooth muscle layer, described only in animal studies, is attributable to extended transanastomotic ingrowth rather than a midgraft healing process (Clowes *et al.* 1987, Zacharias *et al.* 1987).

The structure of knitted and, to a lesser extent, also woven Dacron allows a certain degree of transmural tissue ingrowth. However, the compacted inner fibrin layer forms a barrier and, even if the barrier is overcome

by undifferentiated connective tissue, the capillaries remain unconnected to the poorly endothelialized blood surface (Herring *et al.* 1979, Xue and Greisler 2003).

PTFE (polytetrafluoroethylene, [-CF₂-CF₂-]) was patented by DuPont as Teflon in 1937, and ePTFE was patented by Gore as Gore-Tex in 1969. ePTFE is an expanded polymer which is manufactured by a heating, stretching, and extruding process resulting in a non-textile porous tube composed of irregular-shaped solid membranes ("nodes"). The molecule is relatively biostable, i.e. less prone to deterioration in biological environments than PET (Guidoin *et al.* 1993a,b), and the graft surface is electronegative, which minimizes its reaction with blood components. It is characterized by a node-fibril structure, and its average porosity is described by the internodal distance (IND), which is usually 30 to 90 μm. However, the actual available ingrowth spaces between fibrils are much smaller than IND.

Optimal IND of 60 μm (high porosity) was experimentally proposed for tissue ingrowth and endothelialization of 4mm ePTFE grafts in a baboon model (Golden *et al.* 1990). Though a human trial with high-porosity ePTFE showed capillary ingrowth, it did not extend more than half the distance from the outside, and did not produce an endothelial lining (Kohler *et al.* 1992). Human host responses to standard low-porosity

ePTFE (IND $\leq 30 \mu\text{m}$) material are similar to the responses to Dacron grafts: a thin fibrin coagulum or amorphous platelet-rich material develops over time, and a lack of luminal surface endothelial cellular coverage is found after human implants. Collagenous external encapsulation develops within 1 to 6 months, penetrating the material structure with minimal cellular infiltration (Guidoin *et al.* 1993a). Systematic evaluation and meta-analysis of randomized controlled trials comparing Dacron and ePTFE showed no evidence of an advantage of one material over the other (Roll *et al.* 2008).

Polyurethanes (PU) comprise a large family of elastic polymers containing a urethane [-NH-(CO)-O-] group. Polyurethanes were originally developed in Germany in the 1930s. They were commercialized by DuPont in 1962, and have been available for biomedical applications since the 1960s (Boretos and Pierce 1967). Generally, they are copolymers consisting of three different monomers: crystalline (hard) and amorphous (soft) segments, the former accounting for rigidity and the latter for flexibility, which can be varied by the manufacturer. The third monomer serves as a chain extender (Zdrachala 1996). The disadvantage of the first generation polyester-based PU was hydrolytic biodegradation, which resulted in abortion of one of the clinical trials (Zhang *et al.* 1997). The next generation of polyether-based PU, which is hydrolysis-resistant but more susceptible to oxidation, underwent successful clinical assessment as hemodialysis access graft (Glickman *et al.* 2001), and received Food and Drug Administration (FDA) approval in 2000.

The latest generation of polycarbonate-based PU is hydrolytically and oxidatively stable, and promoted faster luminal endothelialization and less neointimal formation as small-calibre vascular prostheses in a rat experiment compared to ePTFE (Jeschke *et al.* 1999). After 36-month implantation in aorto-iliac position in dogs, a histological analysis of poly(carbonate-urea) PU grafts showed well-developed neointima only in distal anastomosis, only minor hydrolysis of the amorphous segments and, in addition, the grafts retained their compliance with time – a feature not observed in Dacron or ePTFE. These promising results led to a phase I clinical trial (Seifalian *et al.* 2003).

According to their microstructure, polyurethanes can be divided into fibrillar or foamy. Both structures tend to lack communicating spaces for potential capillary ingrowth. Upon implantation, the blood surface of the fibrillar PU is covered by a fibrin layer that is thinner

than on knitted Dacron, and the outer surface is encapsulated by scar formation containing FBGC. In microporous foamy PU with $15 \mu\text{m}$ pore size, relatively little capillary ingrowth can be accomplished, whereas with increasing pore size up to $157 \mu\text{m}$, the undesired inflammatory FBGC reaction could be diminished and capillary sprouting was allowed from the outside in a Chacma baboon model. However, before its completion, transmural ingrowth slowed down when reaching the dense inner fibrin layer that was “squeezed” into the pores from the inside (Zilla *et al.* 2007).

Although PU grafts possess many interesting features, e.g. the presence of EC under poor hemodynamic conditions, excellent healing, good surgical handling and low suture bleeding (Tiwari *et al.* 2002), further investigations are required before more recommendations can be made, because there is a lack of evidence for their use in human peripheral bypass surgery (Dereume *et al.* 1993). An outline of synthetic vascular prostheses is summarized in Table 3.

Vascular grafts with modified lumen

Several attempts have been adopted to suppress the thrombogenicity of synthetic material by affixing chemicals or anticoagulants to the graft lumen: early studies showed decreased platelet deposition on carbon-coated ePTFE grafts, but the overall patency rates were not improved (Kapfer *et al.* 2006). A heparin-bonded Dacron graft exhibited slightly better patency than an untreated ePTFE graft (Devine *et al.* 2004), and heparin-immobilized ePTFE provided better thromboresistance in humans (Bosiers *et al.* 2006). Applying a polyethylene glycol (PEG)-hirudin/iloprost coating to 4 mm ePTFE prostheses reduced IH and led to superior patency in a pig experiment (Heise *et al.* 2006). Heparin coating significantly reduced aortic graft thrombosis in rats both for ePTFE and for PU (Walpoth *et al.* 1998). A dipyridamole (anti-platelet drug) coating positively influenced the patency rate of a 5 mm PU vascular prosthesis in animal experiments (Aldenhoff *et al.* 2001). Fibroblast growth factor-1 and heparin affixation enhanced ePTFE graft capillarization and surface endothelialization without significant IH in a canine aorta model (Gray *et al.* 1994). Antithrombotic agents are of course routinely administered to patients with vascular disease, with an obvious influence on graft patency and overall cardiovascular mortality (Watson *et al.* 1999).

Interestingly, an effort has been made to increase the resistance of synthetic vascular grafts to infectious

Table 3. Synthetic vascular grafts in clinical use.

	Synthetic vascular grafts					
	PET (Dacron, Terylen)		ePTFE (Teflon, Gore-Tex)		Polyurethane	
	Woven	Knitted	Low-porosity (<30 µm IND)	High-porosity (>45 µm IND)	Fibrillar	Foamy
<i>Advantages</i>	Better stability, lower permeability and less bleeding	Greater porosity, tissue ingrowth and radial distensibility	Biostability, no dilation over time	Biostability, better cell ingrowth	Compliance, good hemo- and biocompatibility, less thrombogenicity	
<i>Disadvantages</i>	Reduced compliance and tissue incorporation, low porosity, fraying at edges, infection risk	Dilation over time, infection risk	Stitch bleeding, limited incorporation, infection risk, perigraft seroma formation	Late neointimal desquamation in 90 µm IND, infection risk	Biodegradation in first generation, infection risk, carcinogenic?	
<i>Healing</i>	Inner fibrinous capsule, outer collagenous capsule, scarce endothelial islands	Fibrin luminal coverage, very sporadic endothelium, transanastomotic endothelialization in animals	Luminal fibrin and platelet carpet, connective tissue capsule with foreign body giant cells, no transmural tissue ingrowth	Macrophages and polymorphonuclear invasion, capillary sprouting, fibroblast migration, certain angiogenesis, thicker neointima, endothelialization in animals	Thin inner fibrin layer, outside foreign body cells, limited ingrowth	Better ingrowth with bigger pores
<i>First use</i>	Ku <i>et al.</i> 1957		Norton and Eiseman 1975		Boretos and Pierce 1967	
<i>Review e.g.</i>	Xue and Greisler 2003		Nishibe <i>et al.</i> 2004		Tiwari <i>et al.</i> 2002	

IND (internodal distance).

agents. Antibiotics such as rifampicin bonded to Dacron did not reduce the incidence of vascular graft infection (Earnshaw *et al.* 2000); however, a silver-coated collagen-impregnated Dacron prosthesis offers an alternative approach in the treatment of vascular graft infection (Mirzaie *et al.* 2007).

Composite and vein-interposition vascular grafts

Attempts at improving disadvantageous anastomotic hemodynamics include alternative surgical techniques and interposition of an autologous vein segment.

Despite the theoretical and experimental advantage of end-to-end (straight) rather than end-to-side (Y-shaped) anastomosis between the graft and the artery (O'Brien *et al.* 2007), clinical trials did not prove superiority, and moreover an increased limb amputation rate was revealed in the end-to-end setting, probably due to exclusion of collateral vessels (Schouten *et al.* 2005). Suturing materials and techniques may also decrease the anastomotic compliance difference, e.g. in the case of interrupted sutures (more than one single suture).

However, running sutures are most widely used, because they are quicker and achieve better hemostasis (Tiwari *et al.* 2003).

Own venous tissue is placed in the form of a vein patch or cuff between an ePTFE graft and the artery to reduce the anastomotic compliance mismatch and to improve the prosthesis patency. Procedures implemented during the 1980s and 1990s include the Miller cuff, the Linton patch, the Taylor patch and the St. Mary boot (Moawad and Gagne 2003). When the patient's own vein graft is of insufficient length, feasible options that increase the patency rates may be: a *composite bypass* - prosthesis and vein are spliced together with an additional anastomosis (Bastounis *et al.* 1999), a *sequential bypass*, which is formed as a distal venous extension graft to the preceding proximal prosthetic bypass (Mahmood *et al.* 2002), and a *bridge graft*, which connects two patent distal arteries with a short vein segment (Deutsch *et al.* 2001). Although adjunctive arterio-venous fistula placement increases the blood flow through femoro-distal (i.e. below knee) bypasses, thus theoretically preventing thrombosis, several trials failed to show evidence of

benefit (Laurila *et al.* 2006).

Vein interposition techniques are generally useful in the case of distal anastomosis located below the knee, but are unimportant in the above-the-knee position (Mamode and Scott 2000). The underlying mechanism consists in reducing the compliance mismatch and suppressing IH (Cabrera Fischer *et al.* 2005). Research derived from this evidence led to the evolution of specially formed ePTFE grafts, capable of better harnessing the hemodynamic forces by an enlarged anastomotic hood. Interestingly, clinical studies have shown patency rates of this pre-cuffed ePTFE (Panneton *et al.* 2004) and carbon-lined ePTFE Distaflo graft (Fisher *et al.* 2003) that are comparable to interposition vein cuffs.

Biosynthetic/biohybrid vascular grafts (biografts)

The idea of introducing viable biological components into an artificial material-based vascular graft was established to produce more biocompatible vascular substitutes. A viable endothelial layer is the best antithrombogenic surface. There are three possible sources of graft lumen endothelialization *in vivo* post implantation: trans-anastomotic ingrowth from the native artery, transmural tissue and capillary ingrowth through the prosthesis wall, and a “fall-out” or blood-borne source from circulating progenitor cells. Unlike animals, humans seldom achieve spontaneous endothelialization more than 1-2 cm from an anastomosis, and transmural ingrowth is hampered by the structure of currently-used prostheses (Zilla *et al.* 2007).

The concept of seeding endothelial cells onto the graft lumen before implantation was experimentally implemented (Herring *et al.* 1978), and consequently this autologous EC “transplantation” managed to improve the patency of human Dacron prostheses (Herring *et al.* 1984) in a non-smoker population. Endothelial cells from a subcutaneous vein segment (*e.g.* saphenous, cephalic or jugular vein) are harvested and immediately seeded onto the graft lumen within the timeframe of one operation. The results of the first clinical trials performed during the 1980s were controversial and disappointing, mainly due to insufficient cell density (Bordenave *et al.* 1999). However, sophisticated EC extraction and retention techniques (Tiwari *et al.* 2001) and the search for more abundant cell sources, such as microvascular endothelium from fat tissue or an omentum biopsy, improved the outcome of this single-stage cell seeding method (Alobaid *et al.* 2005).

On the other hand, the double-stage approach involves a prolonged incubation and cell multiplication period (2-4 weeks) between cell harvesting and implantation (Bordenave *et al.* 2005). Nevertheless, *in vitro* flow pretreatment or shear stress preconditioning of the cell-material constructs before exposing them to arterial conditions enhances cell retention by inducing structural changes and adaptation (Rademacher *et al.* 2001). This means that an endothelial cell monolayer is physiologically exposed to certain mechanical forces, namely hydrostatic pressure resulting from blood pressure, shear stress resulting from tangential friction of blood flow against the vessel wall, and finally longitudinal and circumferential cyclic stretch resulting from repeated blood vessel distension due to the cardiac cycle (Lehoux *et al.* 2006). Chronic laminar versus turbulent shear stress seems to be of utmost importance for endothelial and vascular smooth muscle cell function (Chien 2007), regulating signal transduction in these cells (Daculsi *et al.* 2007), their alignment, molecule secretion, cytoskeleton reorganization, gene expression, cell migration, proliferation and survival. These processes ultimately influence thrombogenesis and atherogenesis (Yoshizumi *et al.* 2003), and similar cellular events and phenotypes (Rémy-Zolghadri *et al.* 2004, Fernandez *et al.* 2006) can be observed when cells are seeded on biomaterials (Fernandez *et al.* 2007).

Early results of clinical *in vitro* double-stage endothelialization were promising (Magometschnigg *et al.* 1992, Meinhart *et al.* 1997), and recently published overall 5-year and 10-year patency of endothelialized fibrin glue-precoated ePTFE femoro-popliteal bypass grafts of 69 % and 61 %, respectively (Deutsch *et al.* 2009) has already been correctly reported to close the gap between prosthetic and vein grafts (Meinhart *et al.* 1997). Interestingly, the feasibility of endothelium-seeded vascular prostheses was also confirmed in coronary bypass grafting, though it is not widely used (Laube *et al.* 2000).

To improve the above-mentioned clinical success, additional cell sources and cell retention technologies are being investigated. Vascular cells are anchorage-dependent, and integrin receptor-mediated adhesion occurs via ECM proteins that are adsorbed from the cell culture media *in vitro* or from blood *in vivo* (Bačáková *et al.* 2004). Thus, to regulate cell attachment, biomaterials are coated with ECM proteins such as collagen, laminin and fibronectin (Vara *et al.* 2005) or are modified by covalent bonding of short adhesive peptides

which may be cell specific, such as the Arg-Gly-Asp (RGD) sequence (Bačáková *et al.* 2007). Fibrin, as a natural scaffold for cell migration and healing, plays a pivotal role in tissue engineering of vascular grafts (Filová *et al.* 2009). In addition, less specific physical surface modifications, non-ligand based techniques (Salacinski *et al.* 2001) and surface nanoarchitecturing (de Mel *et al.* 2008) are being explored.

An alternative approach aimed at improving seeding efficiency is to use progenitor cells. Bone marrow cells infiltrated into the ePTFE vascular grafts and implanted in the aortic position of dogs retained complete endothelialization and patency after six months (Noishiki *et al.* 1996). Similarly, human endothelial progenitor cells (EPC) were isolated from peripheral blood (Asahara *et al.* 1997), and it has been shown in a canine model that a subset of CD34+ bone marrow cells can be mobilized to the circulation and can colonize the flow surfaces of Dacron vascular prostheses (Shi *et al.* 1998). Moreover, seeding of bone marrow-derived cells (BMC) accelerated early Dacron graft endothelialization without increasing thrombogenicity in a dog model (Bhattacharya *et al.* 2000). Taken together, due to their high proliferative capacity and differentiation potential, stem cells may represent the next era of cell-sourcing technology (Riha *et al.* 2005).

Genetically-modified cells have also been considered for the construction of vascular replacements. For example, genetically-modified endothelial cells over-expressing tissue plasminogen activator (t-PA) and urokinase-type PA, or bone marrow mesenchymal stem cells transduced to express endothelial nitric oxide synthase (eNOS), would promote cell repopulation of the graft and help to eliminate thrombotic events (Zarbin *et al.* 2007). However, the use of genetically-modified cells inherently raises ethical questions. A conventional approach is directly to load the materials with anti-coagulant, anti-inflammatory and cell growth-regulating substances, such as heparin and heparin-like molecules, as mentioned above (Walpoth *et al.* 1998, Lee *et al.* 2002), hirudin (Heise *et al.* 2006), dipyridamole (Aldenhoff *et al.* 2001), growth factors, such as vascular endothelial growth factor (Ehrbar *et al.* 2005) or fibroblast growth factor (Gray *et al.* 1994, Sato *et al.* 2008), or antimigratory and antiproliferative drugs paclitaxel (Lim *et al.* 2007), sirolimus (Ishii *et al.* 2008) and inhibitors of CDK2 kinase (Brooks *et al.* 1997). These drugs can be incorporated directly into the prosthesis wall or delivered through drug-eluting stents

(Lee *et al.* 2008), catheters and perivascular collars (for review see Sriram and Patterson 2001). Artificial materials releasing NO are also being developed, consisting of synthetic polymers (e.g. polyurethane, PTFE) incorporated with NO donors, such as diazeniumdiolates and S-nitrosothiols (Varu *et al.* 2009).

Living vascular grafts, totally-engineered blood vessels (TEBV)

The concept of completely biological living grafts implies the ability to remodel, grow, self-repair and respond to the immediate environment. Similarly to the native artery, the graft would consist of a functional endothelial cell layer resting on metabolically active smooth muscle cells which are in a contractile (i.e. non-synthetic and anti-atherogenic) phenotype (Muto *et al.* 2007). The graft would also contain enough collagen and elastin proteins to display desirable viscoelastic properties, and would lack any synthetic foreign material that would initiate chronic inflammatory responses or be susceptible to infection. If a synthetic material is used, it should be non-immunogenic, non-thrombogenic and of appropriate compliance. In the ideal case, it should be degradable, providing a temporary scaffold for vascular tissue regeneration, gradually removed and replaced by the newly-forming tissue.

In the construction of tissue-engineered vascular grafts, three major components must be addressed: a scaffold to provide the initial graft shape and strength, adhesive matrix and living vascular cells (Baguneid *et al.* 2006). For the scaffold, four major approaches can be identified: permanent synthetic support, natural acellular tissues, a biodegradable scaffold, and non-scaffold technology (Campbell and Campbell 2007).

The first seminal attempt involved seeding bovine EC, VSMC and fibroblasts onto collagen gel tubes (Weinberg and Bell 1986). An external Dacron mesh reinforcement had to be added because of poor mechanical strength. An example of the use of a natural scaffold involves seeding human cells on decellularized porcine aorta (Bader *et al.* 2000) or small intestinal submucosa implanted as vascular grafts (Lantz *et al.* 1993). The most commonly used biodegradable polymer is polyglycolic acid (PGA). Under shear stress preconditioning, VSMC followed by EC were seeded onto this scaffold, and the resulting vascular grafts were implanted in pigs, with excellent results (Niklason *et al.* 1999). Seeding bone-marrow cells (BMC) onto a biodegradable scaffold enabled the establishment of a

Table 4. Composite, biosynthetic and totally engineered vascular grafts in experimental and clinical use.

	Composite grafts	Biosynthetic grafts		Totally engineered blood vessels	
	Clinical	Experimental	Clinical	Experimental	Clinical
<i>Advantages</i>	Reduced compliance mismatch and intimal hyperplasia, improved patency in femoro-distal bypasses, easier redo surgery	Antithrombogenic, better patency		Responsiveness, non-thrombogenicity, self-repair, growth, metabolically active, potentially cost-effective	
<i>Disadvantages</i>	Technically demanding, prolonged surgery time	No emergency use, cell amplification problems, cell culture contamination risk		Demanding fabrication, time- and cost-consuming, bioreactor cell laboratory, specialized centers only	
<i>Healing</i>	Less intimal hyperplasia	Self-renewing functional endothelium		Complete integration	
<i>First use</i>	Siegman 1979	Herring <i>et al.</i> 1978	Herring <i>et al.</i> 1984	Weinberg and Bell 1986	Shin'oka <i>et al.</i> 2001
<i>Review e.g.</i>	Moawad and Gagne 2003	Seifalian <i>et al.</i> 2002	Bordenave <i>et al.</i> 2008	Isenberg <i>et al.</i> 2006	L'Heureux <i>et al.</i> 2007a

totally-engineered autograft implanted into the inferior vena cava of a dog, and explant analysis showed that BMC were able to differentiate both in endothelial and smooth muscle cells (Matsumura *et al.* 2003).

The non-scaffold or “self-assembly” approach produced a graft composed exclusively of human cells by wrapping and culturing fibroblast and VSMC cellular sheets on a PTFE mandril, which was then removed and EC were seeded in the lumen. This was the first completely biological TEBV to display mechanical resistance comparable to that of human vessels (experimental burst strength up to 2000 mm Hg). A short-term experiment in a canine model demonstrated good surgical handling (L'Heureux *et al.* 1998). Another vascular graft was developed without a scaffold by inducing an inflammatory reaction. A silastic tube was placed into the peritoneal cavity of rats and rabbits and was in 2 weeks spontaneously covered by layers of myofibroblasts, collagen matrix, and a single layer of mesothelium. It was everted to resemble the blood vessel and grafted in carotid or aortic position, and remained patent after 4 months (Campbell *et al.* 1999).

A TEBV construct consisting of autologous bone marrow cells seeded onto a biodegradable scaffold was first clinically implanted to replace the pulmonary artery in a pediatric patient with a cyanotic defect (Shin'oka *et al.* 2001), and there was no graft-related complication in a group of 42 patients (Shin'oka *et al.* 2005). Another clinical success was achieved by TEBV produced by the cell-sheet multilayer method (L'Heureux

et al. 2006) and implanted as a hemodialysis access graft in 10 patients (L'Heureux *et al.* 2007b). A recent paper reports primary 1-month and 6-month patency of 78 % and 60 %, respectively, meeting the approved criteria for a high-risk patient cohort (McAlister *et al.* 2009).

A summary of composite, biohybrid and totally-engineered vascular grafts is outlined in Table 4.

Venous replacements

Unlike arterial reconstructions, venous reconstructions are limited to large-diameter central veins, such as the inferior (Schwarzbach *et al.* 2006) or superior vena cava and the ilio-femoral veins (Kalra *et al.* 2003). Autologous size- and length-matched veins such as superficial femoral vein, internal jugular vein, left renal vein and mainly the gold-standard composite saphenous spiral vein graft (Doty *et al.* 1999) produce the best results, *e.g.* for a bypass for an occlusion of the superior vena cava of iatrogenic origin (catheter-related thrombosis). Venous allogeneous homografts, either fresh or cryopreserved, have been used in experimental and clinical venous reconstruction (Sitzmann *et al.* 1984). The patency of externally supported ePTFE is better than the patency of grafts fashioned from ePTFE alone. However, the use of synthetic prostheses may be limited by low-flow thrombogenicity, contaminated tissue beds, as in venous vascular trauma or tumor resection cases, *e.g.* in portal vein resection for pancreatic carcinoma (Leon *et al.* 2007). Venous reconstructions may be combined with an

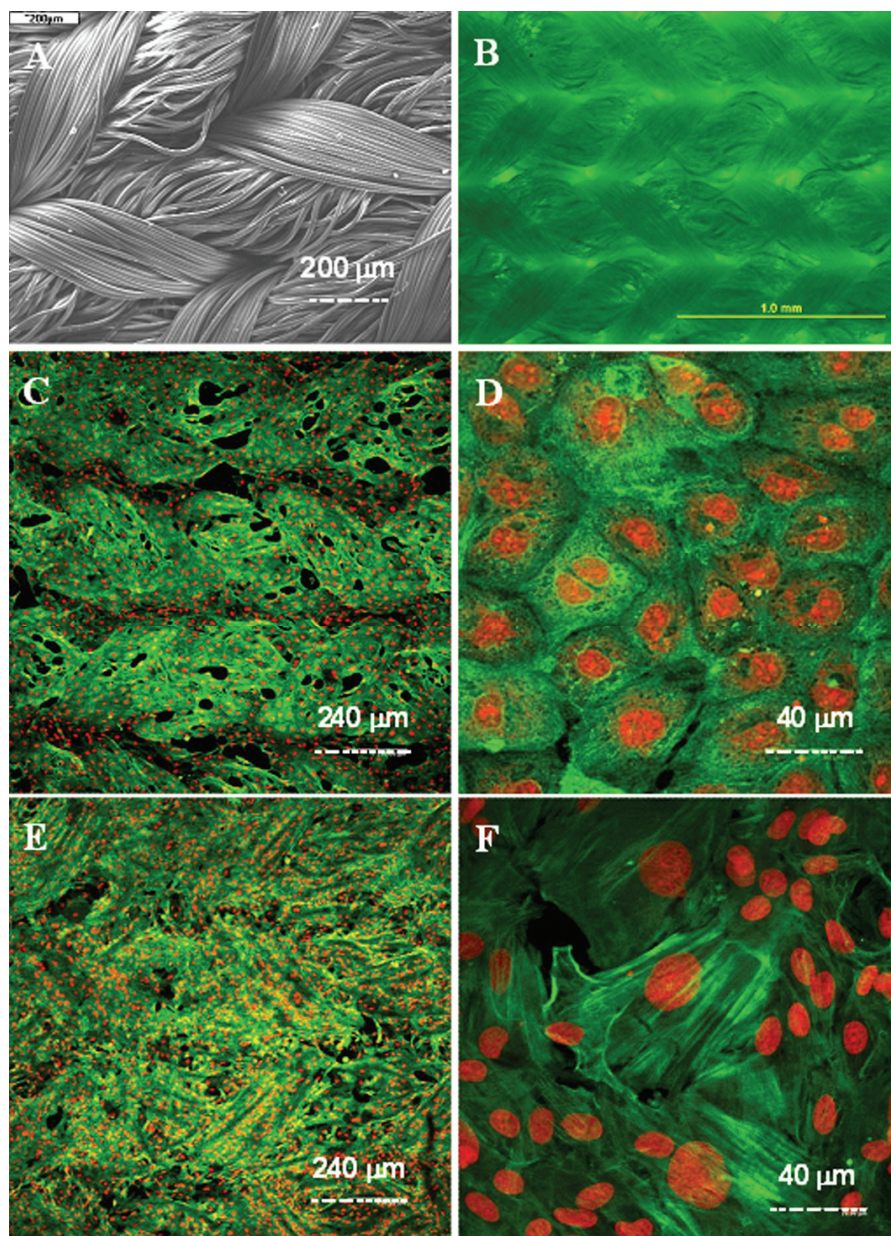


Fig. 3. Innovation of knitted PET vascular prostheses produced by VUP a.s., Brno, Czech Republic. **A:** the rough and highly hydrophobic inner surface of the prosthesis not suitable for cell adhesion; **B:** coating of this surface with defined molecular assemblies such as collagen I (Co), laminin (LM), fibronectin (FN) or fibrin (Fb) makes it more adhesive; **C, D, E, F:** confluent layer of bovine vascular endothelial cells (line CPAE) on CoFb (C, D) and multilayer of rat aortic smooth muscle cells on CoFN (E), CoLM (F) on the inner surface of a knitted wrapped prosthesis. Immunofluorescence of beta-actin (C, D) or alpha-actin (E, F), nuclei counterstained with propidium iodide. NOVA nanoSEM 200 FEI electron microscope (A), Olympus IX 51 fluorescence microscope, obj. 4x (B) or Leica TCS SP2 AOBs confocal microscope, obj. 10x, 29x5 μm (C), 40x, zoom 2x, 4x1 μm (D), obj. 10x, 9x15 μm (E), 40x, zoom 2x, 55x1 μm (F).

adjuvant arterio-venous fistula to improve graft patency. Interestingly, the success rates of repeated percutaneous angioplasty approached those of operative reconstruction (Wisselink *et al.* 1993).

Hemodialysis vascular access

Creating a native arterio-venous fistula for chronic hemodialysis access is obviously superior to creating a prosthetic fistula. However, patients with depleted veins receive a prosthetic access graft, which has to provide sufficient blood flow and sustain repeated puncturing. It is difficult to identify the ideal access graft from a large number of biologic and prosthetic conduits. Randomized trials suggest that cuffed ePTFE grafts offer

reasonable patency and that “early access” PU grafts can provide a safe conduit. Biologic/semibiologic grafts give better results than synthetic grafts, in some respects (Scott and Glickman 2007).

Collective experience of the authors

Static experiments performed in our laboratory enabled us to assess the cell colonization of various biomaterials without flow of culture medium. Clinically-used PET vascular prostheses (produced by VÚP, Brno, Czech Republic) were impregnated with biodegradable polyester-based copolymers (Pamula *et al.* 2008) as a background for further modification with multilayers of adhesive matrix proteins, such as collagen, fibronectin,

laminin and particularly fibrin, which can be derived in autologous form from the patient's blood (Brynda *et al.* 2005, Filová *et al.* 2009). For the purposes of vascular tissue engineering, we explored the adhesion and growth of animal-derived endothelial and vascular smooth muscle cells (Chlupáč *et al.* 2008a) (Fig. 3). Research work carried out on some fibrin-based surfaces (Riedel *et al.* 2009) has led to a common patent (Brynda *et al.* 2008).

Dynamic experiments were conducted under defined medium flow to better mimic *in vivo* conditions. Dacron vascular prostheses were modified on the luminal surface with ECM protein assemblies (Chlupáč *et al.* 2006a), and an investigation was made of the adhesion and growth of human patient-derived EC (Chlupáč *et al.* 2006b). Their resistance to shear stress was also investigated (Chlupáč *et al.* 2006c). The gene expression profile of endothelium seeded on similar planar ECM analogues was also determined (Chlupáč *et al.* 2008b). Hybrid vascular graft constructs *in vitro* seeded with autologous EC and preconditioned under shear stress in a bioreactor (Provitro Co., Berlin Germany) are currently being investigated in a porcine model.

Conclusions

Open arterial surgery in the form of a lower limb bypass remains the standard treatment for extensive and multilevel atherosclerotic disease. Over the past 50 years,

no synthetic alternative has been found to compare with the patency rates of gold-standard autologous conduits. However, appropriate native material is often unavailable and alternative vascular prostheses show poor clinical performance. To address this issue, luminal modifications of grafts, interposition of vein segments and particularly tissue engineering of small-sized blood vessels have been introduced. Biohybrid and tissue-engineered vascular grafts have been manufactured with the help of materials research, complex tissue culture and cell seeding technologies. The outcomes of experimental and clinical implants seem to be favorable. It is likely that cell therapy will become a frequent option in vascular and endovascular surgery in the coming decades.

Conflict of Interest

There is no conflict of interest.

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References

- ALOBALD N, SALACINSKI HJ, SALES KM, HAMILTON G, SEIFALIAN AM: Single stage cell seeding of small diameter prosthetic cardiovascular grafts. *Clin Hemorheol Microcirc* **33**: 209-226, 2005.
- ALDENHOFF YB, VAN DER VEEN FH, TER WOORST J, HABETS J, POOLE-WARREN LA, KOOLE LH: Performance of a polyurethane vascular prosthesis carrying a dipyridamole (Persantin) coating on its luminal surface. *J Biomed Mater Res* **54**: 224-233, 2001.
- ALI AT, MODRALL JG, HOCKING J, VALENTINE RJ, SPENCER H, EIDT JF, CLAGETT GP: Long-term results of the treatment of aortic graft infection by in situ replacement with femoral popliteal vein grafts. *J Vasc Surg* **50**: 30-39, 2009.
- ASAHARA T, MUROHARA T, SULLIVAN A, SILVER M, VAN DER ZEE R, LI T, WITZENBICHLER B, SCHATTEMAN G, ISNER JM: Isolation of putative progenitor endothelial cells for angiogenesis. *Science* **275**: 964-967, 1997.
- BAČÁKOVÁ L, FILOVÁ E, RYPÁČEK F, ŠVORČÍK V, STARÝ V: Cell adhesion on artificial materials for tissue engineering. *Physiol Res* **53** (Suppl 1): S35-S45, 2004.
- BAČÁKOVÁ L, FILOVÁ E, KUBIES D, MACHOVÁ L, PROKS V, MALINOVÁ V, LISÁ V, RYPÁČEK F: Adhesion and growth of vascular smooth muscle cells in cultures on bioactive RGD peptide-carrying polylactides. *J Mater Sci Mater Med* **18**: 1317-1323, 2007.

- BADER A, STEINHOFF G, STROBL K, SCHILLING T, BRANDES G, MERTSCHING H, TSIKAS D, FROELICH J, HAVERICH A: Engineering of human vascular aortic tissue based on a xenogeneic starter matrix. *Transplantation* **70**: 7-14, 2000.
- BAGUNEID MS, SEIFALIAN AM, SALACINSKI HJ, MURRAY D, HAMILTON G, WALKER MG: Tissue engineering of blood vessels. *Br J Surg* **93**: 282-290, 2006.
- BASTOUNIS E, GEORGOPOULOS S, MALTEZOS C, ALEXIOU D, CHIOTOPOULOS D, BRAMIS J: PTFE-vein composite grafts for critical limb ischaemia: a valuable alternative to all-autogenous infrageniculate reconstructions. *Eur J Vasc Endovasc Surg* **18**: 127-132, 1999.
- BEGHI C, NICOLINI F, BUDILLON AM, BORRELLO B, BALLORE L, REVERBERI C, GHERLI T.: Midterm clinical results in myocardial revascularization using the radial artery. *Chest* **122**: 2075-2079, 2002.
- BERGER K, SAUVAGE L, RAO A, WOOD S: Healing of arterial prostheses in man: its incompleteness. *Ann Surg* **175**: 118-127, 1972.
- BHATTACHARYA V, McSWEENEY PA, SHI Q, BRUNO B, ISHIDA A, NASH R, STORB RF, SAUVAGE LR, HAMMOND WP, WU MH: Enhanced endothelialization and microvessel formation in polyester grafts seeded with CD34⁺ bone marrow cells. *Blood* **95**: 581-585, 2000.
- BLAKEMORE AH, VOORHEES AB Jr: The use of tubes constructed from vinyon N cloth in bridging arterial defects; experimental and clinical. *Ann Surg* **140**: 324-334, 1954.
- BLUMENBERG RM, GELFAND ML, BARTON EA, BOWERS CA, GITTLEMAN DA: Clinical significance of aortic graft dilation. *J Vasc Surg* **14**: 175-180, 1991.
- BORDENAVE L, RÉMY-ZOLGHADRI M, FERNANDEZ P, BAREILLE R, MIDY D: Clinical performance of vascular grafts lined with endothelial cells. *Endothelium* **6**: 267-275, 1999.
- BORDENAVE L, FERNANDEZ P, RÉMY-ZOLGHADRI M, VILLARS S, DACULSI R, MIDY D: In vitro endothelialized ePTFE prostheses: clinical update 20 years after the first realization. *Clin Hemorheol Microcirc* **33**: 227-234, 2005.
- BORDENAVE L, MENU P, BAQUEY C: Developments towards tissue-engineered, small-diameter arterial substitutes. *Expert Rev Med Devices* **5**: 337-347, 2008.
- BORETOS JW, PIERCE WS: Segmented polyurethane: a new elastomer for biomedical applications. *Science* **158**: 1481-1482, 1967.
- BOSIERS M, DELOOSE K, VERBIST J, SCHROË H, LAUWERS G, LANSINK W, PEETERS P: Heparin-bonded expanded polytetrafluoroethylene vascular graft for femoropopliteal and femorocrural bypass grafting: 1-year results. *J Vasc Surg* **43**: 313-319, 2006.
- BREWSTER DC: Current controversies in the management of aortoiliac occlusive disease. *J Vasc Surg* **25**: 365-379, 1997.
- BROOKS EE, GRAY NS, JOLY A, KERWAR SS, LUM R, MACKMAN RL, NORMAN TC, ROSETE J, ROWE M, SCHOW SR, SCHULTZ PG, WANG X, WICK MM, SHIFFMAN D: CVT-313, a specific and potent inhibitor of CDK2 that prevents neointimal proliferation. *J Biol Chem* **272**: 29207-29211, 1997.
- BRYNDA E, PACHERNÍK J, HOUSKA M, PIENKA Z, DVOŘÁK P: Surface immobilized protein multilayers for cell seeding. *Langmuir* **21**: 7877-7883, 2005.
- BRYNDA E, RIEDEL T, DYR J, HOUSKA M, BAČÁKOVÁ L, FILOVÁ E, CHLUPÁČ J, LESNÝ P, JENDELOVÁ P, SYKOVÁ E: Mode of preparation of controlled fibrin assemblies on solid surfaces (Způsob přípravy regulovaných vrstev fibrinu na pevných površích), *Czech Patent Application*, registered in the UPV Office under No. 2006-821, accepted in 2008.
- CABRERA FISCHER EI, BIA SANTANA D, CASSANELLO GL, ZÓCALO Y, CRAWFORD EV, CASAS RF, ARMENTANO RL: Reduced elastic mismatch achieved by interposing vein cuff in expanded polytetrafluoroethylene femoral bypass decreases intimal hyperplasia. *Artif Organs* **29**: 122-130, 2005.
- CALLIGARO KD, SYREK JR, DOUGHERTY MJ, RUA I, RAVIOLA CA, DELAURENTIS DA: Use of arm and lesser saphenous vein compared with prosthetic grafts for infrapopliteal arterial bypass: are they worth the effort? *J Vasc Surg* **26**: 919-927, 1997.
- CAMPBELL JH, EFENDY JL, CAMPBELL GR: Novel vascular graft grown within recipient's own peritoneal cavity. *Circ Res* **85**: 1173-1178, 1999.

- CAMPBELL GR, CAMPBELL JH: Development of tissue engineered vascular grafts. *Curr Pharm Biotechnol* **8**: 43-50, 2007.
- CHANG BB, PATY PS, SHAH DM, LEATHER RP: The lesser saphenous vein: an underappreciated source of autogenous vein. *J Vasc Surg* **15**: 152-157, 1992.
- CHEMLA ES, MORSY M: Randomized clinical trial comparing decellularized bovine ureter with expanded polytetrafluoroethylene for vascular access. *Br J Surg* **96**: 34-39, 2009.
- CHESHIRE NJW, WOLFE MS, NOONE MA, DAVIES L, DRUMMOND M: The economics of femorocrural reconstruction for critical limb ischaemia with and without autologous vein. *J Vasc Surg* **15**: 167-175, 1992.
- CHIEN S: Mechanotransduction and endothelial cell homeostasis: the wisdom of the cell. *Am J Physiol* **292**: H1209-H1224, 2007.
- CHLUPÁČ J, FILOVÁ E, RIEDEL T, BRYNDA E, REMY-ZOLGHADRI M, BAREILLE R, FERNANDEZ P, DACULSI R, BORDENAVE L, BAČÁKOVÁ L: Human endothelium on vascular prostheses modified by extracellular matrix proteins in a flow experiment. *Eng Biomater* **58-60**: 10-13, 2006a.
- CHLUPÁČ J, FILOVÁ E, RIEDEL T, BRYNDA E, REMY-ZOLGHADRI M, BAREILLE R, FERNANDEZ P, DACULSI R, BORDENAVE L, BAČÁKOVÁ L: Innovation of clinically used vascular prostheses by protein assemblies on their luminal surface. *Physiol Res* **55**: 24P, 2006b.
- CHLUPÁČ J, FILOVÁ E, RIEDEL T, BRYNDA E, RÉMY-ZOLGHADRI M, BAREILLE R, FERNANDEZ P, DACULSI R, BORDENAVE L, BAČÁKOVÁ L: Endothelial cells seeded on protein-coated vascular prostheses under experimental shear stress conditions. *Atherosclerosis Suppl* **7**: 159, 2006c.
- CHLUPÁČ J, FILOVÁ E, RIEDEL T, BRYNDA E, PAMULA E, LISÁ V, BAČÁKOVÁ L: Endothelial cells on PET vascular prostheses impregnated with polyester-based copolymers and coated with cell-adhesive protein assemblies. *Eng Biomater* **81-84**: 108-111, 2008a.
- CHLUPÁČ J, FILOVÁ E, RIEDEL T, BRYNDA E, RÉMY-ZOLGHADRI M, BAREILLE R, FERNANDEZ P, DACULSI R, BORDENAVE L, BAČÁKOVÁ L: Endothelial cells seeded on protein multilayer assemblies under shear stress. *Physiol Res* **57**: 77P, 2008b.
- CLOWES AW, ZACHARIAS RK, KIRKMAN TR: Early endothelial coverage of synthetic arterial grafts: porosity revisited. *Am J Surg* **153**: 501-504, 1987.
- CONTE MS: The ideal small arterial substitute: a search for the Holy Grail? *FASEB J* **12**: 43-45, 1998.
- COOPER GJ, UNDERWOOD MJ, DEVERALL PB: Arterial and venous conduits for coronary artery bypass. A current review. *Eur J Cardiothorac Surg* **10**: 129-140, 1996.
- DACULSI R, RÉMY-ZOLGHADRI M, GRELLIER M, CONRAD V, FERNANDEZ P, BAREILLE R, BORDENAVE L: Signal transduction and procoagulant state of human cord blood-progenitor-derived endothelial cells after interleukin-1 α stimulation. *Endothelium* **14**: 163-171, 2007.
- DAHL SL, KOH J, PRABHAKAR V, NIKLASON LE: Decellularized native and engineered arterial scaffolds for transplantation. *Cell Transplant* **12**: 659-666, 2003.
- DARDIK H, WENGERTER K, QIN F, PANGILINAN A, SILVESTRI F, WOLODIGER F, KAHN M, SUSSMAN B, IBRAHIM IM: Comparative decades of experience with glutaraldehyde-tanned human umbilical cord vein graft for lower limb revascularization: an analysis of 1275 cases. *J Vasc Surg* **35**: 64-71, 2002.
- DE MEL A, BOLVIN C, EDIRISINGHE M, HAMILTON G, SEIFALIAN AM: Development of cardiovascular bypass grafts: endothelialization and applications of nanotechnology. *Expert Rev Cardiovasc Ther* **6**: 1259-1277, 2008.
- DE MOL VAN OTTERLOO JC, VAN BOCKEL JH, PONFOORT ED, BRIET E, BROMMER EJ, HERMANS J, DAHA MR: Systemic effects of collagen-impregnated aortoiliac Dacron vascular prostheses on platelet activation and fibrin formation. *J Vasc Surg* **14**: 59-66, 1991.
- DEREUME JP, VAN ROMPHEY A, VINCENT G, ENGELMANN E: Femoropopliteal bypass with a compliant, composite polyurethane/Dacron graft: short-term results of a multicentre trial. *Cardiovasc Surg* **1**: 499-503, 1993.
- DEUTSCH M, MEINHART J, HOWANIETZ N, FRÖSCHL A, HEINE B, MOIDL R, MENDEL H, SISEL A, STÜMPFLEN A, ZILLA P: The bridge graft: a new concept for infrapopliteal surgery. *Eur J Vasc Endovasc Surg* **21**: 508-512, 2001.

- DEUTSCH M, MEINHART J, ZILLA P, HOWANIETZ N, GORLITZER M, FROESCHL A, STUEMPFLEN A, BEZUIDENHOUT D, GRABENWOEGER M: Long-term experience in autologous in vitro endothelialization of infrainguinal ePTFE grafts. *J Vasc Surg* **49**: 352-362, 2009.
- DEVINE C, MCCOLLUM C, THE NORTH WEST FEMOROPLOPLITEAL TRIAL PARTICIPANTS: Heparin-bonded Dacron and polytetrafluoroethylene for femoropopliteal bypass: five years results of a perspective randomized multi-centre clinical trial. *J Vasc Surg* **40**: 924-931, 2004.
- DOTY JR, FLORES JH, DOTY DB: Superior vena cava obstruction: bypass using spiral vein graft. *Ann Thorac Surg* **67**: 1111-1116, 1999.
- EARNSHAW JJ, WHITMAN B, HEATHER BP: Two-year results of a randomized controlled trial of rifampicin-bonded extra-anatomic dacron grafts. *Br J Surg* **87**: 758-759, 2000.
- EHRBAR M, METTERS A, ZAMMARETTI P, HUBBELL JA, ZISCH AH: Endothelial cell proliferation and progenitor maturation by fibrin-bound VEGF variants with differential susceptibilities to local cellular activity. *J Control Release* **101**: 93-109, 2005.
- EMRECAN B, YILIK L, OZBEK C, GÜRBÜZ A: Bovine ureter graft for haemodialysis access surgery. *Nephrol Dial Transplant* **21**: 2290-2291, 2006.
- FAHNER PJ, IDU MM, VAN GULIK TM, LEGEMATE DA: Systematic review of preservation methods and clinical outcome of infrainguinal vascular allografts. *J Vasc Surg* **44**: 518-524, 2006.
- FARIES PL, LOGERFO FW, ARORA S, PULLING MC, ROHAN DI, AKBARI CM, CAMPBELL DR, GIBBONS GW, POMPOSELLI FB: Arm vein conduit is superior to composite prosthetic-autogenous grafts in lower extremity revascularization. *J Vasc Surg* **31**: 1119-1127, 2000.
- FERNANDEZ P, DACULSI R, RÉMY-ZOLGHADRI M, BAREILLE R, BORDENAVE L: Endothelial cells cultured on engineered vascular grafts are able to transduce shear stress. *Tissue Eng* **12**: 1-7, 2006.
- FERNANDEZ P, BOURGET C, BAREILLE R, DACULSI R, BORDENAVE L: Gene response in endothelial cells cultured on engineered surfaces is regulated by shear stress. *Tissue Eng* **13**: 1607-1614, 2007.
- FILOVÁ E, BRYNDA E, RIEDEL T, BAČÁKOVÁ L, CHLUPÁČ J, LISÁ V, HOUSKA M, DYR JE: Vascular endothelial cells on two-and three-dimensional fibrin assemblies for biomaterial coatings. *J Biomed Mater Res A* **90**: 55-69, 2009.
- FISHER RK, KIRKPATRICK UJ, HOW TV, BRENNAN JA, GILLING-SMITH GL, HARRIS PL: The distaflo graft: a valid alternative to interposition vein? *Eur J Vasc Endovasc Surg* **25**: 235-239, 2003.
- GLICKMAN MH, STOKES GK, ROSS JR, SCHUMAN ED, STERNBERGH WC, LINDBERG JS, MONEY SM, LORBER MI: Multicenter evaluation of a polytetrafluoroethylene vascular access graft as compared with the expanded polytetrafluoroethylene vascular access graft in hemodialysis applications. *J Vasc Surg* **34**: 465-473, 2001.
- GOLDEN MA, HANSON SR, KIRKMAN TR, SCHNEIDER PA, CLOWES AW: Healing of polytetrafluoroethylene arterial grafts is influenced by graft porosity. *J Vasc Surg* **11**: 838-844, 1990.
- GOYANNES J: Nuevos trabajos de cirugía vascular, substitucion plastica de las arterias por las venas o arterioplastia venosa aplicada como nuevo metodo al tratamiento de los aneurysmas. *El Siglo Med* **53**: 446, 1906.
- GRAY JL, KANG SS, ZENNI GC, KIM DU, KIM PI, BURGESS WH, DROHAN W, WINKLES JA, HAUDENSCHILD CC, GREISLER HP: FGF-1 affixation stimulates ePTFE endothelialization without intimal hyperplasia. *J Surg Res* **57**: 596-612, 1994.
- GROSS RE, HURWITT ES, BILL AH JR, PIERCE EC Jr: Preliminary observations on the use of human arterial grafts in the treatment of certain cardiovascular defects. *N Engl J Med* **239**: 578, 1948.
- GUIDOIN R, CHAKFÉ N, MAUREL S, HOW T, BATT M, MAROIS M, GOSSELIN C: Expanded polytetrafluoroethylene arterial prostheses in humans: histopathological study of 298 surgically excised grafts. *Biomaterials* **14**: 678-693, 1993a.
- GUIDOIN R, MAUREL S, CHAKFÉ N, HOW T, ZHANG Z, THERRIEN M, FORMICHI M, GOSSELIN C: Expanded polytetrafluoroethylene arterial prostheses in humans: chemical analysis of 79 explanted specimens. *Biomaterials* **14**: 694-704, 1993b.

- GUIDOIN R, MAROIS Y, DENG X, CHAKFÉ N, MAROIS M, ROY R, KING MW, DOUVILLE Y. Can collagen impregnated polyester arterial prostheses be recommended as small diameter blood conduits? *ASAIO J* **42**: 974-983, 1996.
- GUYTON RA: Coronary artery bypass is superior to drug-eluting stents in multivessel coronary artery disease. *Ann Thorac Surg* **81**: 1949-1957, 2006.
- HARUGUCHI H, TERAOKA S: Intimal hyperplasia and haemodynamic factors in arterial bypass and arteriovenous grafts: a review. *J Artif Organs* **6**: 227-235, 2003.
- HEISE M, SCHMIDMAIER G, HUSMANN I, HEIDENHAIN C, SCHMIDT J, NEUHAUS P, SETTMACHER U: PEG-hirudin/iloprost coating of small diameter ePTFE grafts effectively prevents pseudointima and intimal hyperplasia development. *Eur J Vasc Endovasc Surg* **32**: 418-424, 2006.
- HERRING M, GARDNER A, GLOVER J: A single-staged technique for seeding vascular grafts with autogenous endothelium. *Surgery* **84**: 498-504, 1978.
- HERRING MB, DILLEY R, JERSILD RA JR, BOXER L, GARDNER A, GLOVER J: Seeding arterial prostheses with vascular endothelium. The nature of the lining. *Ann Surg* **190**: 84-90, 1979.
- HERRING M, GARDNER A, GLOVER J: Seeding human arterial prostheses with mechanically derived endothelium. The detrimental effect of smoking. *J Vasc Surg* **1**: 279-289, 1984.
- HURT AV, BATELLO-CRUZ M, SKIPPER BJ, TEAF SR, STERLING WA Jr: Bovine carotid artery heterografts versus polytetrafluoroethylene grafts. A prospective, randomized study. *Am J Surg* **146**: 844-847, 1983.
- ISENBERG BC, WILLIAMS C, TRANQUILLO RT: Small-diameter artificial arteries engineered in vitro. Review. *Circ Res* **98**: 25-35, 2006.
- ISHII Y, SAKAMOTO S, KRONENGOLD RT, VIRMANI R, RIVERA EA, GOLDMAN SM, PRECHTEL EJ, HILL JG, DAMIANO RJ Jr: A novel bioengineered small-caliber vascular graft incorporating heparin and sirolimus: excellent 6-month patency. *J Thorac Cardiovasc Surg* **135**: 1237-1245, 2008.
- JABOULAY M, BRIAU E: Recherches expérimentelles sur la suture et la greffe artérielles. *Lyon Méd* **81**: 97-99, 1896.
- JESCHKE MG, HERMANUTZ V, WOLF SE, KÖVEKER GB: Polyurethane vascular prostheses decreases neointimal formation compared with expanded polytetrafluoroethylene. *J Vasc Surg* **29**: 168-176, 1999.
- JOHNSON WC, LEE KK: A comparative evaluation of polytetrafluoroethylene, umbilical vein, and saphenous vein bypass grafts for femoral-popliteal above-knee revascularization: a prospective randomized Department of Veterans Affairs cooperative study. *J Vasc Surg* **32**: 268-277, 2000.
- KACHLIK D, BAČA V, STINGL J, SOSNA B, LAMETSCHWANDTNER A, MINNICH B, SETINA M: Architectonic arrangement of the vasa vasorum of the human great saphenous vein. *J Vasc Res* **44**: 157-166, 2007.
- KAKISIS JD, LIAPIS CD, BREUER C, SUMPIO BE: Artificial blood vessel: the Holy Grail of peripheral vascular surgery. *J Vasc Surg* **41**: 349-354, 2005.
- KALRA M, GLOVICZKI P, ANDREWS JC, CHERRY KJ JR, BOWER TC, PANNETON JM, BJARNASON H, NOEL AA, SCHLECK C, HARMSSEN WS, CANTON LG, PAIROLERO PC: Open surgical and endovascular treatment of superior vena cava syndrome caused by nonmalignant disease. *J Vasc Surg* **38**: 215-223, 2003.
- KAPFER X, MEICHELBOECK W, GROEGLER FM: Comparison of carbon-impregnated and standard ePTFE prostheses in extra-anatomical anterior tibial artery bypass: a prospective randomized multicenter study. *Eur J Vasc Endovasc Surg* **32**: 155-168, 2006.
- KATZMAN HE, GLICKMAN MH, SCHILD AF, FUJITANI RM, LAWSON JH: Multicenter evaluation of the bovine mesenteric vein bioprosthesis for hemodialysis access in patients with an earlier failed prosthetic graft. *J Am Coll Surg* **201**: 223-230, 2005.
- KIEFFER E, GOMES D, CHICHE L, FLÉRON MH, KOSKAS F, BAHNINI A: Allograft replacement for infrarenal aortic graft infection: early and late results in 179 patients. *J Vasc Surg* **39**: 1009-1017, 2004.
- KLINKERT P, POST PN, BRESLAU PJ, VAN BOCKEL JH: Saphenous vein versus PTFE for above-knee femoropopliteal bypass. A review of the literature. *Eur J Vasc Endovasc Surg* **27**: 357-362, 2004.
- KOHLER TR, STRATTON JR, KIRKMAN TR, JOHANSEN KH, ZIERLER BK, CLOWES AW: Conventional versus high-porosity polytetrafluoroethylene grafts: clinical evaluation. *Surgery* **112**: 901-907, 1992.

- LANTZ GC, BADYLAK SF, HILES MC, COFFEY AC, GEDDES LA, KOKINI K, SANDUSKY GE, MORFF RJ: Small intestinal submucosa as a vascular graft: a review. *J Invest Surg* **6**: 297-310, 1993.
- LAUBE HR, DUWE J, RUTSCH W, KONERTZ W: Clinical experience with autologous endothelial cell-seeded polytetrafluoroethylene coronary artery bypass grafts. *J Thorac Cardiovasc Surg* **120**: 134-141.
- LAURILA K, LUTHER M, ROTH WD, VILKKO P, KANTONEN I, TEITTINEN K, SIHVO EI, IHLBERG L, ALBÄCK A, LEPÄNTALO M: Adjuvant arteriovenous fistula as means of rescue for infrapopliteal venous bypass with poor runoff. *J Vasc Surg* **44**: 985-992, 2006.
- LAWSON JA, TANGELDER MJ, ALGRA A, EIKELBOOM BC: The myth of the in situ graft: superiority in infrainguinal bypass surgery? *Eur J Vasc Endovasc Surg* **18**: 149-157, 1999.
- LEE HJ, HONG JK, GOO HC, LEE WK, PARK KD, KIM SH, YOO YM, KIM YH: Improved blood compatibility and decreased VSMC proliferation of surface-modified metal grafted with sulfonated PEG or heparin. *J Biomater Sci Polym Ed* **13**: 939-952, 2002.
- LEE SW, PARK SW, KIM YH, YUN SC, PARK DW, LEE CW, HONG MK, RHEE KS, CHAE JK, KO JK, PARK JH, LEE JH, CHOI SW, JEONG JO, SEONG IW, CHO YH, LEE NH, KIM JH, CHUN KJ, KIM HS, PARK SJ: A randomized comparison of sirolimus versus paclitaxel-eluting stent implantation in patients with diabetes mellitus. *J Am Coll Cardiol* **52**: 727-733, 2008.
- LEHOUX S, CASTIER Y, TEDGUI A: Molecular mechanisms of the vascular responses to haemodynamic forces. *J Intern Med* **259**: 381-392, 2006.
- L'HEUREUX N, PÂQUET S, LABBÉ R, GERMAIN L, AUGER FA: A completely biological tissue-engineered human blood vessel. *FASEB J* **12**: 47-56, 1998.
- L'HEUREUX N, DUSSERRE N, KONIG G, VICTOR B, KEIRE P, WIGHT TN, CHRONOS NA, KYLES AE, GREGORY CR, HOYT G, ROBBINS RC, McALLISTER TN: Human tissue-engineered blood vessels for adult arterial revascularization. *Nat Med* **12**: 361-365, 2006.
- L'HEUREUX N, DUSSERRE N, MARINI A, GARRIDO S, DE LA FUENTE L, McALLISTER T: Technology insight: the evolution of tissue-engineered vascular grafts – from research to clinical practice. *Nat Clin Pract Cardiovasc Med* **4**: 389-395, 2007a.
- L'HEUREUX N, McALLISTER TN, DE LA FUENTE LM: Tissue-engineered blood vessel for adult arterial revascularization. *N Engl J Med* **357**: 1451-1453, 2007b.
- LEON LR JR, HUGHES JD, PSALMS SB, GUERRA R, BISWAS A, PRASAD A, KROUSE RS: Portomesenteric reconstruction during Whipple procedures: review and report of a case. *Vasc Endovascular Surg* **41**: 537-546, 2007.
- LIM HJ, NAM HY, LEE BH, KIM DJ, KO JY, PARK JS: A novel technique for loading of paclitaxel-PLGA nanoparticles onto ePTFE vascular grafts. *Biotechnol Prog* **23**: 693-697, 2007.
- LINTON RR: Some practical considerations in the surgery of blood vessel grafts. *Surgery* **38**: 817-834, 1955.
- LOPEZ PP, GINZBERG E: Vascular trauma. In: *Surgery: Basic Science and Clinical Evidence*. JA NORTON, PS BARIE, RR BOLLINGER, AE CHANG, S LOWRY, SJ MULVIHILL, HI PASS, RW THOMPSON (eds), Springer, New York, 2008, pp 521-544.
- MAGOMETSCHNIGG H, KADLETZ M, VODRAZKA M, DOCK W, GRIMM M, GRABENWÖGER M, MINAR E, STAUDACHER M, FENZL G, WOLNER E: Clinical study with in vitro endothelial cell lining of expanded polytetrafluoroethylene grafts in crural repeat reconstruction. *J Vasc Surg* **15**: 527-535, 1992.
- MAHMOOD A, GARNHAM A, SINTLER M, SMITH SR, VOHRA RK, SIMMS MH: Composite sequential grafts for femorocrural bypass reconstruction: experience with a modified technique. *J Vasc Surg* **36**: 772-778, 2002.
- MAMODE N, SCOTT RN: Graft type for femoro-popliteal bypass surgery. *Cochrane Database Syst Rev* **2**: CD001487, 2000.
- MAROIS Y, CHAKFÉ N, GUIDOIN R, DUHAMEL RC, ROY R, MAROIS M, KING MW, DOUVILLE Y: An albumin-coated polyester arterial graft: in vivo assessment of biocompatibility and healing characteristics. *Biomaterials* **17**: 3-14, 1996.
- MATIA I, JANOUŠEK L, MARADA T, ADAMEC M: Cold-stored venous allografts in the treatment of critical limb ischaemia. *Eur J Vasc Endovasc Surg* **34**: 424-431, 2007.

- MATIA I, ADAMEC M, VARGA M, JANOUŠEK L, LIPAR K, VIKLICKÝ O: Aortoiliac reconstruction with allograft and kidney transplantation as a one-stage procedure: long term results. *Eur J Vasc Endovasc Surg* **35**: 353-357, 2008.
- MATSUMURA G, MIYAGAWA-TOMITA S, SHIN'OKA T, IKADA Y, KUROSAWA H: First evidence that bone marrow cells contribute to the construction of tissue-engineered vascular autografts in vivo. *Circulation* **108**: 1729-1734, 2003.
- MATSUURA JH, BLACK KS, LEVITT AB, ROSENTHAL D, WELLONS ED, FALLON MT, DAVENPORT CK, GOODMAN CL, PAGELSEN ND, OLLERENSHAW JD: Cellular remodeling of depopulated bovine ureter used as an arteriovenous graft in the canine model. *J Am Coll Surg* **198**: 778-783, 2004.
- McALLISTER TN, MARUSZEWSKI M, GARRIDO SA, WYSTRYCHOWSKI W, DUSSERRE N, MARINI A, ZAGALSKI K, FIORILLO A, AVILA H, MANGLANO X, ANTONELLI J, KOCHER A, ZEMBALA M, CIERPKA L, DE LA FUENTE LM, L'HEUREUX N: Effectiveness of haemodialysis access with an autologous tissue-engineered vascular graft: a multicentre cohort study. *Lancet* **373**: 1440-1446, 2009.
- MEINHART J, DEUTSCH M, ZILLA P: Eight years of clinical endothelial cell transplantation. Closing the gap between prosthetic grafts and vein grafts. *ASAIO J* **43**: M515-M521, 1997.
- MET R, VAN LIENDEN KP, KOELEMAY MJ, BIPAT S, LEGEMATE DA, REEKERS JA: Subintimal angioplasty for peripheral arterial occlusive disease: a systematic review. *Cardiovasc Intervent Radiol* **31**: 687-697, 2008.
- MIRZAI M, SCHMITTO JD, TIRILOMIS T, FATEHPUR S, LIAKOPOULOS OJ, TEUCHER N, DÖRGE H, SCHÖNDUBE FA: Surgical management of vascular graft infection in severely ill patients by partial resection of the infected prosthesis. *Eur J Vasc Endovasc Surg* **33**: 610-613, 2007.
- MOAWAD J, GAGNE P: Adjuncts to improve patency of infrainguinal prosthetic bypass grafts. *Vasc Endovascular Surg* **37**: 381-386, 2003.
- MUTO A, FITZGERALD TN, PIMIENTO JM, MALONEY SP, TESO D, PASZKOWIAK JJ, WESTVIK TS, KUDO FA, NISHIBE T, DARDIK A: Smooth muscle cell signal transduction: implications of vascular biology for vascular surgeons. *J Vasc Surg* **45** (Suppl A): A15-A24, 2007.
- NEZIĆ DG, KNEZEVIĆ AM, MILOJEVIĆ PS, DUKANOVIĆ BP, JOVIĆ MD, BORZANOVIĆ MD, NESKOVIĆ AN: The fate of the radial artery conduit in coronary artery bypass grafting surgery. *Eur J Cardiothorac Surg* **30**: 341-346, 2006.
- NIKLASON LE, GAO J, ABBOTT WM, HIRSCHI KK, HOUSER S, MARINI R, LANGER R: Functional arteries grown in vitro. *Science* **284**: 489-493, 1999.
- NISHIBE T, KONDO Y, MUTO A, DARDIK A: Optimal prosthetic graft design for small diameter vascular grafts. *Vascular* **15**: 356-360, 2007.
- NOISHIKI Y, TOMIZAWA Y, YAMANE Y, MATSUMOTO A: Autocrine angiogenic vascular prosthesis with bone marrow transplantation. *Nat Med* **2**: 90-93, 1996.
- NORGREN L, HIATT WR, DORMANDY JA, NEHLER MR, HARRIS KA, FOWKES FG: TASC II Working Group. Inter-Society Consensus for the Management of Peripheral Arterial Disease (TASC II). *J Vasc Surg* **45** (Suppl S): S5-S67, 2007.
- NORTON L, EISEMAN B: Replacement of portal vein during pancreatectomy for carcinoma. *Surgery* **77**: 280-284, 1975.
- O'BRIEN TP, WALSH MT, KAVANAGH EG, FINN SP, GRACE PA, MCGLOUGHLIN TM: Surgical feasibility study of a novel polytetrafluoroethylene graft design for the treatment of peripheral arterial disease. *Ann Vasc Surg* **21**: 611-617, 2007.
- PAMULA E, BAČÁKOVÁ L, FILOVÁ E, BUCZYNSKA J, DOBRZYNSKI P, NOSKOVÁ L, GRAUSOVÁ L: The influence of pore size on colonization of poly(L-lactide-glycolide) scaffolds with human osteoblast-like MG 63 cells in vitro. *J Mater Sci Mater Med* **19**: 425-435, 2008.
- PANNETON JM, HOLLIER LH, HOFER JM: Multicenter randomized prospective trial comparing a pre-cuffed polytetrafluoroethylene graft to a vein cuffed polytetrafluoroethylene graft for infragenicular arterial bypass. *Ann Vasc Surg* **18**: 199-206, 2004.

- QUARMBY JW, BURNAND KG, LOCKHART SJ, DONALD AE, SOMMERVILLE KM, JAMIESON CW, BROWSE NL: Prospective randomized trial of woven versus collagen-impregnated knitted prosthetic Dacron grafts in aortoiliac surgery. *Br J Surg* **85**: 775-777, 1998.
- RADEMACHER A, PAULITSCHKE M, MEYER R, HETZER R: Endothelialization of PTFE vascular grafts under flow induces significant cell changes. *Int J Artif Organs* **24**: 235-242, 2001.
- RAHLF G, URBAN P, BOHLE RM: Morphology of healing in vascular prostheses. *Thorac Cardiovasc Surg* **34**: 43-48, 1986.
- RÉMY-ZOLGHADRI M, LAGANIÈRE J, OLIGNY JF, GERMAIN L, AUGER FA: Endothelium properties of a tissue-engineered blood vessel for small-diameter vascular reconstruction. *J Vasc Surg* **39**: 613-620, 2004.
- RIEDEL T, BRYNDA E, DYR JE, HOUSKA M: Controlled preparation of thin fibrin films immobilized at solid surfaces. *J Biomed Mater Res A* **88**: 437-447, 2009.
- RICOTTA JJ: Vascular conduits: an overview. In: *Vascular Surgery*. RB RUTHERFORD (ed), Elsevier-Saunders, Philadelphia, 2005, pp 688-695.
- RIHA GM, LIN PH, LUMSDEN AB, YAO Q, CHEN C: Review: application of stem cells for vascular tissue engineering. *Tissue Eng* **11**: 1535-1552, 2005.
- ROLL S, MÜLLER-NORDHORN J, KEIL T, SCHULZ H, EIDT D, GREINER W, WILLICH SN: Dacron vs. PTFE as bypass materials in peripheral vascular surgery – systematic review and meta-analysis *BMC Surgery* **8**: 22, 2008.
- SALACINSKI HJ, TIWARI A, HAMILTON G, SEIFALIAN AM: Cellular engineering of vascular bypass grafts: role of chemical coatings for enhancing endothelial cell attachment. *Med Biol Eng Comput* **39**: 609-618, 2001.
- SARJEANT JM, RABINOVITCH M: Understanding and treating vein graft atherosclerosis. *Cardiovasc Pathol* **11**: 263-271, 2002.
- SARKAR S, SALACINSKI HJ, HAMILTON G, SEIFALIAN AM: The mechanical properties of infrainguinal vascular bypass grafts: their role in influencing patency. *Eur J Vasc Endovasc Surg* **31**: 627-636, 2006.
- SASAKI H: The right gastroepiploic artery in coronary artery bypass grafting. *J Card Surg* **23**: 398-407, 2008.
- SATO S, NITTA Y, SAIKI Y, KAWAMOTO S, IGUCHI A, KAKU M, TABATA Y, TABAYASHI K: Enhanced perigraft angiogenesis prevents prosthetic graft infection. *Ann Thorac Surg* **86**: 1278-1284, 2008.
- SAUVAGE LR: Dacron arterial grafts. In: *Vascular Graft Update: Safety and Performance*. HE KAMBIC, A KANTROWITZ, P SUNG (eds), ASTM Publication, Williamsburg, 1986, pp 16-24.
- SCHMIDT CE, BAIER JM: Acellular vascular tissues: natural biomaterials for tissue repair and tissue engineering. *Biomaterials* **21**: 2215-2231, 2000.
- SCHOUTEN O, HOEDT MT, WITTENS CH, HOP WC, VAN SAMBEEK MR, VAN URK H: VASCAN Study Group. End-to-end versus end-to-side distal anastomosis in femoropopliteal bypasses; results of a randomized multicenter trial. *Eur J Vasc Endovasc Surg* **29**: 457-462, 2005.
- SCHWARZBACH MH, HORMANN Y, HINZ U, LEOWARDI C, BÖCKLER D, MECHTERSHEIMER G, FRIESS H, BÜCHLER MW, ALLENBERG JR: Clinical results of surgery for retroperitoneal sarcoma with major blood vessel involvement. *J Vasc Surg* **44**: 46-55, 2006.
- SCOTT EC, GLICKMAN MH: Conduits for hemodialysis access. *Semin Vasc Surg* **20**: 158-163, 2007.
- SEIFALIAN AM, TIWARI A, HAMILTON G, SALACINSKI HJ: Improving the clinical patency of prosthetic vascular and coronary bypass grafts: the role of seeding and tissue engineering. *Artif Organs* **26**: 307-320, 2002.
- SEIFALIAN AM, SALACINSKI HJ, TIWARI A, EDWARDS A, BOWALD S, HAMILTON G: In vivo biostability of a poly(carbonate-urea)urethane graft. *Biomaterials* **24**: 2549-2557, 2003.
- SHI Q, WU MH, ONUKI Y, GHALI R, HUNTER GC, JOHANSEN KH, SAUVAGE LR: Endothelium on the flow surface of human aortic Dacron vascular grafts. *J Vasc Surg* **25**: 736-742, 1997.
- SHIN H, JO S, MIKOS AG: Biomimetic materials for tissue engineering. *Biomaterials* **24**: 4353-4364, 2003.
- SHIN'OKA T, IMAI Y, IKADA Y: Transplantation of a tissue-engineered pulmonary artery. *N Engl J Med* **344**: 532-533, 2001.

- SHIN'OKA T, MATSUMURA G, HIBINO N, NAITO Y, WATANABE M, KONUMA T, SAKAMOTO T, NAGATSU M, KUROSAWA H: Midterm clinical result of tissue-engineered vascular autografts seeded with autologous bone marrow cells. *J Thorac Cardiovasc Surg* **129**: 1330-1338, 2005.
- SIEGMAN FA: Use of the venous cuff for graft anastomosis. *Surg Gynecol Obstet* **148**: 930, 1979.
- SITZMANN JV, IMBEMBO AL, RICOTTA JJ, McMANAMA GP, HUTCHINS GM: Dimethylsulfoxide-treated, cryopreserved venous allografts in the arterial and venous systems. *Surgery* **95**: 154-159, 1984.
- SRIRAM V, PATTERSON C: Cell cycle in vasculoproliferative diseases: potential interventions and routes of delivery. *Circulation* **103**: 2414-2419, 2001.
- STARZL TE: A trip south. In: *The Puzzle People: Memoirs of a Transplant Surgeon*. TE STARZL (ed), University of Pittsburgh Press, Pittsburgh, 2003, pp 47-58.
- STEHOUWER CD, CLEMENT D, DAVIDSON C, DIEHM C, ELTE JW, LAMBERT M, SERENI D, the EFIM Vascular Medicine Working Group: Peripheral arterial disease: a growing problem for the internist. *Eur J Intern Med* **20**: 132-138, 2009.
- STEWART GJ, ESSA N, CHANG KH, REICHLER FA: A scanning and transmission electron microscope study of the luminal coating on Dacron prostheses in the canine thoracic aorta. *J Lab Clin Med* **85**: 208-226, 1975.
- SWENNE CL, BOROWIEC J, CARLSSON M, LINDHOLM C: Prediction of and risk factors for surgical wound infection in the saphenous vein harvesting leg in patients undergoing coronary artery bypass. *Thorac Cardiovasc Surg* **54**: 300-306, 2006.
- TIWARI A, SALACINSKI HJ, HAMILTON G, SEIFALIAN AM: Tissue engineering of vascular bypass grafts: role of endothelial cell extraction. *Eur J Vasc Endovasc Surg* **21**: 193-201, 2001.
- TIWARI A, SALACINSKI H, SEIFALIAN AM, HAMILTON G: New prostheses for use in bypass grafts with special emphasis on polyurethanes. *Cardiovasc Surg* **10**: 191-197, 2002.
- TIWARI A, CHENG KS, SALACINSKI H, HAMILTON G, SEIFALIAN AM: Improving the patency of vascular bypass grafts: the role of suture materials and surgical techniques on reducing anastomotic compliance mismatch. *Eur J Vasc Endovasc Surg* **25**: 287-295, 2003.
- VARA DS, SALACINSKI HJ, KANNAN RY, BORDENAVE L, HAMILTON G, SEIFALIAN AM: Cardiovascular tissue engineering: state of the art. *Pathol Biol (Paris)* **53**: 599-612, 2005.
- VARU VN, TSIHLIS ND, KIBBE MR: Basic science review: nitric oxide-releasing prosthetic materials. *Vasc Endovascular Surg* **43**: 121-131, 2009.
- VOORHEES AB Jr, JARETZKI A, BLAKEMORE AH: The use of tubes constructed from vinyon "N" cloth in bridging arterial defects. *Ann Surg* **135**: 332-336, 1952.
- VROMAN L, ADAMS AL: Identification of rapid changes at plasma-solid interfaces. *J Biomed Mater Res* **3**: 43-67, 1969.
- WALPOTH BH, ROGULENKO R, TIKHVINSKAIA E, GOGOLEWSKI S, SCHAFFNER T, HESS OM, ALTHAUS U: Improvement of patency rate in heparin-coated small synthetic vascular grafts. *Circulation* **98** (Suppl II): II-319-II-324, 1998.
- WATSON HR, BELCHER G, HORROCKS M: Adjuvant medical therapy in peripheral bypass surgery. *Br J Surg* **86**: 981-991, 1999.
- WEINBERG CB, BELL E: A blood vessel model constructed from collagen and cultured vascular cells. *Science* **231**: 397-400, 1986.
- WISSELINK W, MONEY SR, BECKER MO, RICE KL, RAMEE SR, WHITE CJ, KAZMIER FJ, HOLLIER LH: Comparison of operative reconstruction and percutaneous balloon dilatation for central venous obstruction. *Am J Surg* **166**: 200-205, 1993.
- WU MH, KOUCHI Y, ONUKI Y, SHI Q, YOSHIDA H, KAPLAN S, VIGGERS RF, GHALI R, SAUVAGE LR: Effect of differential shear stress on platelet aggregation, surface thrombosis, and endothelialization of bilateral carotid-femoral grafts in the dog. *J Vasc Surg* **22**: 382-390, 1995.
- XUE L, GREISLER HP: Biomaterials in the development and future of vascular grafts. *J Vasc Surg* **37**: 472-480, 2003.
- YOSHIZUMI M, ABE J, TSUCHIYA K, BERK BC, TAMAKI T: Stress and vascular responses: atheroprotective effect of laminar fluid shear stress in endothelial cells: possible role of mitogen-activated protein kinases. *J Pharmacol Sci* **91**: 172-176, 2003.

- ZACHARIAS RK, KIRKMAN TR, CLOWES AW: Mechanisms of healing in synthetic grafts. *J Vasc Surg* **6**: 429-436, 1987.
- ZARBIV G, PREIS M, BEN-YOSEF Y, FLUGELMAN MY: Engineering blood vessels by gene and cell therapy. *Expert Opin Biol Ther* **7**: 1183-1191, 2007.
- ZDRAHALA RJ: Small caliber vascular grafts. Part II: Polyurethanes revisited. *J Biomater Appl* **11**: 37-61, 1996.
- ZHANG Z, MAROIS Y, GUIDOIN RG, BULL P, MAROIS M, HOW T, LAROCHE G, KING MW: Vascugraft polyurethane arterial prosthesis as femoro-popliteal and femoro-peroneal bypasses in humans: pathological, structural and chemical analyses of four excised grafts. *Biomaterials* **18**: 113-124, 1997.
- ZILLA P, BEZUIDENHOUT D, HUMAN P: Prosthetic vascular grafts: wrong models, wrong questions and no healing. *Biomaterials* **28**: 5009-5027, 2007.
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