

## Pharmacokinetics of Boldine in Control and Mrp2-Deficient Rats

J. CERMANOVA<sup>1</sup>, A. PRASNICKA<sup>1</sup>, E. DOLEZELOVA<sup>3</sup>, L. ROZKYDALOVA<sup>1,4</sup>,  
M. HROCH<sup>2</sup>, J. CHLÁDEK<sup>1</sup>, P. TOMSIK<sup>2</sup>, I. KLOETING<sup>5</sup>, S. MICUDA<sup>1</sup>

<sup>1</sup>Department of Pharmacology, <sup>2</sup>Department of Medical Biochemistry, Faculty of Medicine, Charles University, Hradec Kralove, Czech Republic, <sup>3</sup>Department of Biological and Medical Sciences, <sup>4</sup>Department of Pharmacology and Toxicology, Faculty of Pharmacy, Charles University, Hradec Kralove, Czech Republic, <sup>5</sup>The Institute of Pathophysiology, University Medicine Greifswald, Karlsburg, Germany

Received October 1, 2016

Accepted October 22, 2016

### Summary

The aim of the present study was to describe the currently poorly understood pharmacokinetics (PK) of boldine in control rats (LW, Lewis rats), and Mrp2 transporter-deficient rats (TR). Animals from the LW and TR groups underwent a bolus dose study with 10 mg/kg of boldine applied either orally or intravenously in order to evaluate the major PK parameters. The TR rats demonstrated significantly reduced total clearance with prolonged biological half-life (LW 12±4.6 versus TR 20±4.4 min), decreased volume of distribution (LW 3.2±0.4 l/kg versus TR 2.4±0.4 l/kg) and reduced bioavailability (LW 7 % versus TR 4.5 %). Another set of LW and TR rats were used for a clearance study with continuous intravenous administration of boldine. The LW rats showed that biliary and renal clearance formed less than 2 % of the total clearance of boldine. The treatment of samples with β-glucuronidase showed at least a 38 % contribution of conjugation reactions to the overall clearance of boldine. The TR rats demonstrated reduced biliary clearance of boldine and its conjugates, which was partly compensated by their increased renal clearance. In conclusion, this study presents the PK parameters of boldine and shows the importance of the Mrp2 transporter and conjugation reactions in the elimination of the compound.

### Key words

Boldine • Mrp2 • Pharmacokinetics • Elimination • Bioavailability

### Corresponding author

S. Micuda, Department of Pharmacology, Charles University, Faculty of Medicine, Simkova 870, 500 03 Hradec Kralove, Czech Republic. E-mail: micuda@lfhk.cuni.cz

### Introduction

Boldine is an alkaloid isolated from the leaf and bark of the Chilean Boldo tree (*Peumus boldus* Molina, Monimiaceae). The agent has shown positive effects on several preclinical *in vitro* and *in vivo* models of different pathologies such as hepatotoxicity (Lanhers *et al.* 1991, Fernandez *et al.* 2009, Zagorova *et al.* 2015), atherosclerosis (Santanam *et al.* 2004), diabetic nephropathy (Hernandez-Salinas *et al.* 2013), altered GIT motility (Muthna *et al.* 2013) and malignant or inflammatory diseases (Backhouse *et al.* 1994, Tomsik *et al.* 2016). These effects were the result of the antioxidant, anti-inflammatory, anti-proliferative, prokinetic, choleric and anti-infective action of boldine (O'Brien *et al.* 2006, Muthna *et al.* 2013). The majority of these pathologies demand long-term repeated administration, preferably orally. Information about bioavailability (BAV), the achievement of the desired concentration in plasma, distribution and elimination, i.e. pharmacokinetics (PK), is therefore crucial for further effective use of boldine. In contrast to detailed data about the effects/pharmacodynamics, much less is known about the PK of the agent.

In the few studies conducted so far, boldine plasma and tissue concentration have been analysed in rats (Jimenez and Speisky 2000, Hroch *et al.* 2013). The results of these studies consistently showed a rapid decline in boldine plasma concentrations with a terminal half-life of 31 min. Other PK parameters that describe BAV, distribution or metabolism and excretion are

missing. Only Jimenez *et al.* (2000) have provided more specific data. Their study demonstrated that treatment of urine samples with  $\beta$ -glucuronidase increased the recovery of boldine three to four-fold. This produced indirect evidence of the extensive formation of boldine glucuronides. Following this, more detailed analysis through the use of the LC-MS method confirmed glucuronide and sulfate conjugates as the major metabolites of boldine in rats (Hroch *et al.* 2013). Our group recently showed that concentrations of boldine in bile exceed its concentrations in plasma (Hroch *et al.* 2013, Cermanova *et al.* 2015). This means that the agent is actively concentrated in bile by an ABC transporter on the apical membrane of hepatocytes. Considering substrate spectra, sulfate and glucuronide conjugated metabolites are typically transported by multidrug resistance-associated protein 2 (Mrp2) (Nies and Keppler 2007). The role of Mrp2 in the kinetics of boldine is unknown. Our previous experiments with the infusion of boldine into Mrp2-deficient rats did not show a discrepancy in the choleric activity of the compound or an inhibitory effect on Mrp2 in MDCK-MRP2 cells (Cermanova *et al.* 2015). It is therefore important to perform a direct kinetic study in Mrp2-deficient animals to uncover the exact role of Mrp2 in the disposition of boldine.

The aim of this study was to describe the PK of boldine with the specification of individual parameters in control and Mrp2-deficient rats. Special attention was paid to the BAV of the agent and its elimination by hepatic and renal pathways. The contribution of glucuronidation and sulfation to these processes was determined by the incubation of plasma, bile and urine samples with  $\beta$ -glucuronidase, which also contained sulfatase.

## Materials and Methods

### Chemicals

Boldine was purchased from Sigma-Aldrich (St. Louis, MO, US) and dissolved in distilled water (pH adjusted by HCl to 1.0) with consequent titration to pH 7.0 by NaOH. The stock solution of boldine was further dissolved in saline for oral and intravenous administration to rats.  $\beta$ -glucuronidase Type HP2 from *Helix pomatia* was purchased from Sigma-Aldrich (Cat. no. G7017, St. Louis, MO, US).

### Animals

Mrp2-deficient Lewis transport-deficient rats (TR<sup>-</sup>) or complementary Lewis (LW) rats (Institut für Pathophysiologie, Karlsburg, Germany) were used throughout the study. The animals were housed within controlled environmental conditions. All experiments were performed in accordance with the "Guide for the Care and Use of Laboratory Animals" published by the US National Institutes of Health (NIH publication, 1996) and under the supervision of the Ethical Committee of the Faculty of Medicine in Hradec Kralove.

### Experimental design

The examination of BAV was performed in 12 TR<sup>-</sup> and 12 LW rats. Animals from each strain were randomised into two groups (six animals per group). After 12 hours of fasting, one group of LW and TR<sup>-</sup> rats received boldine by i.v. bolus (10 mg/kg of bw) and another group of LW and TR<sup>-</sup> rats were applied with the same dose orally *via* intragastric probe. Blood samples (300  $\mu$ l) were taken from the orbital plexus in short inhaled anesthesia at 4, 10, 30, 60, 120 and 180 min after the administration of boldine. Plasma was obtained by immediate centrifugation at 3000 g and stored at -80 °C. At the end of the experiment, the rats were sacrificed by exsanguination from the carotid artery.

### Clearance study

The evaluation of biliary and urinary excretion of boldine was performed in LW and TR<sup>-</sup> rats after 12 hour of fasting (one group each, six rats per group) during general anaesthesia induced by pentobarbital (50 mg/kg, intraperitoneally). The rats were fixed in a supine position on a heated platform to maintain body temperature at 37 °C and the v. jugularis (for substance administration), carotid artery (for blood sampling), bile duct (bile collection) and urine bladder (for urine collection) were cannulated. After an initial 30-min stabilisation period, the study began with boldine i.v. bolus of 15 mg/kg applied over 10 min followed by an i.v. infusion of boldine (50 mg/kg/h). A parallel evaluation of the glomerular filtration rate was enabled by the simultaneous administration of sinistrin (1.62 mg/kg/h). The infused volume was 6 ml/kg/h to replace fluid losses. Bile and urine were collected in pre-weighted tubes at 30-min intervals. Blood samples were taken in the middle of the collection intervals. At the end of the experiment, the rats were sacrificed by exsanguination from the carotid artery and samples of

plasma, bile, urine, liver and kidney were snap frozen in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  until analysis.

#### *Western blot*

This method was performed as described previously (Kadova *et al.* 2015). Briefly, crude membrane fraction from the liver and the kidney was prepared by ultracentrifugation (100000 g for 1 h) of supernatant obtained by centrifugation (3000 g for 10 min) of tissue homogenate. Samples were then separated on polyacrylamide gels, transferred to PVDF membrane and processed by primary and secondary antibodies with consequent chemiluminescent detection. GAPDH was used as a loading control.

#### *Analytical methods*

Boldine was detected by the previously described HPLC method (Hroch *et al.* 2013). For the identification of phase II boldine metabolites (conjugates) in body fluids, 50  $\mu\text{l}$  of urine or bile (250  $\mu\text{l}$  plasma) was mixed with 50  $\mu\text{l}$  (250  $\mu\text{l}$  plasma) of acetate buffer (pH 5; 59 ml of 0.2 M acetic acid and 141 ml of 0.2 M sodium acetate) and 50  $\mu\text{l}$  of  $\beta$ -glucuronidase (or only acetate buffer pH 5) was added. The mixture was incubated at  $37^{\circ}\text{C}$  for 16 h (Nobilis *et al.* 2004). After centrifugation (10000 g for 5 min), the diluted samples (plasma) were directly injected into the chromatographic column. Samples of bile and urine were then diluted with mobile phase and analysed for boldine.

#### *PK analysis*

The calculation of the PK parameters after the bolus i.v. dose of boldine was completed by a non-compartmental analysis of plasma concentration versus time curves using Kinetica software (Thermo Fisher Scientific, Inc.), as reported previously (Laho *et al.* 2016). The evaluation of biliary and urinary excretion of boldine was performed during constant i.v. infusion of the agent. Biliary and urinary excretion was calculated by multiplication of bile/urine flow (ml/min) with the concentration of boldine in the sample (measured before and after treatment with  $\beta$ -glucuronidase). Clearance parameters were evaluated during steady-state of plasma concentrations, which was reached between 60-120 min of the clearance study. The calculation was based on the division of the rate of administration (for total clearance) or biliary/renal excretion (for biliary/renal clearance) by the steady-state concentration of boldine in plasma. The glomerular filtration rate was calculated as the total

clearance of sinistrin during the clearance study by the division of the rate of administration by its plasma concentrations.

#### *Statistical analysis*

The results are presented as mean  $\pm$  SD. Students't-test was employed for a two-group comparison. Differences were considered significant at a *p*-value of less than 0.05. All analyses were performed using GraphPad Prism 6.0 software (San Diego, US).

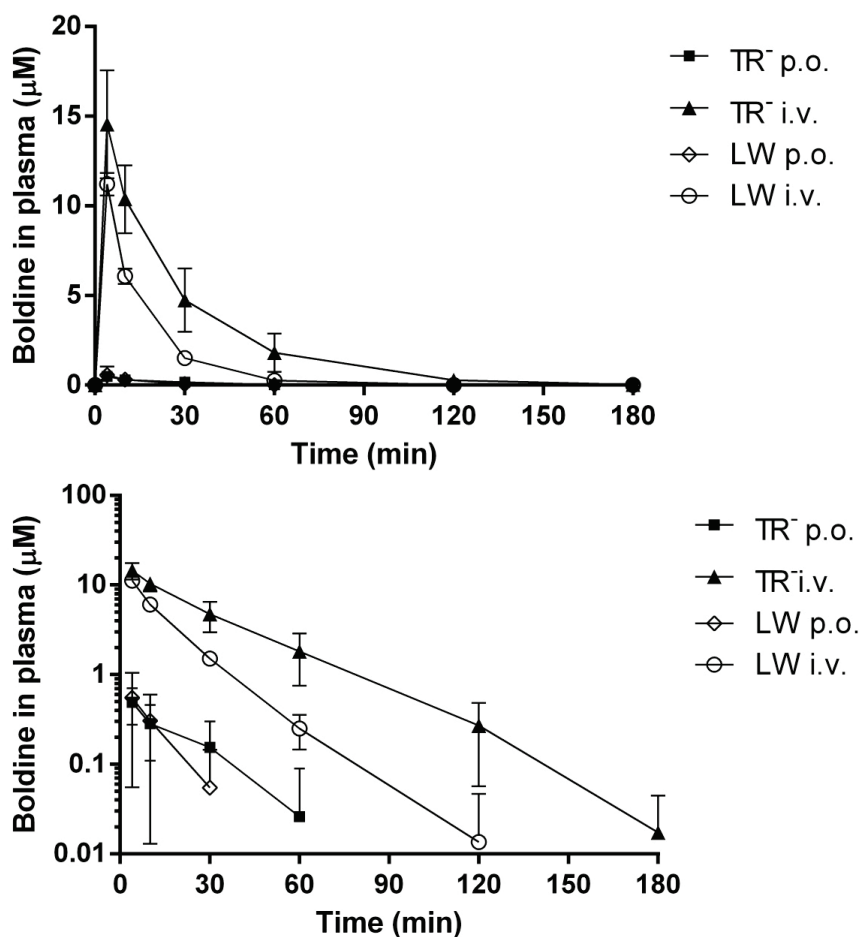
## **Results**

#### *PK parameters of boldine after bolus administration to control LW animals*

Oral and i.v. bolus doses of boldine yielded markedly different profiles of concentrations in plasma of the LW animals (Fig. 1). The ratio of areas under the curve of plasma concentrations ( $\text{AUC}_{0-\infty}$ ) pointed to very low BAV (Table 1). The concentration in plasma fell rapidly below the limit of detection, especially in animals who received oral administration. An analysis of the PK parameters was therefore possible only in i.v.-applied animals. Extrapolated AUC from the last measured concentration till infinity was below 2 %, which supports the optimal duration of evaluation and guarantees the accuracy of further calculations. Short half-life ( $T_{1/2}$ ) emphasized the rapid elimination of boldine, as confirmed by the high total clearance of the agent. The profile of plasma concentrations during elimination suggested first order kinetic with single compartment distribution, i.e. avid equilibrium between plasma and tissues. This was also supported by the rapid attainment of maximum concentration ( $C_{\text{max}}$ ) within 4 min, even after oral administration. The distribution volume above volume of total body water (Table 1) indicated the effective intracellular penetration of boldine and complied with its ability to perform an anti-oxidative effect within the cells.

#### *PK parameters after bolus administration to Mrp2-deficient TR<sup>-</sup> rats*

The plasma concentration versus time plots after i.v. administration of boldine in TR<sup>-</sup> rats demonstrated a significant increase in  $C_{\text{max}}$  and the prolongation of the elimination phase in comparison with LW animals (Fig. 1). Consequent PK analysis confirmed that Mrp2 deficiency increased  $\text{AUC}_{0-\infty}$ , prolonged  $T_{1/2}$  mean residence time (MRT) and reduced total clearance and



**Fig. 1.** Plasma concentration time curves of boldine administered (10 mg/kg) orally or intravenously to LW, control and Mrp2-deficient TR<sup>-</sup> rats. Data are means  $\pm$  SD (six animals per group).

distribution volume of boldine. Although a tendency towards the prolongation of the elimination period was also evident after the oral administration of boldine in TR<sup>-</sup> rats (Fig. 1), the interindividual variability precluded the attainment of a statistically significant change of  $AUC_{0-\infty}$  when compared with oral administration in LW rats. Thus, the ration of  $AUC_{0-\infty}$  after oral and intravenous administration of boldine showed reduced bioavailability of the agent in TR<sup>-</sup> rats when compared to control LW rats as a consequence of prolonged elimination.

#### *Elimination of boldine in heathy LW rats*

A constant rate infusion of boldine with an analysis of urine and bile in LW rats showed preferential urinary excretion of boldine (Table 2). The ratio of renal clearance to the glomerular filtration rate markedly below one uncovered active reabsorption of the parent compound in renal tubules. The sum of biliary and renal clearances formed less than 2% of the total clearance, indicating the significant contribution of metabolism to the total clearance of boldine. The

consequent incubation of all samples with  $\beta$ -glucuronidase markedly increased the recovery of boldine and pointed to mainly biliary excretion of boldine conjugates. The total clearance of boldine conjugates approached 38% of the total clearance of the parent compound.

#### *Elimination of boldine in TR<sup>-</sup> rats*

The absence of Mrp2 in TR<sup>-</sup> rats led to a marked reduction of biliary excretion and clearance of boldine during its infusion, while its urinary excretion and clearance was increased. As a consequence, the steady-state plasma concentration and total clearance of parent compound were not changed in TR<sup>-</sup> rats when compared with LW animals. In comparison, biliary excretion and the clearance of boldine conjugates were nearly abolished in TR<sup>-</sup> rats. Increased urinary excretion was not sufficient to compensate for the reduced biliary excretion of boldine conjugates. Thus, TR<sup>-</sup> rats showed reduced total clearance of conjugated boldine metabolites with their accumulation in plasma.

**Table 1.** The effect of an Mrp2 transporter deficit on the primary PK parameters of boldine after i.v. administration in a dose of 10 mg/kg.

	LW	TR <sup>-</sup>
$C_{max}$ ( $\mu M$ )	11±0.6	15±3.0*
$AUC_{0-\infty}$ (mg/ml/min)	0.05±0.01	0.13±0.04**
$AUC_{extra}$ (%)	1.9±0.7	1.7±1.1
$T_{1/2}$ (min)	12±4.6	20±4.4*
MRT (min)	17±2.9	30±6.6**
$CL_{Tot}$ (ml/min/kg)	187±17	88±40***
$V_d$ (l/kg)	3.2±0.4	2.4±0.4**
BAV (%)	7	4.5

LW – control group; TR<sup>-</sup> – Mrp2-deficient group;  $C_{max}$  – peak plasma concentration;  $AUC_{0-\infty}$  – area under the plasma concentration curve extrapolated to infinity;  $AUC_{extra}$  – % of extrapolation of  $AUC_{0-\infty}$  from last measured concentration to infinity;  $T_{1/2}$  – elimination half-life; MRT – mean residence time of a drug in the body;  $CL_{Tot}$  (ml/min/kg) – total clearance;  $V_d$  – apparent volume of distribution; BAV – bioavailability of the drug. Data are presented as means ± SD from groups of six animals significantly different from the control group (\*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ ).

**Table 2.** Comparison between renal and biliary clearance of boldine and its conjugates with glucuronic acid and sulphate during i.v. infusion.

	Boldine		Boldine conjugates	
	LW	TR <sup>-</sup>	LW	TR <sup>-</sup>
$C_{SS}$ ( $\mu M$ )	29±5.5	33±6.6	80±16	150±56*
BE (nmol/min/kg)	3.4±0.3	1.7±0.7***	240±46	29±18***
UE (nmol/min/kg)	13±7.7	26±12*	68±36	270±100**
$CL_B$ (ml/min/kg)	0.12±0.03	0.06±0.03**	3.1±0.73	0.23±0.17***
$CL_R$ (ml/min/kg)	0.42±0.24	0.75±0.28*	0.91±0.58	1.8±0.6*
$CL_R/GFR$	0.17±0.12	0.24±0.15	0.3±0.1	0.5±0.12*
$CL_{Tot}$ (ml/min/kg)	53±9.1	49±9.2	20±3.5	11±3.5**

LW – control group; TR<sup>-</sup> – Mrp2-deficient group;  $C_{SS}$  – steady-state plasma concentration; BE – biliary excretion; UE – urinary excretion;  $CL_B$  – biliary clearance;  $CL_R$  – renal clearance;  $CL_{Tot}$  – total body clearance; GFR – glomerular filtration rate. Data are presented as means ± SD from groups of six animals significantly different from the control group (\*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ ).

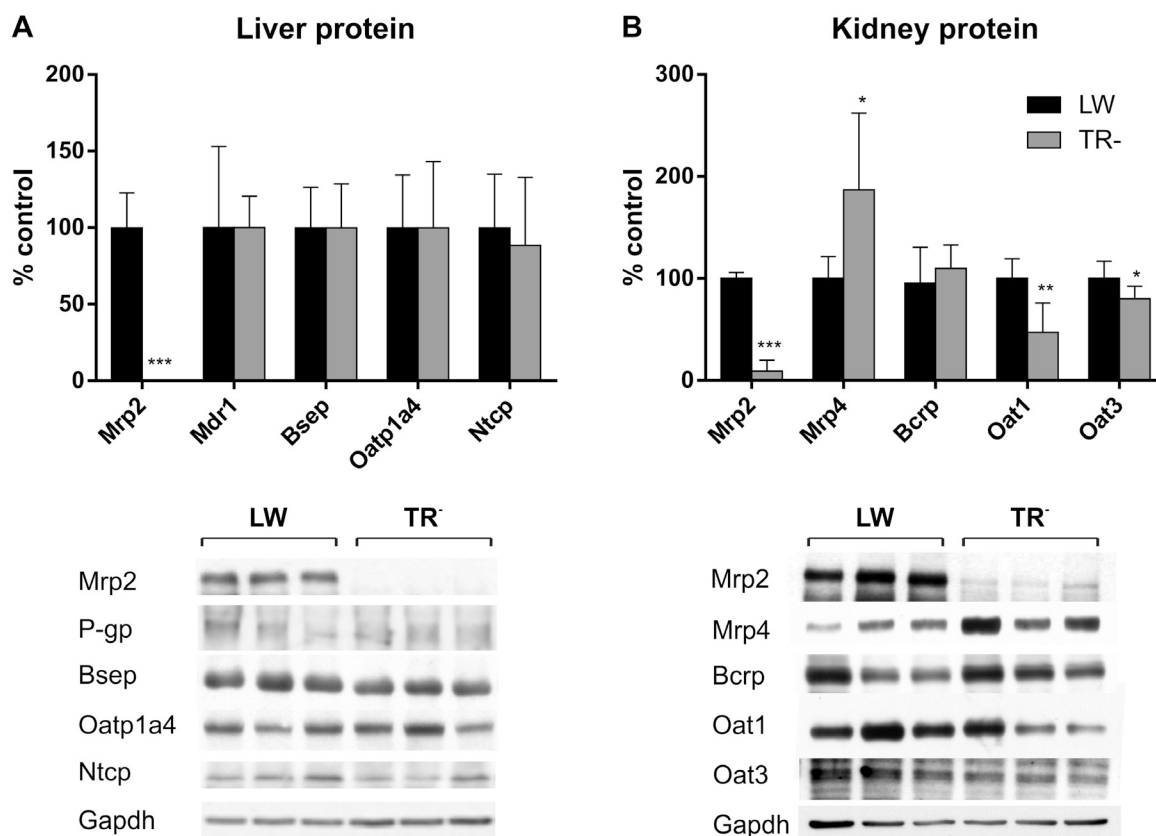
#### Protein expression of selected transporters in the liver and kidney

Western blot analysis of the major drug-transporting proteins showed the sole absence of Mrp2 in the liver of TR<sup>-</sup> rats without influence on other proteins (Fig. 2A). Mrp2 was also nearly absent in kidneys but herein we also detected a reduced expression of uptake Oat1 and Oat3 transporters and an increased expression of Mrp4 (Fig. 2B).

## Discussion

Bioavailability is an essential PK parameter describing the proportion of the dose applied by

extravascular administration that reaches systemic circulation in an active form. Knowledge of these parameters is extremely important for agents, which are intended for mainly oral application. The value of BAV for boldine was not known. In the only study presenting plasma concentration versus time plots of boldine in rats after oral or i.v. administration, the authors used different doses and different timings for blood sampling for either application route (Jimenez and Speisky 2000). Only a rough comparison was possible between the administration of 20 mg/kg intravenously and 25 mg/kg of boldine orally. Under these settings, average  $C_{max}$  after intravenous administration was approximately 31  $\mu M$ , while the oral dose yielded a concentration of 7  $\mu M$ .



**Fig. 2.** Protein expression of selected drug transporters in the liver (**A**) and kidneys (**B**) of LW and TR<sup>-</sup> rats. Bands present representative analysis of six animals per group. Asterisks show significant difference from control group (\*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ ).

Within 60 min of administration, plasma concentrations declined to 5  $\mu\text{M}$  and to 3  $\mu\text{M}$  after intravenous and oral administration, respectively. Differences in  $C_{\text{max}}$  therefore suggest poor absorption of boldine from GIT. Our data from the administration of 10 mg/kg by both routes and the standardization of blood sampling yielded very low oral bioavailability of 7 %. The exact cause of low BAV of boldine is unknown. A major problem seems to be the reduced water solubility of the agent at a physiological or slightly basic pH in the intestine. The agent is most soluble in an acidic environment, as detected in dissolution experiments during the optimization of administration (unpublished observation). This may explain the extremely rapid absorption of boldine from the stomach after its gastric gavage when  $C_{\text{max}}$  was attained within 4 min. An important implication of low BAV of boldine in terms of further oral administration is that the agent must be applied in sufficiently large doses to reach the plasma and tissue concentrations required for its health-promoting effect. Necessary concentrations for its intense free radical scavenging properties are 10-50  $\mu\text{M}$  (Muthna *et al.* 2013). Together with low BAV, this means that a boldine oral dose of at least 10 mg

per kg of BW must be applied to produce the chance of a sufficient pharmacodynamic response. A combination of large doses with low BAV may consequently produce high local concentrations of boldine in GIT. Furthermore, the intensive topical effect of rectally-applied boldine was demonstrated by the suppression of inflammation in experimental colitis (Gotteland *et al.* 1997).

The PK parameters and the routes of boldine elimination from organisms were completely unknown. The only available data showed a very short plasma elimination half-life of 31 min (Jimenez and Speisky 2000). The administration of boldine to control LW animals in our study demonstrated an even shorter half-life of 12 min. An explanation for such a difference is still needed. The difference may be the effect of strain difference when compared with previously used Wistar rats. We also note that i.v. sampling under short anesthesia in our study reduced the stress of animals compared to immobilisation used in the previous study. The rapid elimination of boldine after the i.v. bolus injection corresponded with its high total clearance. However, the bile and urine sampling indicated that bile and renal clearance of the parent compound formed only

1 % of the total clearance. The study showed that boldine is mainly metabolized. Previous studies detected glucuronide and/or sulfate conjugates as major metabolites of boldine (Hroch *et al.* 2013). Indeed, treatment of samples with  $\beta$ -glucuronidase also containing sulfatase increased the recovery of boldine, particularly from bile. The clearance of boldine by these enzymatic pathways consequently formed at least 38 % of the total clearance of the compound. The pathway eliminating the remaining part of the administered dose is unknown. However, the major drawback of the study is that the exact concentration of glucuronide and sulfate conjugates could not be exactly measured due to the unavailability of the LC-MS/MS method. Therefore our data may have underestimated the contribution of conjugation to overall clearance. On the other hand, the method was able to compare the mutual contribution of biliary and renal clearance and showed preferential renal excretion of the parent compound. The conjugated metabolites revert this ration in favor of biliary excretion, pointing to their higher affinity for apical transporting proteins in hepatocytes. An interesting finding was also the marked reabsorption of boldine, as well as its conjugates, in kidney tubules. Whether this effect is carried out by passive reabsorption or active transport deserves further research.

Sulfate and glucuronide conjugates, which seem to be a major elimination pathway for boldine (Hroch *et al.* 2013), are typically transported by the Mrp2 protein (Nies and Keppler 2007). We therefore compared the kinetics of boldine between control and Mrp2-deficient animals. A significant prolongation of the elimination phase after the bolus i.v. administration of boldine and its reduced biliary excretion during infusion in TR<sup>-</sup> rats confirmed the important role of Mrp2 in the elimination of boldine itself. Moreover, the incubation of samples with glucuronidase/sulfatase uncovered a massive reduction in biliary secretion of these boldine conjugates in Mrp2-deficient animals and showed that Mrp2 is a major transporter for these metabolites in the liver. The opposite occurred in the kidneys, where an Mrp2 deficit led to increased urinary excretion and renal clearance of boldine as well as its conjugates. This increase corresponds with the up-regulation of the Mrp4 transporter seen in current and previous studies (Chen *et al.* 2005). The shared substrate specificity between Mrp2 and Mrp4 (Deeley *et al.* 2006, Zhou *et al.* 2008) supports the role of Mrp4 in the elimination of boldine and its metabolites.

Another factor that must be considered is

a possible change in the activity of glucuronosyltransferases in Mrp2-deficient rats. Johnson *et al.* (2006) reported a 3.5-5.5-fold higher expression of the UGT1A enzyme family proteins in the liver and kidney of TR<sup>-</sup> rats compared to wild-type rats. This mechanism is supported by markedly increased plasma concentrations of boldine conjugates and a rise in their urinary excretion. In addition, the Mrp3 basolateral protein, which serves as a cytoprotective mechanism by transporting Mrp2 substrates including conjugates with glucuronic acid from hepatocytes and proximal tubular cells back to blood, is also augmented in the liver and kidney of TR<sup>-</sup> animals, as previously reported (Johnson *et al.* 2006).

The Mrp2 transporter is also localised at the apical membrane of enterocytes and it reduces the absorption of its substrates. The absence of this transporter is therefore associated with the increased bioavailability of such compounds (Dahan and Amidon 2009, Zamek-Gliszczynski *et al.* 2012). However, Mrp2 deficiency does not seem to modify the absorption of boldine from GIT because TR<sup>-</sup> rats demonstrated reduced bioavailability. We suggest that this reduction rose from unchanged AUC<sub>0-∞</sub> after oral administration between control and TR<sup>-</sup> animals and increased AUC<sub>0-∞</sub> due to the prolongation of the elimination phase in TR<sup>-</sup> animals after the i.v. bolus of boldine. Comparable AUC<sub>0-∞</sub> after oral administration in LW and TR<sup>-</sup> rats indicates the involvement of other intestinal transporters in the absorption of boldine. Indeed, Johnson *et al.* (2006) described marked down-regulation of the Mrp3 protein in the jejunum and ileum of TR<sup>-</sup> rats. Mrp3 is an efflux transporter at the basolateral membrane of enterocytes, which facilitates the absorption of compounds from GIT. It is therefore possible that similar substrate specificity with Mrp2 also enables the transportation of boldine by Mrp3 and that the reduction of Mrp3 in enterocytes compensates for the absence of Mrp2.

In conclusion, our study has demonstrated for the first time low bioavailability, intracellular distribution and quantification of mutual hepatic and renal elimination of boldine in rats. We have also noted the significant role of Mrp2 and conjugation with glucuronic acid and sulphates in the overall PK of boldine. The results also indicated that boldine may be a substrate for other Mrp members, namely Mrp4.

### Conflict of Interest

There is no conflict of interest.

## Acknowledgements

This study was supported by grants from the Grant

Agency of Charles University Prvok P37/05 and SVV-2016-260287.

## References

- BACKHOUSE N, DELPORTE C, GIVERNAU M, CASSELS BK, VALENZUELA A, SPEISKY H: Anti-inflammatory and antipyretic effects of boldine. *Agents Actions* **42**: 114-117, 1994.
- CERMANOVA J, KADOVA Z, ZAGOROVA M, HROCH M, TOMSIK P, NACHTIGAL P, KUDLACKOVA Z, PAVEK P, DUBECKA M, CECKOVA M, STAUD F, LAHO T, MICUDA S: Boldine enhances bile production in rats via osmotic and farnesoid X receptor dependent mechanisms. *Toxicol Appl Pharmacol* **285**: 12-22, 2015.
- CHEN C, SLITT AL, DIETER MZ, TANAKA Y, SCHEFFER GL, KLAASSEN CD: Up-regulation of Mrp4 expression in kidney of Mrp2-deficient TR- rats. *Biochem Pharmacol* **70**: 1088-1095, 2005.
- DAHAN A, AMIDON GL: Small intestinal efflux mediated by MRP2 and BCRP shifts sulfasalazine intestinal permeability from high to low, enabling its colonic targeting. *Am J Physiol Gastrointest Liver Physiol* **297**: G371-G377, 2009.
- DEELEY RG, WESTLAKE C, COLE SP: Transmembrane transport of endo- and xenobiotics by mammalian ATP-binding cassette multidrug resistance proteins. *Physiol Rev* **86**: 849-899, 2006.
- FERNANDEZ J, LAGOS P, RIVERA P, ZAMORANO-PONCE E: Effect of boldo (*Peumus boldus* Molina) infusion on lipoperoxidation induced by cisplatin in mice liver. *Phytother Res* **23**: 1024-1027, 2009.
- GOTTELAND M, JIMENEZ I, BRUNSER O, GUZMAN L, ROMERO S, CASSELS BK, SPEISKY H: Protective effect of boldine in experimental colitis. *Planta Med* **63**: 311-315, 1997.
- HERNANDEZ-SALINAS R, VIELMA AZ, ARISMENDI MN, BORIC MP, SAEZ JC, VELARDE V: Boldine prevents renal alterations in diabetic rats. *J Diabetes Res* **2013**: 593672, 2013.
- HROCH M, MICUDA S, CERMANOVA J, CHLADEK J, TOMSIK P: Development of an HPLC fluorescence method for determination of boldine in plasma, bile and urine of rats and identification of its major metabolites by LC-MS/MS. *J Chromatogr B Analyt Technol Biomed Life Sci* **936**: 48-56, 2013.
- JIMENEZ I, SPEISKY H: Biological disposition of boldine: in vitro and in vivo studies. *Phytother Res* **14**: 254-260, 2000.
- JOHNSON BM, ZHANG P, SCHUETZ JD, BROUWER KL: Characterization of transport protein expression in multidrug resistance-associated protein (Mrp) 2-deficient rats. *Drug Metab Dispos* **34**: 556-562, 2006.
- KADOVA Z, DOLEZELOVA E, CERMANOVA J, HROCH M, LAHO T, MUCHOVA L, STAUD F, VITEK L, MOKRY J, CHLADEK J, HAVLINOVA Z, HOLECEK M, MICUDA S: IL-1 receptor blockade alleviates endotoxin-mediated impairment of renal drug excretory functions in rats. *Am J Physiol Renal Physiol* **308**: F388-F399, 2015.
- LAHO T, CLARKE JD, DZIERLENGA AL, LI H, KLEIN DM, GOEDKEN M, MICUDA S, CHERRINGTON NJ: Effect of nonalcoholic steatohepatitis on renal filtration and secretion of adefovir. *Biochem Pharmacol* **115**: 144-151, 2016.
- LANHERS MC, JOYEUX M, SOULIMANI R, FLEURENTIN J, SAYAG M, MORTIER F, YOUNOS C, PELT JM: Hepatoprotective and anti-inflammatory effects of a traditional medicinal plant of Chile, *Peumus boldus*. *Planta Med* **57**: 110-115, 1991.
- MUTHNA D, CMIELOVA J, TOMSIK P, REZACOVA M: Boldine and related aporphines: from antioxidant to antiproliferative properties. *Nat Prod Commun* **8**: 1797-1800, 2013.
- NIES AT, KEPPLER D: The apical conjugate efflux pump ABCC2 (MRP2). *Pflugers Arch* **453**: 643-659, 2007.
- NOBILIS M, HOLCAPEK M, KOLAROVA L, KOPECKY J, KUNES M, SVOBODA Z, KVETINA J: Identification and determination of phase II nabumetone metabolites by high-performance liquid chromatography with photodiode array and mass spectrometric detection. *J Chromatogr A* **1031**: 229-236, 2004.
- O'BRIEN P, CARRASCO-POZO C, SPEISKY H: Boldine and its antioxidant or health-promoting properties. *Chem Biol Interact* **159**: 1-17, 2006.



- 
- SANTANAM N, PENUMETCHA M, SPEISKY H, PARTHASARATHY S: A novel alkaloid antioxidant, Boldine and synthetic antioxidant, reduced form of RU486, inhibit the oxidation of LDL in-vitro and atherosclerosis in vivo in LDLR(-/-) mice. *Atherosclerosis* **173**: 203-210, 2004.
- TOMSIK P, MICUDA S, MUTHNA D, CERMAKOVA E, HAVELEK R, RUDOLF E, HROCH M, KADOVA Z, REZACOVA M, CMIELOVA J, ZIVNY P: Boldine inhibits mouse mammary carcinoma in vivo and human MCF-7 breast cancer cells in vitro. *Planta Med* **82**: 1416-1424, 2016.
- ZAGOROVA M, PRASNICKA A, KADOVA Z, DOLEZELOVA E, KAZDOVA L, CERMANOVA J, ROZKYDALOVA L, HROCH M, MOKRY J, MICUDA S: Boldine attenuates cholestasis associated with nonalcoholic fatty liver disease in hereditary hypertriglyceridemic rats fed by high-sucrose diet. *Physiol Res* **64** (Suppl. 4): S467-S476, 2015.
- ZAMEK-GLISZCZYNSKI MJ, BEDWELL DW, BAO JQ, HIGGINS JW: Characterization of SAGE Mdr1a (P-gp), Bcrp, and Mrp2 knockout rats using loperamide, paclitaxel, sulfasalazine, and carboxydichlorofluorescein pharmacokinetics. *Drug Metab Dispos* **40**: 1825-1833, 2012.
- ZHOU SF, WANG LL, DI YM, XUE CC, DUAN W, LI CG, LI Y: Substrates and inhibitors of human multidrug resistance associated proteins and the implications in drug development. *Curr Med Chem* **15**: 1981-2039, 2008.
-