

REVIEW

VIP/PACAP Signaling as an Alternative Target During Hyperoxic Exposure in Preterm Newborns

Qëndrim THAÇI^{1,3}, Shkëlzen REÇICA¹, Islam KRYEZIU¹, Vadim MITROKHIN², Andre KAMKIN², Ramadan SOPI¹, Nikola HADZI-PETRUSHEV², Mitko MLADENOV^{2,3}

¹Department of Premedical Courses, Faculty of Medicine, University of Prishtina, Prishtina, Kosovo, ²Department of Fundamental and Applied Physiology, Russian National Research Medical University, Moscow, Russia, ³Institute of Biology, Faculty of Natural Science and Mathematics, Ss Cyril and Methodius University, Skopje, North Macedonia

Received January 5, 2021

Accepted March 25, 2021

Epub Ahead of Print June 2, 2021

Summary

The use of oxygen therapy (high doses of oxygen - hyperoxia) in the treatment of premature infants results in their survival. However, it also results in a high incidence of chronic lung disease known as bronchopulmonary dysplasia, a disease in which airway hyper-responsiveness and pulmonary hypertension are well known as consequences. In our previous studies, we have shown that hyperoxia causes airway hyper-reactivity, characterized by an increased constrictive and impaired airway smooth muscle relaxation due to a reduced release of relaxant molecules such as nitric oxide, measured under *in vivo* and *in vitro* conditions (extra- and intrapulmonary) airways. In addition, the relaxation pathway of the vasoactive intestinal peptide (VIP) and/or pituitary adenylate cyclase activating peptide (PACAP) is another part of this system that plays an important role in the airway caliber. Peptide, which activates VIP cyclase and pituitary adenylate cyclase, has prolonged airway smooth muscle activity. It has long been known that VIP inhibits airway smooth muscle cell proliferation in a mouse model of asthma, but there is no data about its role in the regulation of airway and tracheal smooth muscle contractility during hyperoxic exposure of preterm newborns.

Key words

Lung • Bronchopulmonary dysplasia • Hyperoxia • Vasoactive intestinal peptide • Pituitary adenylate cyclase-activating polypeptide • Preterm newborns

Corresponding authors

Q. Thaçi, Department of Biology, Faculty of Medicine, University of Prishtina, Mother Teresa 5, Prishtina 10000, Kosovo, E-mail: qendrimthaqi214@hotmail.com and M. Mladenov, Institute of Biology, Faculty of Natural Science and Mathematics, Ss Cyril and Methodius University, 1000, Skopje, North Macedonia, e-mail: m.mitko@gmail.com

Bronchopulmonary dysplasia

Bronchopulmonary dysplasia (BPD) was first described in 1967 by Northway *et al.* as a chronic lung disease in premature infants, (usually those treated with high oxygen partial pressure), because their alveoli are not enough developed to perform respiration (Jobe and Bancalari 2001, Zaban 2019). Airway hyper-responsiveness and pulmonary hypertension (PH) are well known consequences of BPD (Hershenson *et al.* 1994). Previous studies have shown that hyperoxia (treatment with high doses of oxygen) causes airway smooth muscle (ASM) hyperreactivity due to the reduced release of relaxant molecules such as nitric oxide (NO), changes in prostaglandin E₂ (PGE₂) levels, etc (Sopi *et al.* 2012, Stamenkovska *et al.* 2020). The data published through last few decades indicates that hyperreactivity involves many different molecular signaling mechanisms, among which the non-adrenergic-noncholinergic inhibitory system (iNANC) (Anaid *et al.* 2007), is one of

the mainly affected systems. Vasoactive intestinal peptide/pituitary adenylate cyclase-activating polypeptide (VIP/PACAP) relaxation pathway, is considered to be another part of this system and plays an important role in the airway caliber (Ao *et al.* 2011).

The hyperoxic exposure leads to generation of reactive oxygen species (ROS) in the lungs, such as superoxide radical anion ($O_2^{\cdot-}$), peroxy radicals (ROO^{\cdot}), and hydroxyl radical (HO^{\cdot}). The non-radical derivatives of molecular oxygen (O_2), like hydrogen peroxide (H_2O_2), hypochlorous acid (HOCl), singlet oxygen (1O_2), and peroxynitrite (ONOO $^{\cdot}$), are all strongly associated with the pathophysiology of BPD (Berkelhamer *et al.* 2013). Another major risk factor for developing BPD is pneumonia, which occurs when pro-inflammatory cytokines such as tumor necrosis factor alpha (TNF- α), interleukin 1 beta (IL-1 β), interleukin 6 (IL-6), receptor of chemokine 2 (CXCR2), and interleukin 11 (IL-11), are released in response to prenatal and neonatal trigger factors such as mechanical ventilation (Federico *et al.* 2007). Many of these pro-inflammatory cytokines have been detected in aspirated fluids of neonates with BPD (Bose *et al.* 2008).

VIP/PACAP (Vasoactive Intestinal Peptide/Pituitary Adenylate Cyclase-Activating Polypeptide)

The vasoactive intestinal peptide (VIP), also known as the vasoactive intestinal polypeptide, is a 28-amino acid peptide first isolated from the upper intestine in 1975 by Said and later found in many mammalian organs and tissues including the intestines (Costa and Furness 1983), lungs (Dey *et al.* 1981),

kidneys (Barajas *et al.* 1983), heart (Weihe and Remecke 1981), skin (Bloom and Polak, 1983), pancreas, suprachiasmatic nuclei of the hypothalamus, and widely distributed in the central and peripheral nervous systems (Said 1986), with approximately two minutes of blood half-life (Henning and Sawmiller 2001). The human VIP gene located in the chromosome 6q24 contains 7 introns and 6 exons, of which 5 are encoded (Hahn and Eidem 1998), whereas this gene in the rat is located in the chromosome 1p11 (Lamperti *et al.* 1991). VIP belongs to the super-family of structurally related peptide hormones which includes glucagon, glucagon-like peptide (GLP), helodermin, secretin, gastric inhibitory polypeptide (GIP), growth hormone releasing factor (GRF), and ligand II protein-receptors (Umetsu *et al.* 2011). VIPs may also contain sequences, encoding several additional biological neuroendocrine peptides, including the peptide histidine isoleucine [PHI, in low mammals] (Tatemoto and Mutt 1981), peptide histidine methionine [PHM], the human equivalent of PHI (Itoh 1983), histidine valine peptide [PHV] and C-terminal extended form of the PHI and PHM (Yiangou 1987). PHI, PHM, and PHV presumably perform their biological function through the same receptors as VIP (Fahrenkrug 1993).

The pituitary adenylate cyclase (AC), activating polypeptide (PACAP) was firstly isolated from ovine adrenaline secretion, insulin secretion, and immunosuppression (Arimura and Shioda 1995, Ghatei *et al.* 1993). Its half-life in human blood ranges between 5 and 10 min (Mentlein 1999). PACAP in humans is encoded by the *ADCYAP1* gene and is located in the chromosome 18p11 (Hosoya *et al.* 1992). Two types of this peptide have been identified to date: 38 amino acid peptides (PACAP-38) isolated from the sheep

Table 1. The amino acid sequences of VIP/PACAP and its related peptides

Peptide	Amino acid sequence								
	5	10	15	20	25	30	35	40	45
<i>VIP</i>	HSDAV	FTDNY	TRIRK	QMAVK	KYLNS	ILN			
<i>PACAP38</i>	HSDGI	FTDSY	SRYRK	QMAVK	KYLAA	VLGKR	YKQRV	KNK	
<i>PACAP27</i>	HSDGI	FTDSY	SRYRK	QMAVK	KYLAA	VL			
<i>PHI</i>	HADGV	FTSDF	SRLLG	QLSAL	KYLES	LI			
<i>Helodermin</i>	HSDAI	FTOY	SKLLA	KLALO	KYLAS	ILGSR	TSPPP		
<i>Secretin</i>	HSDGT	FTSEL	SRLRD	SARLQ	RLLQG	LV			
<i>Glucagon</i>	HSQGT	FTSDY	SKYLD	SRRAQ	DFVQV	LMNT			
<i>GRF</i>	YADAI	FTNSY	RKVLG	QLSAR	KLLQD	YMSRQ	QGESN	QERGA	RARL
<i>GIP</i>	YADGT	TFISD	YSIAM	DKIHO	ODFVN	WLLAO	KGKKN	DVKHN	ITO

hypothalamus that stimulates AC in rat anterior pituitary cells in culture (Miyata *et al.* 1989) and 27 amino acid peptide (PACAP-27), isolated from the same source (Miyata *et al.* 1990) (Table 1).

VIP/PACAP receptors in the airways

The biological effects of VIP and PACAP are mediated by three types of G-protein-coupled receptors (GPCR), VPAC1, VPAC2 and PAC1. VPAC1 and VPAC2 receptors are binding sites for both VIP and PACAP, while PAC1 is a binding site for PACAP only (Laburthe *et al.* 2002, Ito *et al.* 2001) (Fig. 1). The G protein receptor family is classified into 3 groups (A, B and C), generally as 7-pass trans-membrane protein receptors. The VIP/PACAP receptor belongs to group B from the GPCR family, which consists of 437-459 amino acid residues (Ulrich *et al.* 1998). VPAC1 was the first VIP and PACAP receptor isolated from rat lungs by (Ishihara *et al.* 1992). VPAC1 is also found in the central nervous system (CNS), predominantly in the cerebral cortex and hippocampus (Ishihara *et al.* 1992, Usdin *et al.* 1994), in peripheral tissues including the liver, lungs, intestines (Usdin *et al.* 1994, Sreedharan *et al.* 1995), as well as in T lymphocytes (Delgado *et al.* 1996). VPAC2 is the second receptor to respond to VIP and PACAP, cloned by Harmar and coworkers (1995), from a rat's odor bulb and later confirmed by (Usdin *et al.* 1994). Messenger RNA encoding the VPAC2 receptor is also found in the central nervous system (CNS), and most commonly in the thalamus and supra chiasmatic nucleus, as well as in the lower parts like hippocampus, brainstem, spinal cord, and dorsal root ganglia (Ito *et al.* 2001). The receptor is also present in many peripheral tissues, including the smooth muscles of the cardiovascular, gastrointestinal, and reproductive system (Adamou *et al.* 1995, Wei and Mojsov 1996). The PAC1 receptor for the first time was cloned by Pisegna and Wank in 1993, from the acinar pancreatic carcinoma cell line (AR4-2J) in rats, with a much greater ability to bind to PACAP-27 and PACAP-38 in comparison to VIP. The DNA sequences of the related mouse (Hashimoto *et al.* 1996a), bovine (Miyamoto *et al.* 1994), human (Ogi *et al.* 1993) and a series of rat receptors were published independently by several groups of authors (Hashimoto *et al.* 1993, Svoboda *et al.* 1993). PAC1 is highly expressed in the CNS, in the olfactory bulb, thalamus, hypothalamus, hippocampus, granular cells of the cerebellum [Hashimoto *et al.* 1996b, Shioda *et al.* 1997] and in a number of peripheral tissues, most commonly in the adrenal medulla (Moller *et al.* 1996) (Fig. 1).

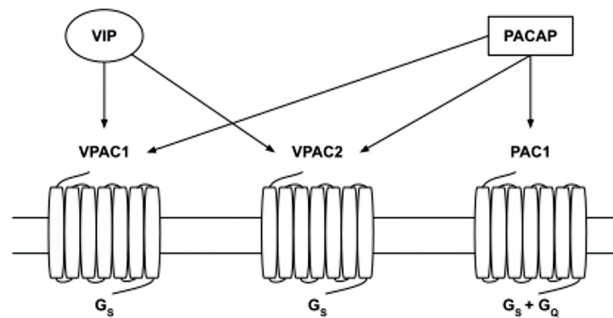


Fig. 1. Schematic representation of the signal transduction pathways of vasoactive intestinal peptide (VIP) / pituitary adenylate cyclase activating polypeptide (PACAP) receptors. Three receptors to PACAP have been described: VPAC1, VPAC2 and PAC1. VIP and PACAP show similar affinity for VPAC1 and VPAC2, whereas PACAP is more selective for PAC1 receptor.

VIP/PACAP signaling pathway in the airways

High-density VIP and PACAP expressing nerve fibers are found in the tracheobronchial tree, especially in the smooth muscle layer around submucosal, mucosal and serous glands, in the lamina propria, and the walls of pulmonary and bronchial arteries (Dey *et al.* 1981). As mentioned before the physiological effects of VIP and PACAP are mediated by three types of G-protein-coupled receptors VPAC1, VPAC2, and PAC1. These physiological actions include relaxation of the airways smooth muscle, bronchodilation (Diamond *et al.* 1983, Kanazawa *et al.* 1996), and pulmonary vasodilation (Linden *et al.* 1999). In different *in vivo* and *in vitro* studies, with various subjects including guinea pigs, rabbits, dogs and humans, VIP was shown to cause a reduction of the constrictive effects of histamine, prostaglandin $F_{2\alpha}$, kallikrein, leukotriene D_4 , neurokinins A and B and endothelin in isolated tracheal or bronchial segments (Hamasaki *et al.* 1983, Boomsma *et al.* 1990). On the other hand, calcium (Ca^{2+}) ions as an important player in the mechanisms of the muscle contraction/relaxation processes, may be released by the sarcoplasmic reticulum (SR), or transported from extracellular space (Groneberg *et al.* 2001, Kuo *et al.* 2003). After Ca^{2+} binding to the calmodulin, the myosin light chain kinase (MLCK) activates-(phosphorylate) myosin light chains (MLC), and allows the myosin cross-bridge to bind to the actin filaments, leading to contraction (Roux *et al.* 1997). In relation to VIP/PACAP, it was found that after their binding to corresponding receptors, they causes activation of the membrane-bound AC, which further generates cyclic adenosine monophosphate (cAMP) from adenosine triphosphate (ATP) (Robinson and Colbran 2013, Ganz

et al. 1986). The intracellular accumulation of cAMP on the level of the airways causes activation of a group of cAMP-dependent protein kinases A (PKA) (Francis *et al.* 1988, Hedlund *et al.* 1995). PKA phosphorylates phospholamban (PLN), a protein that normally interferes with the Ca^{2+} pump within the membrane of the SR. Reducing the level of free cytoplasmic Ca^{2+} or increasing Ca^{2+} uptake by internal stores like SR or mitochondria, results with smooth muscle relaxation (Mueller *et al.* 1979, Somlyo and Somlyo 1994). However, it is important to note that Ca^{2+} uptake by mitochondria is not cAMP regulated (Borie 1981). Other previous studies in rats, guinea pigs and humans, suggests that cAMP induces relaxation of ASM by interacting with various signaling pathways, including K^+ channels, more likely by membrane hyperpolarisation followed by a reduction in the Ca^{2+} influx via *voltage-dependent Ca^{2+} channels* (Nuttall and Farley 1996, Prakash *et al.* 1997). In addition, there is evidence that the reduction in the intracellular Na^+ by the Na^+/K^+ ATPase, caused increased Ca^{2+} efflux via $\text{Na}^+/\text{Ca}^{2+}$ exchanger, (the exchanger could be activated by PKA or directly by cAMP). The interaction of these channels would therefore be expected to induce ASM relaxation (Hall 2000, McGrogan *et al.* 1995, Gunst and Strop 1988). Additional mechanisms may contribute to the decreasing in the intracellular Ca^{2+} concentration, like inositol 1,4,5-triphosphate (IP_3)-gated Ca^{2+} release channels in the membrane of SR. IP_3 plays a substantial role in the opening of these channels, and different studies suggest for PKA prevented formation of the intracellular IP_3 , consequently followed by a reduced concentration of the intracellular Ca^{2+} (Yang *et al.* 1996, Ding *et al.* 1997). Moreover, activated PKA usually causes MLCK inactivation and reduces its ability to activate the MLCs, which is essential for ASM contraction, and bronchodilatation (Giembicz and Newton 2006).

In addition, VIP is degraded by proteases that are present at/or near the airway mucosa, including mast-cell tryptase and chymase and by neutral endopeptidase ("enkephalinase") (Caughey *et al.* 1988, Goetzl *et al.* 1989), whereas, PACAP is metabolized by dipeptidyl peptidase IV (Li *et al.* 2007).

Involvement of the VIP/PACAP signalling in the inflammation

As indicated before, another major risk factor for the development of BPD is inflammation. Particular types of pro-inflammatory cytokines and chemokines such as

$\text{TNF}\alpha$, IL-1 β , IL-6, chemokine receptor 2 (CXCR2) and CXCL8, IL-11 and IL-12 are related to inflammation. Numerous studies, in animal and human models, showed that VIP/PACAP signaling plays a key role in the balance between pro- and anti-inflammatory factors and possesses essential role in the successful control of inflammation (Gomariz *et al.* 2006, Ambalavanan *et al.* 2009). Transcription of the nuclear factor κB (NF- κB), leads to increased production of TNF- α , IL-1 β and IL-6. VIP/PACAP on the other hand is able to inhibit NF- κB translocation through a cAMP independent mechanism, further stimulating production of anti-inflammatory cytokines, such as IL-10, IL-11 and transforming growth factor- β (TGF- β), and at the same time prevent inflammation (Delgado *et al.* 1998, Trepicchio *et al.* 1996, Tsunawaki *et al.* 1988, Delgado *et al.* 1999). The VIP/PACAP cause inhibition of the production of pro-inflammatory cytokines mainly by involvement of the VPAC1-receptor, and lesser involvement of the VPAC2-receptor too (Delgado and Genea 1999, Di Benedetto *et al.* 2019). The main producers of cytokines are macrophages (Laskin and Pendino 1995, Juarranz *et al.* 2004). Moreover, VIP/PACAP was found to modulate inflammatory responses by regulation of the different functions in other cells, including the mast cells, microglia, dendritic cells and synovial fibroblasts (Tuncel *et al.* 2000, Abad *et al.* 2003). VIP also reduces the pro-inflammatory T helper1 (Th1) and T helper 17 (Th17) responses (Delgado *et al.* 2001, Abad *et al.* 2011, Benitez *et al.* 2018, Austin and Loyd 2014).

Involvement of the VIP/PACAP signaling in the pulmonary hypertension

Another well-known consequence of BPD is pulmonary hypertension (PH), which pathobiology is not yet completely clear. PH represents high blood pressure in the arteries of the lungs, which occurs when blood vessels in the lungs are narrowed, blocked or destroyed, and as a consequence blood flow through the lungs slows (Lau *et al.* 2017, Maarman *et al.* 2017). Other major determinants in the prognosis of the PH, are pulmonary artery pressure greater than 25 mmHg and right ventricular hypertrophy (Maarman *et al.* 2017). Several abnormal signaling pathways related to the PH have been identified, including reduced synthesis of prostacyclin and nitric oxide, and increased production of thromboxane and endothelin-1 (Giaida and Saleh 1995, Petkov *et al.* 2003). The recent studies have focused on the possible implication of the VIP/PACAP system in

patients with PH. A low level of VIP in the lungs is found in patients suffering from PH with an over-expression of both types VPAC receptors. Conversely, Said *et al.* (2007), have shown that VIP inhalation improves hemodynamics and lung capacity in the patients suffering from PH, proposing the peptide as a potential new treatment for PH. Previous observations in mice suggested that genetic knockout of the VIP gene, led to hemodynamic and histomorphological features of arterial PH, whereas intraperitoneal injections of VIP, has been shown to improve vascular pulmonary and right ventricular remodeling (Busto *et al.* 2000). Same as in other organs and tissues, the effect of VIP/PACAP in human pulmonary artery smooth muscle cells is mediated by VIP receptors VPAC1, VPAC2 and PAC1, which are primarily Gas-coupled receptors (Said *et al.* 2007). The VPAC2 receptor is highly expressed in human pulmonary artery smooth muscle cells (Said *et al.* 2007). Gas-coupled receptor activation causes an increase in cAMP, by activating AC, which can increase the activity of downstream mediators such as PKA, or induce expression of the protein directly activated by cAMP. PKA also phosphorylates targets such as MLCK to decrease its activity, resulting with vasodilatation and decreased proliferation of pulmonary artery smooth muscle cells (Fig. 2).

Conclusion

This review describes the physiological importance of VIP and PACAP in pulmonary diseases including BPD and PH. VIP/PACAP expresses a variety

References

- ABAD C, WASCHEK JA. Immunomodulatory roles of VIP and PACAP in models of multiple sclerosis. *Curr Pharm Des* 17: 1025-1035, 2011. <https://doi.org/10.2174/138161211795589364>
- ABAD C, MARTINEZ C, JUARRANZ MG, ARRANZ A, LECETA J, DELGADO M, GOMARIZ RP: Therapeutic effects of vasoactive intestinal peptide in the trinitrobenzene sulfonic acid mice model of Crohn's disease. *Gastroenterology* 124: 961-971, 2003. <https://doi.org/10.1053/gast.2003.50141>
- ADAMOUE JE, AIYAR N, VAN HORN S, ELSHOUBAGY NA: Cloning and functional characterization of the human vasoactive intestinal peptide (VIP)-2 receptor. *Biochem Biophys Res Commun* 209: 385-392, 1995. <https://doi.org/10.1006/bbrc.1995.1515>
- AMBALAVANAN N, CARLO WA, D'ANGIO CT, McDONALD SA, DAS A, SSHENDEL D, THORSEN P, HIGGINS RD; KENNEDY ESH: river National Institute of Child Health and Human Development, Neonatal Research Network. Cytokines associated with bronchopulmonary dysplasia or death in extremely low birth weight infants. *Pediatrics* 123: 1132-1141, 2009. <https://doi.org/10.1542/peds.2008-0526>

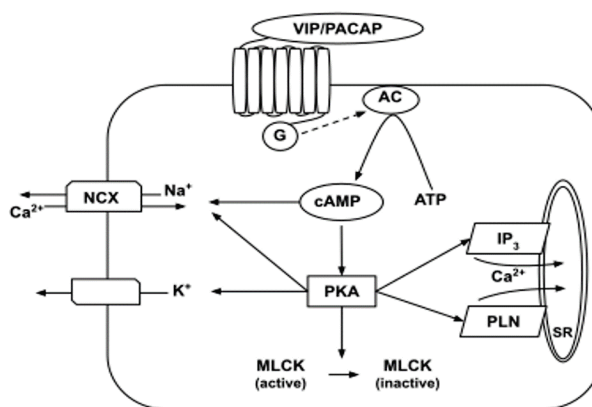


Fig. 2. Molecular actions of VIP/PACAP in induction of relaxation in airway smooth muscle cells. AC- adenylyl cyclase; cAMP - cyclic adenosine monophosphate; ATP - adenosine triphosphate; PKA - protein kinases A; IP₃ -inositol 1,4,5-triphosphate; PLN - phospholamban; MLCK - myosin light chain kinase.

of actions, including potent dilatory actions in the pulmonary blood vessels and ASM and a potent anti-inflammatory and anti-proliferative actions. Based on all mentioned above, our opinion is that VIP/PACAP signaling might have an important role in the regulation of airway and tracheal smooth muscle contractility during hyperoxic exposure of preterm newborns. The need for additional investigation may be suggested, that will lead VIP/PACAP or some other player from their airway/tracheal signaling to be classified as a medication in the potential treatment of BPD and PH.

Conflict of Interest

There is no conflict of interest.

- ANAID S, PETKOV V, BAYKUSCHEVA-GENTSHEVA T, HOEGER H, PAINSIPP E, HOLZER P, MOSGOELLER W: Involvement of endothelial NO in the dilator effect of VIP on rat isolated pulmonary artery. *Regul Pept* 139: 102-108, 2007. <https://doi.org/10.1016/j.regpep.2006.10.012>
- AO X, FANG F, XU F: Role of vasoactive intestinal peptide in hyperoxia-induced injury of primary type II alveolar epithelial cells. *Indian J Pediatr* 78: 535-539, 2011. <https://doi.org/10.1007/s12098-010-0248-1>
- ARIMURA A, SHIODA S: Pituitary adenylate cyclase activating polypeptide (PACAP) and its receptors: neuroendocrine and endocrine interaction. *Front Neuroendocrinol* 16: 53-88, 1995. <https://doi.org/10.1006/frne.1995.1003>
- AUSTIN ED, LOYD JE. The genetics of pulmonary arterial hypertension. *Circ Res* 115: 189-202, 2014. <https://doi.org/10.1161/CIRCRESAHA.115.303404>
- BARAJAS L, SOKOLSKI DN, LECHAGO J: Vasoactive intestinal polypeptide-immunoreactive nerves in the kidney. *Neurosci Lett* 43: 263-269, 1983. [https://doi.org/10.1016/0304-3940\(83\)90199-4](https://doi.org/10.1016/0304-3940(83)90199-4)
- BENITEZ R, DELGADO-MAROTO V, CARO M, FORTE-LAGO I, DURAN-PRADO M, O'VALLE F, LICHTMAN AH, GONZALEZ-REY E, DELGADO M: Vasoactive intestinal peptide ameliorates acute myocarditis and atherosclerosis by regulating inflammatory and autoimmune responses. *J Immunol* 200: 3697-3710, 2018. <https://doi.org/10.4049/jimmunol.1800122>
- BERKELHAMER SK, KIM GA, RADDER JE, WEDGWOOD S, CZECH L, STEINHORN RH, SCHUMACKER PT: Developmental differences in hyperoxia-induced oxidative stress and cellular responses in the murine lung. *Free Radic Bio Med* 61: 51-60, 2013. <https://doi.org/10.1016/j.freeradbiomed.2013.03.003>
- BLOOM SR, POLAK JM: Regulatory peptides and the skin. *Clin Exp Dermatol* 8: 3-18, 1983. <https://doi.org/10.1111/j.1365-2230.1983.tb01738.x>
- BOOMSMA JD, FODA HD, SAID SI: Vasoactive intestinal peptide (VIP) reverses endothelin-induced contractions of guinea pig trachea and pulmonary artery. *Am Rev Respir Dis* 141: A485, 1990.
- BORIE AB. Control, modulation and regulation of cell calcium. *Rev Physiol Biochem Pharmacol* 90: 13-153, 1981. <https://doi.org/10.1007/BFb0034078>
- BOSE CL, DAMMANN CEL, LAUGHON MM: Bronchopulmonary dysplasia and inflammatory biomarkers in the premature neonate. *Arch Dis Child Fetal Neonatal* 93: 455-461, 2008. <https://doi.org/10.1136/adc.2007.121327>
- BUSTO R, PRIETO JC, BODEGA G, ZAPATERO J, CARRERO I: Immunohistochemical localization and distribution of VIP/PACAP receptors in human lung. *Peptides* 21: 265-269, 2000. [https://doi.org/10.1016/S0196-9781\(99\)00202-8](https://doi.org/10.1016/S0196-9781(99)00202-8)
- CAUGHEY GH, LEIDIG F, VIRO NF, NADEL JA: Substance P and vasoactive intestinal peptide degradation by mast cell tryptase and chymase. *J Pharmacol Exp Ther* 244: 133-137, 1988.
- COSTA M, FURNESS JB. The origins, pathways and terminations of neurons with VIP-like immunoreactivity in the guinea-pig small intestine. *Neuroscience* 8: 665-676, 1983. [https://doi.org/10.1016/0306-4522\(83\)90002-7](https://doi.org/10.1016/0306-4522(83)90002-7)
- DELGADO M, ABAD C, MARTINEZ C, LECETA J, GOMARIZ RP: Vasoactive intestinal peptide prevents experimental arthritis by downregulating both autoimmune and inflammatory components of the disease. *Nat Med* 7: 563-568, 2001. <https://doi.org/10.1038/87887>
- DELGADO M, MUNOZ-ELIAS EJ, GOMARIZ RP, GANEA D: Vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide enhance IL-10 production by murine macrophages: in vitro and in vivo studies. *J Immunol* 162: 1707-1716, 1999.
- DELGADO M, GANEA D: Vasoactive intestinal peptide and pituitary adenylate cyclase activating polypeptide inhibit IL-12 transcription by regulating NF- κ B and Ets activation. *J Biol Chem* 274: 31930-31940, 1999. <https://doi.org/10.1074/jbc.274.45.31930>
- DELGADO M, MUNOZ-ELIAS EJ, KAN Y, GOZES I, FRIDKIN M, BRENNEMAN DE, GOMARIZ RP, GANEA D: Vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide inhibit tumor necrosis factor alpha transcriptional activation by regulating nuclear factor- κ B and cAMP response element-binding protein/c-Jun. *J Biol Chem* 273: 31427-31436, 1998. <https://doi.org/10.1074/jbc.273.47.31427>
- DELGADO M, MARTINEZ C, JOHNSON MC, GOMARIZ RP, GANEA D: Differential expression of vasoactive intestinal peptide receptors 1 and 2 (VIP-R1 and VIP-R2) mRNA in murine lymphocytes. *J Neuroimmunol* 68: 27-38, 1996. [https://doi.org/10.1016/0165-5728\(96\)00063-X](https://doi.org/10.1016/0165-5728(96)00063-X)

- DEY RD, SHANNON WA, STUD SI: Localization of VIP-immunoreactive nerves in airways and pulmonary vessels of dogs, cats and human subjects. *Cell Tissue Res* 220: 231-238, 1981. <https://doi.org/10.1007/BF00210505>
- DIAMOND L, SZAREK LJ, GILLESPIE MN, ALTIERE RJ: In vivo bronchodilator activity of vasoactive intestinal peptide in the cat. *Am Rev Respir Dis* 12: 827-832, 1983.
- DI BENEDETTO P, RUSCITTI P, VADAS Z, TOUBI E, GIACOMELLI R: Macrophages with regulatory functions, a possible new therapeutic perspective in autoimmune diseases. *Autoimmun Rev* 18: 102369, 2019. <https://doi.org/10.1016/j.autrev.2019.102369>
- DING KH, HUSAIN S, AKHTAR RA, ISALES CM, ABDEL-LATIF AA: Inhibition of muscarinic-stimulated polyphosphoinositide hydrolysis and Ca²⁺ mobilization in cat iris sphincter smooth muscle cells by cAMP-elevating agents. *Cell Signal* 9: 411-421, 1997. [https://doi.org/10.1016/S0898-6568\(97\)00018-1](https://doi.org/10.1016/S0898-6568(97)00018-1)
- FAHRENKRUG J. Transmitter role of vasoactive intestinal peptide. *Pharmacol Toxicol* 72: 354-363, 1993. <https://doi.org/10.1111/j.1600-0773.1993.tb01344.x>
- FARBER HW, LOSCALZO J. Pulmonary arterial hypertension. *N Engl J Med* 351: 1655-1665, 2004. <https://doi.org/10.1056/NEJMra035488>
- FEDERICO A, MORGILLO F, TUSSILLO C, CIARDIELLO F, LOGUERCIO C: Chronic inflammation and oxidative stress in human carcinogenesis. *Int J Cancer* 121: 2381-2386, 2007. <https://doi.org/10.1002/ijc.23192>
- FRANCIS SH, NOBLETT BD, TODD BW, WELLS JN, CORBIN JD: Relaxation of vascular and tracheal smooth muscle by cyclic nucleotide analogs that preferentially activate purified cGMP-dependent protein kinase. *Mol Pharmacol* 34: 506-517, 1988.
- GANZ P, SANDROCK AW, LANDIS SC, LEOPOLD J, Jr GIMBRONE MA, ALEXANDER RW: Vasoactive intestinal peptide: vasodilation and cyclic AMP generation. *Am J Physiol* 250: 755-760, 1986. <https://doi.org/10.1152/ajpheart.1986.250.5.H755>
- GHATEI MA, TAKAHASHI K, SUZUKI Y, GARDINER J, JONES PM, BLOOM SR: Distribution, molecular characterization of pituitary adenylate cyclase-activating polypeptide and its precursor encoding messenger RNA in human and rat tissues. *J Endocrinol* 136: 159-166, 1993. <https://doi.org/10.1677/joe.0.1360159>
- GIAID A, SALEH D: Reduced expression of endothelial nitric oxide synthase in the lungs of patients with pulmonary hypertension. *N Engl J Med* 333: 214-221, 1995. <https://doi.org/10.1056/NEJM199507273330403>
- GIEMBYCZ MA, NEWTON R: Beyond the dogma: novel beta2-adrenoceptor signalling in the airways. *Eur Respir J* 27: 1286-1306, 2006. <https://doi.org/10.1183/09031936.06.00112605>
- GOETZL EJ, SREEDHARAN SP, TRRCK CW, BRIDENBAUGH R, MALFROY B: Preferential cleavage of amino- and carboxylterminal oligopeptides from vasoactive intestinal polypeptide by human recombinant enkephalinase (neutral endopeptidase, EC 3.4.24.11). *Biochem Biophys Res Commun* 158: 850-854, 1989. [https://doi.org/10.1016/0006-291X\(89\)92800-3](https://doi.org/10.1016/0006-291X(89)92800-3)
- GOMARIZ RP, JUARRANZ Y, ABAD C, ARRANZ A, LECETA J, MARTINEZ C: VIP-PACAP system in immunity: New insights for multi target therapy. *Ann NY Acad Sci* 1070: 51-74, 2006. <https://doi.org/10.1196/annals.1317.031>
- GRONEBERG DA, SPRINGER J, FISCHER A: Vasoactive intestinal polypeptide as mediator of asthma. *Pulm Pharmacol Ther* 14: 391-401, 2001. <https://doi.org/10.1006/pupt.2001.0306>
- GUNST SS, STROPP JQ: Effect of Na⁺/K⁺ adenosine triphosphatase activity on relaxation of canine tracheal smooth muscle. *Appl Physiol* 64: 635-641, 1988. <https://doi.org/10.1152/jappl.1988.64.2.635>
- HAHM SH, EIDEN LE: Cis-regulatory elements controlling basal and inducible VIP gen transcription. *Ann NY Acad Sci* 865: 10-26, 1998. <https://doi.org/10.1111/j.1749-6632.1998.tb11158.x>
- HALL IP. Second messengers, ion channels and pharmacology of airway smooth muscle. *Eur Respir J* 15: 1120-1127, 2000. <https://doi.org/10.1034/j.1399-3003.2000.01523.x>
- HAMASAKI Y, MOJARAD M, SAID SI: Relaxant action of VIP on cat pulmonary artery: Comparison with acetylcholine, Isoproterenol & PGEI. *J Appl Physiol* 54: 1607-1611, 1983. <https://doi.org/10.1152/jappl.1983.54.6.1607>

- HASHIMOTO H, NOGI H, MORI K, OHISHI H, SHIGEMOTO R, YAMAMOTO K, MATSUDA T, MIZUNO N, NAGATA S, BABA A: Distribution of the mRNA for a pituitary adenylate cyclase-activating polypeptide receptor in the rat brain: an in situ hybridization study. *J Comp Neurol* 371: 567-577, 1996a. [https://doi.org/10.1002/\(SICI\)1096-9861\(19960805\)371:4<567::AID-CNE6>3.0.CO;2-2](https://doi.org/10.1002/(SICI)1096-9861(19960805)371:4<567::AID-CNE6>3.0.CO;2-2)
- HASHIMOTO H, YAMAMOTO K, HAGIGARA N, OGAWA N, NISHINO A, AINO H, NOGI, KIMANISHI H, MATSUDA T, BABA A: cDNA cloning of a mouse pituitary adenylate cyclase-activating polypeptide receptor. *Biochim Biophys Acta*. 1281: 129-133, 1996b. [https://doi.org/10.1016/0005-2736\(96\)00056-9](https://doi.org/10.1016/0005-2736(96)00056-9)
- HASHIMOTO H, ISHIHARA T, SHIGEMOTO R, MORI K, NAGATA S: Molecular cloning and tissue distribution of a receptor for pituitary adenylate cyclase-activating polypeptide. *Neuron* 11: 333-342, 1993. [https://doi.org/10.1016/0896-6273\(93\)90188-W](https://doi.org/10.1016/0896-6273(93)90188-W)
- HEDLUND P, ALM P, EKSTROM P, FAHRENKRUG J, HANNIBAL J, HEDLUND H, LARSSON B, ANDERSSON KE: Pituitary adenylate cyclase-activating polypeptide, helospectin, and vasoactive intestinal polypeptide in human corpus cavernosum. *Br J Pharmacol*. 116: 2258-2266, 1995. <https://doi.org/10.1111/j.1476-5381.1995.tb15062.x>
- HENNING RJ, SAWMILLER DR: Vasoactive intestinal peptide: cardiovascular effects. *Cardiovasc Res* 49: 27-37, 2001. [https://doi.org/10.1016/S0008-6363\(00\)00229-7](https://doi.org/10.1016/S0008-6363(00)00229-7)
- HERSHENSON MB, WYLAM ME, PUNJABI N, UMANS JG, SCHUMACKER PT, MITCHELL RW, SOLWAY J: Exposure of immature rats to hyperoxia increases tracheal smooth muscle stress generation in vitro. *J Appl Physiol* 76: 743-749, 1994. <https://doi.org/10.1152/jappl.1994.76.2.743>
- HOSOYA M, KIMURA C, OGI K, OHKUBO S, MIYAMOTO Y, KUGOH H, SHIMIZU M, ONDA H, OSHIMURA MARIMURA A, FUJINO M: Structure of the human pituitary adenylate cyclase activating polypeptide (PACAP) gene. *Biochim Biophys Acta* 1129: 199-206, 1992. [https://doi.org/10.1016/0167-4781\(92\)90488-L](https://doi.org/10.1016/0167-4781(92)90488-L)
- ISHIHARA T, SHIGEMOTO R, MORI K, TAKAHASHI K, NAGATA S: Functional expression and tissue distribution of a novel receptor for vasoactive intestinal polypeptide. *Neuron* 8: 811-819, 1992. [https://doi.org/10.1016/0896-6273\(92\)90101-I](https://doi.org/10.1016/0896-6273(92)90101-I)
- ITO T, IGARASHI H, PRADHAN TK, HOU W, MANTEY SA, TAYLOR JE, MURPHY WA, COY DH, JENSEN RT: GI side-effects of a possible therapeutic GRF analogue in monkeys are likely due to VIP receptor agonist activity. *Peptides* 22: 1139-1151, 2001. [https://doi.org/10.1016/S0196-9781\(01\)00436-3](https://doi.org/10.1016/S0196-9781(01)00436-3)
- ITOH N, OBATA K, YANAIHARA N, OKAMOTO H: Human preprovasoactive intestinal polypeptide contains a novel PHI-27-like peptide, PHM-27. *Nature* 304: 547-549, 1983. <https://doi.org/10.1038/304547a0>
- JOBE AH, BANCALARI E: Bronchopulmonary dysplasia. *Am J Respir Crit Care Med* 163: 1723-1729, 2001. <https://doi.org/10.1164/ajrccm.163.7.2011060>
- JUARRANZ MG, SANTIAGO B, TORROBA M, GUTIERREZ-CANAS I, PALAO G, GALINDO M, ABAD C, MARTINEZ C, LECETA J, PABLOS JL, GOMARIZ RP: Vasoactive intestinal peptide modulates proinflammatory mediator synthesis in osteoarthritic and rheumatoid synovial cells. *Rheumatology* 43: 416-422, 2004. <https://doi.org/10.1093/rheumatology/keh061>
- KANAZAWA H, KAWAGUCHI T, FUJII T, SHOJI S, HIRATA K, KUDOH S, KURIHARA N, YOSHIKAWA J: Potentiation of the bronchoprotective effects of vasoactive intestinal peptide, isoprenaline, and theophylline against histamine challenge in anaesthetized guinea pigs by adrenomedullin. *Thorax* 51: 1199-1202, 1996. <https://doi.org/10.1136/thx.51.12.1199>
- KUO KH, DAI J, SEOW CY, LEE CH, VAN BREEMEN C: Relationship between asynchronous Ca²⁺ waves and force development in intact smooth muscle bundles of the porcine trachea. *Am J Physiol Lung Cell Mol Physiol* 285: 1345-1353, 2003. <https://doi.org/10.1152/ajplung.00043.2003>
- LABURTHER M, COUVINEAU A, MARIE JC: VPAC receptors for VIP and PACAP. *Recept Channels* 8: 137-153, 2002. <https://doi.org/10.1080/10606820213680>
- LAMPERTI ED, ROSEN KM, VILLA-KOMAROFF L: Characterization of the gene and messages for vasoactive intestinal polypeptide (VIP) in rat and mouse. *Brain Res Mol Brain Res* 9: 217-231. 1991. [https://doi.org/10.1016/0169-328X\(91\)90005-I](https://doi.org/10.1016/0169-328X(91)90005-I)
- LASKIN DL, PENDINO KJ: Macrophages and inflammatory mediators in tissue injury. *Ann Rev Pharmacol Toxicol* 35: 655-677, 1995. <https://doi.org/10.1146/annurev.pa.35.040195.003255>

- LAU EMT, GIANNOULATOU E, CELERMAJER DS, HUMBERT M: Epidemiology and treatment of pulmonary arterial hypertension. *Nat Rev Cardiol* 14: 603-614, 2017. <https://doi.org/10.1038/nrcardio.2017.84>
- LINDEN A, CARDELL LO, YOSHIHARA S, NADELL JA: Bronchodilation by pituitary adenylate cyclase-activating peptide and related peptides. *Eur Respir J* 14: 443-451, 1999. <https://doi.org/10.1183/09031936.99.14244399>
- LI M, MADERDRUT JL, LERTORA JJ, BATUMAN V: Intravenous infusion of pituitary adenylate cyclase-activating polypeptide (PACAP) in a patient with multiple myeloma and myeloma kidney: a case study. *Peptides* 28: 1891-1895, 2007. <https://doi.org/10.1016/j.peptides.2007.05.002>
- MAARMAN GJ, SCHULZ R, SLIWA K, SCHERMULY RT, LECOUC S: Novel putative pharmacological therapies to protect the right ventricle in pulmonary hypertension: a review of current literature. *Br J Pharmacol* 174: 497-511, 2017. <https://doi.org/10.1111/bph.13721>
- MCGROGAN I, LU S, HIPWORTH S, SORMAZ L, ENG R, PREOCANIN D, DANIEL EE: Mechanisms of cyclic nucleotide-induced relaxation in canine tracheal smooth muscle. *Am J Physiol* 268: 407-413, 1995. <https://doi.org/10.1152/ajplung.1995.268.3.L407>
- MENTLEIN R. Dipeptidyl-peptidase IV (CD26)-role in the inactivation of regulatory peptides. *Regul Pept* 85: 9-24, 1999. [https://doi.org/10.1016/S0167-0115\(99\)00089-0](https://doi.org/10.1016/S0167-0115(99)00089-0)
- MIYAMOTO Y, HABATA Y, OHTAKI T, MASUDA Y, OGI K, ONDA H, FUJINO M: Cloning and expression of a complementary DNA encoding the bovine receptor for pituitary adenylate cyclase activating polypeptide (PACAP). *Biochim Biophys Acta* 1218: 297-307, 1994. [https://doi.org/10.1016/0167-4781\(94\)90181-3](https://doi.org/10.1016/0167-4781(94)90181-3)
- MIYATA A, JIANG L, DAHL RR, KITADA C, KUBO K, FUJINO M, MINAMINO N, ARIMURA A: Isolation of a neuropeptide corresponding to N-terminal 27 residues of pituitary adenylate cyclase activating polypeptide with 38 residues (PACAP38). *Biochem Biophys Res Commun* 170: 643-648, 1990. [https://doi.org/10.1016/0006-291X\(90\)92140-U](https://doi.org/10.1016/0006-291X(90)92140-U)
- MIYATA A, ARIMURA A, DAHL DH, MINAMINO N, UEHARA A, JIANG L, CULLER MD, COY DH: Isolation of a novel 38 residue hypothalamic polypeptide which stimulates adenylate cyclase in pituitary cells. *Biochem Biophys Res Commun* 164: 567-574, 1989. [https://doi.org/10.1016/0006-291X\(89\)91757-9](https://doi.org/10.1016/0006-291X(89)91757-9)
- MOLLER K, SUNDLER F: Expression of pituitary adenylate cyclase activating peptide (PACAP) and PACAP type I receptors in the rat adrenal medulla. *Regul Pept* 63: 129-139, 1996. [https://doi.org/10.1016/0167-0115\(96\)00033-X](https://doi.org/10.1016/0167-0115(96)00033-X)
- MUELLER E, VAN BREEMAN C: Role of intracellular Ca^{2+} sequestration in beta adrenergic relaxation of airway smooth muscle. *Nature* 281: 682-683, 1979. <https://doi.org/10.1038/281682a0>
- NORTHWAY Jr WH, ROSAN RC, PORTER DY: Pulmonary disease following respiratory therapy of hyaline-membrane disease. Bronchopulmonary dysplasia. *N Engl J Med* 276: 357-368, 1967. <https://doi.org/10.1056/NEJM196702162760701>
- NUTTLE LC, FARLEY JM: Frequency modulation of acetylcholine-induced oscillations in Ca^{2+} and Ca^{2+} -activated Cl^{-} current by cAMP in tracheal smooth muscle. *J Pharmacol Exp Ther* 277: 753-760, 1996.
- OGI K, MIYAMOTO Y, MASUDA Y, HABATA Y, HOSOYA M, OHTAKI T, MASUO Y, ONDA H, FUJINO M: Molecular cloning and functional expression of a cDNA encoding a human pituitary adenylate cyclase-activating polypeptide receptor. *Biochem Biophys Res Commun* 196: 1511-1521, 1993. <https://doi.org/10.1006/bbrc.1993.2423>
- PETKOV V, MOSGOELLER W, ZIESCHE R: Vasoactive intestinal peptide as new drug for treatment of primary pulmonary hypertension. *J Clin Invest* 111: 1339-1346, 2003. <https://doi.org/10.1172/JCI17500>
- PISEGNA JR, WANK SA: Molecular cloning and functional expression of the pituitary adenylate cyclase-activating polypeptide type I receptor. *Proc Natl Acad Sci USA*. 90: 6345-6349, 1993. <https://doi.org/10.1073/pnas.90.13.6345>
- PRAKASH YS, VAN DER HEIJDEN HF, KANNAN MS, SIECK GC: Effects of salbutamol on intracellular calcium oscillations in porcine airway smooth muscle. *J Appl Physiol* 82: 1836-1843, 1997. <https://doi.org/10.1152/jappl.1997.82.6.1836>
- ROBINSON A, COLBRAN R: Calcium/Calmodulin-Dependent Protein Kinases. *Encyclopedia of Biological Chemistry* 2: 304-309, 2013. <https://doi.org/10.1016/B978-0-12-378630-2.00493-X>

- ROUX E, GUIBERT C, SAVINEAU JP, MARTHAN R: [Ca²⁺]_i oscillations induced by muscarinic stimulation in airway smooth muscle cells: receptor subtypes and correlation with the mechanical activity. *Br J Pharmacol* 120: 1294-1301, 1997. <https://doi.org/10.1038/sj.bjp.0701061>
- SAID SI, HAMIDI SA, DICKMAN KG, SZEMA AM, LYUBSKY S, LIN RZ, JIANG YP, CHEN JJ, WASCHE JA, KORT S: Moderate pulmonary arterial hypertension in male mice lacking the vasoactive intestinal peptide gene. *Circulation* 115: 1260-1268, 2007. <https://doi.org/10.1161/CIRCULATIONAHA.106.681718>
- SAID SI: Vasoactive intestinal peptide. *J Endocrinol Invest* 9: 191-200, 1986. <https://doi.org/10.1007/BF03348097>
- SAID SI: Vasoactive intestinal polypeptide: widespread distribution in normal gastrointestinal organs. Proceedings of the 57th Annual Meeting of the Endocrine Society, New York. 1975.
- SHERWOOD NM, KRUECKI SL, McRORY JE: The origin and function of the pituitary adenylate cyclase-activating polypeptide (PACAP)/glucagon superfamily. *Endocr Rev* 21: 619-670, 2000. <https://doi.org/10.1210/edrv.21.6.0414>
- SHEWARD WJ, LUTZ EM, HARMAR AJ: The distribution of vasoactive intestinal peptide-2 receptor messenger RNA in the rat brain and pituitary gland as assessed by in situ hybridization. *Neuroscience* 67: 409-418, 1995. [https://doi.org/10.1016/0306-4522\(95\)00048-N](https://doi.org/10.1016/0306-4522(95)00048-N)
- SHIODA S, SHUTO Y, SOMOGYVARI-VIGH A, LEGRADI G, ONDA H, COY DH, NAKAJO S, ARIMURA A: Localization and gene expression of the receptor for pituitary adenylate cyclase-activating polypeptide in the rat brain. *Neurosci Res* 28: 345-354, 1997. [https://doi.org/10.1016/S0168-0102\(97\)00065-5](https://doi.org/10.1016/S0168-0102(97)00065-5)
- SOMLYO AP, SOMLYO AV: Signal transduction and regulation in smooth muscle. *Nature* 372: 231-236, 1994. <https://doi.org/10.1038/372231a0>
- SOPI RB, ZAIDI SIA, MLADENOV M, SAHITI H, ISTREFI Z, GJORGOSKI I, LAJCI A, JAKUPAJ M: L-citrulline supplementation reverses the impaired airway relaxation in neonatal rats exposed to hyperoxia. *Respir Res* 13: 68, 2012. <https://doi.org/10.1186/1465-9921-13-68>
- SREEDHARAN SP, HUANG JX, CHEUNG MC, GOETZL EJ: Structure, expression, and chromosomal localization of the type I human vasoactive intestinal peptide receptor gene. *Proc Natl Acad Sci USA* 92: 2939-2943, 1995. <https://doi.org/10.1073/pnas.92.7.2939>
- STAMENKOVSKA M, THACI Q, HADZIPETRUSHEV N, ANGELOVSKI M, BOGD-ANOV J, RECICA S, KRYEZIU I, GAGOV H, MITROKHIN V, KAMKIN A, SCHUBERT R, MLADENOV M, SOPI RB: Curcumin analogs (B2BrBC and C66) supplementation attenuates airway hyperreactivity and promote airway relaxation in neonatal rats exposed to hyperoxia. *Physiol Rep* 8: e14555, 2020. <https://doi.org/10.14814/phy2.14555>
- SVOBODA M, TASTENOY M, CICCARELLI E, STIEVENART M, CHRISTOPHE J: Cloning of a splice variant of the pituitary adenylate cyclase-activating polypeptide (PACAP) type I receptor. *Biochem Biophys Res Commun* 195: 881-888, 1993. <https://doi.org/10.1006/bbrc.1993.2127>
- TATEMOTO K, MUTT V: Isolation and characterization of the intestinal peptide porcine PHI (PHI-27), a new member of the glucagon-secretin family. *Proc Natl Acad Sci USA* 78: 6603-6607, 1981. <https://doi.org/10.1073/pnas.78.11.6603>
- TREPICCHIO W, BOZZA M, PEDNEAULT G, DORNER A: Recombinant human IL-11 attenuates the inflammatory response through down-regulation of proinflammatory cytokine release and nitric oxide production. *J Immunol* 157: 3627-3634, 1996.
- TSUNAWAKI S, SPORN M, DING A, NATHAN CF: Deactivation of macrophages by transforming growth factor. *Nature* 334: 260-262, 1988. <https://doi.org/10.1038/334260a0>
- TUNCEL N, TORE F, SAHINTURK V, AK D, TUNCEL M: Vasoactive intestinal peptide inhibits degranulation and changes granular content of mast cells: A potential therapeutic strategy in controlling septic shock. *Peptides* 21: 81-89, 2000. [https://doi.org/10.1016/S0196-9781\(99\)00177-1](https://doi.org/10.1016/S0196-9781(99)00177-1)
- ULRICH CD, HOLTMANN M, MILLER LJ: Secretin and vasoactive intestinal peptide receptors: members of a unique family of G protein coupled receptors. *Gastroenterology* 114: 382-397, 1998. [https://doi.org/10.1016/S0016-5085\(98\)70491-3](https://doi.org/10.1016/S0016-5085(98)70491-3)

-
- UMETSU Y, TENNO T, GODA N, SHIRAKAWA M, IKEGAMI T, HIROAKI H: Structural difference of vasoactive intestinal peptide in two distinct membrane-mimicking environments. *Biochim Biophys Acta* 1814: 724-730, 2011. <https://doi.org/10.1016/j.bbapap.2011.03.009>
- USDIN TB, BONNER TI, MEZEY E: Two receptors for vasoactive intestinal polypeptide with similar specificity and complementary distributions. *Endocrinology* 135: 2662-2680, 1994. <https://doi.org/10.1210/endo.135.6.7988457>
- WEI Y, MOJSOV S: Tissue specific expression of different human receptor types for pituitary adenylate cyclase activating polypeptide and vasoactive intestinal polypeptide: implications for their role in human physiology. *J Neuroendocrinol* 8: 811-817, 1996. <https://doi.org/10.1046/j.1365-2826.1996.05191.x>
- WEIHE E, REMECKE M: Peptidergic innervation of the mammalian sinus nodes: vasoactive intestinal polypeptide, neurotensin, substance P. *Neuro Sci Lett* 26: 283-288, 1981. [https://doi.org/10.1016/0304-3940\(81\)90146-4](https://doi.org/10.1016/0304-3940(81)90146-4)
- YANG CM, HSU MC, TSAO HL, CHIU CT, ONG R, HSIEH JT, FAN LW: Effect of cAMP elevating agents on carbachol-induced phosphoinositide hydrolysis and calcium mobilization in cultured canine tracheal smooth muscle cells. *Cell Calcium* 19: 243-254, 1996. [https://doi.org/10.1016/S0143-4160\(96\)90025-1](https://doi.org/10.1016/S0143-4160(96)90025-1)
- YIANGOU Y, DI MARZO V, SPOKES RA, PANICO M, MORRIS HR, BLOOM SR: Isolation, characterization, and pharmacological actions of peptide histidine valine 42, a novel prepro-vasoactive intestinal peptide-derived peptide. *J Biol Chem* 262: 14010-14013, 1987.
- ZOBAN P: Optimal oxygen saturation in extremely premature neonates. *Physiol Res* 68: 171-178, 2019. <https://doi.org/10.33549/physiolres.933987>
-